

## Research Article

# Herpesvirus-Mediated Delivery of a Genetically Encoded Fluorescent $\text{Ca}^{2+}$ Sensor to Canine Cardiomyocytes

János Prorok,<sup>1</sup> Péter P. Kovács,<sup>1</sup> Attila A. Kristóf,<sup>2</sup> Norbert Nagy,<sup>1</sup> Dóra Tombác,<sup>3</sup> Judit S. Tóth,<sup>3</sup> Balázs Ördög,<sup>3</sup> Norbert Jost,<sup>2</sup> László Virág,<sup>1</sup> Julius G. Papp,<sup>1,2</sup> András Varró,<sup>1</sup> András Tóth,<sup>1</sup> and Zsolt Boldogkői<sup>3</sup>

<sup>1</sup> Department of Pharmacology and Pharmacotherapy, Faculty of Medicine, University of Szeged, H-6720 Szeged, Hungary

<sup>2</sup> Division of Cardiovascular Pharmacology, Hungarian Academy of Sciences, H-6720 Szeged, Hungary

<sup>3</sup> Department of Medical Biology, Faculty of Medicine, University of Szeged, H-6720 Szeged, Hungary

Correspondence should be addressed to Zsolt Boldogkői, boldogkoi@yahoo.com

Received 27 February 2009; Accepted 4 May 2009

Recommended by Mohamed Boutjdir

We report the development and application of a pseudorabies virus-based system for delivery of troponin, a fluorescent  $\text{Ca}^{2+}$  sensor to adult canine cardiomyocytes. The efficacy of transduction was assessed by calculating the ratio of fluorescently labelled and nonlabelled cells in cell culture. Interaction of the virus vector with electrophysiological properties of cardiomyocytes was evaluated by the analysis of transient outward current ( $I_{to}$ ), kinetics of the intracellular  $\text{Ca}^{2+}$  transients, and cell shortening. Functionality of transferred troponin was verified by FRET analysis. We demonstrated that the transfer efficiency of troponin to cultured adult cardiac myocytes was virtually 100%. We showed that even after four days neither the amplitude nor the kinetics of the  $I_{to}$  current was significantly changed and no major shifts occurred in parameters of  $[\text{Ca}^{2+}]_i$  transients. Furthermore, we demonstrated that infection of cardiomyocytes with the virus did not affect the morphology, viability, and physiological attributes of cells.

Copyright © 2009 János Prorok et al. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

## 1. Introduction

The past decade has witnessed an explosive progress of virus-based gene delivery technologies. The reason for this is that, albeit traditional approaches, such as calcium phosphate precipitation, electroporation, or liposome-mediated gene transfer perform excellently in immortalized cells of various origins, they mostly fail in primary cultured cells and under in vivo conditions. Viruses had millions of years to evolve several means for effective infection of cells that can be employed by utilizing viruses as vectors for delivering exogenous genes to the desired cells. Virus-mediated gene transfer methods have also become powerful and widely used experimental tools in cardiovascular research. The most important prerequisites for successful in vitro gene transfer into adult cardiomyocytes include high quality of primary cell culture and an effective transfer vector with limited cytotoxicity and other side effects. However, adult cardiomyocytes are difficult to transduce using methods that work well

in other systems because cardiac cells do not divide, have a relatively short lifetime in culture, and are highly sensitive to toxic effects. Gene transfer methods used in cardiovascular research can be divided into two groups: physicochemical and viral vector-based systems. Nonviral methods involve cationic liposome/plasmid DNA complexes, incubation with naked DNA, and calcium phosphate precipitation [1, 2]. All of these methods suffer from severe limitations such as cytotoxicity, very low transfection efficiency especially in vivo applications, and short-term expression of transduced genes due to intracellular degradation of foreign DNA. The above limitations urged the application of more efficient virus vector-based approaches for gene delivery to the cardiovascular system. The following viral systems have been applied in molecular cardiology: adenovirus vectors (Ads), adeno-associated viruses (AAVs), retroviruses (RV) like lentivirus (HIV-1), and herpes simplex virus (HSV-1)-derived vectors [1, 2]. Currently, Ads and AAVs are the most frequently used tools for delivering genes into the

cells of the cardiovascular system, both in vivo and in vitro [3, 4]. While Ads-based vectors allow relatively highly efficient delivery of transgenes to cardiac cells, this system provides only transient expression of transferred genes since the virus does not integrate into the host genome [5]. The application of Ads have further limitations including strong immune responses by the host organism, limited space for transgene integration, and during adaptation may have moderately difficult quality control. An additional problem with Ads systems is related to the infection efficiency. The coxsackie adenovirus receptor (CAR) is a key determinant for the attachment and cellular uptake of adenoviruses [6]. However, CAR expression is maximal in neonates and gets reduced rapidly after birth in several organs such as heart, muscle, and brain resulting in lower penetration rate of adenovirus vectors [7].

Recombinant AAV vectors are able to effectively transduce foreign genes to a variety of cell types including both dividing and postmitotic cells in both in vitro and in vivo experimental systems [8]. The AAV-based systems have a number of favourable attributes, such as lack of parental agent pathogenicity and vector-related cytotoxicity, minimal immunogenicity, and the capacity for stable long-term transgene expression. The main disadvantages of AAV vector-based approaches include the difficulty of producing high-titer virus stocks of consistent purity and bioactivity, and the limited packaging capacity of a maximum 4.8 kb insert-size [9]. Lentivirus-based gene transfer has been reported in a wide variety of cell types, including cardiomyocytes [10]. Current lentiviral vectors are capable of transducing mitotically quiescent cells, particularly within the cardiovascular system. The strengths of this system include the ability of long-term stable transgene expression, an increased packaging capacity compared with AAV, and the other commonly used integrating vector [11]. The disadvantages of lentivirus-based gene delivery systems are the relative low level of transgene expression and the limited transgene carrying capacity of the virus.

Pseudorabies virus (PRV), a causative agent of Aujeszky's disease of swine, is an alpha-herpesvirus belonging to family of Herpesviridae. Several previous reports have been successful in construction of PRVs for delivering foreign genes to neurons [12]. PRV is especially an important tool for labelling neural circuits [13], which was combined with delivery of activity markers to labelled neurons [14].

In this study, we have developed a method for short-term culture of isolated myocytes that retains their morphological as well as physiological integrity and a pseudorabies virus (PRV)-based system for delivery of foreign genes to adult cardiac muscle cells. Troponin (TN-L15) [15], an FRET-based activity marker was selected as a model transgene system. Troponin is a genetically-encoded fluorescent  $Ca^{2+}$  sensor that is suitable for the imaging of cellular activity in neurons, muscle, and cardiac cells. Pseudorabies virus was modified to disable to replicate on postmitotic cells but retaining its capacity for high titer multiplication on dividing cultured epithelial cells. The strengths of the presented system is a very high gene transfer efficiency, low toxic effect, the capacity for relative long-term transgene expression, and

the delivery of large or multiple transgenes. The suitability of PRV-based gene delivery technique was evaluated in several ways. We performed simultaneous measurements in non-infected control and infected myocytes following isolation and infection. Retained functionality of the sarcolemmal ion channels was tested by measuring a characteristic multi-channel current  $I_{to}$ . Intactness of the excitation-contraction coupling (ECC) machinery and contractile function was demonstrated by monitoring intracellular calcium transients and cell shortening. Finally, undisturbed functioning of the transmitted gene was verified by FRET monitoring of the calcium transient currents.

## 2. Materials and Methods

**2.1. Cells and Viruses.** The wild type Kaplan strain [16] of pseudorabies virus (PRV) was used as parent strain for the generation of recombinant viruses used in this study. Viruses were propagated on subconfluent monolayer of porcine kidney (PK-15) cells. Cells were grown in DMEM supplemented with 5% fetal calf serum at 37°C with 5% CO<sub>2</sub>. The stocks of PRV were prepared as follows: PK-15 cells were infected with PRV at a multiplicity of 1 p.f.u./cell, harvested after 24 hours, followed by freezing and thawing three times. The cells were then centrifuged and the pellet discarded. The supernatant fluid was stored in 400 µL aliquots at -80°C.

**2.2. Reporter Genes.** Tn-15 (troponin), an FRET-based fluorescent calcium sensor [15], was used as an activity marker in our experiments. The troponin gene was placed under the control of the major immediate early promoter of human cytomegalovirus (pCMV), which provided a high level of gene expression. The marker gene expression cassette also contained simian virus 40 (SV40)-derived sequences including polyadenylation signal and transcription termination sequences. In addition, a lacZ gene equipped with the above regulatory sequences was also used as a reporter gene for the identification of mutant viruses.

**2.3. Construction of Targeting Vectors.** In our system, a typical targeting plasmid contains a marker gene expression cassette flanked by subcloned viral DNA sequences, which serves as homologous DNA regions for the insertion of marker genes via homologous recombination. The DNA sequences used for mutagenesis and as insertion sites for reporter genes are listed on Figure 1.

**2.3.1. Ribonucleotide Reductase (RR) Gene.** The details of the construction of RR gene targeting vector was described elsewhere [17]. Briefly, a 5-kbp *SalI*-F fragment of PRV DNA containing both subunits of RR gene was isolated and cloned to pRL494, a palindrome-containing positive selection vector [18]. This plasmid was cut with *ScaI* and *MluI* restriction endonucleases generating a 1805-bp deletion, which included a 1789-bp DNA fragment from the 3' end of large (RR1) and a 7-bp DNA sequence from the 5' end of small subunit (RR2) of ribonucleotide reductase gene. Subsequently, free DNA ends were filled up by Klenow

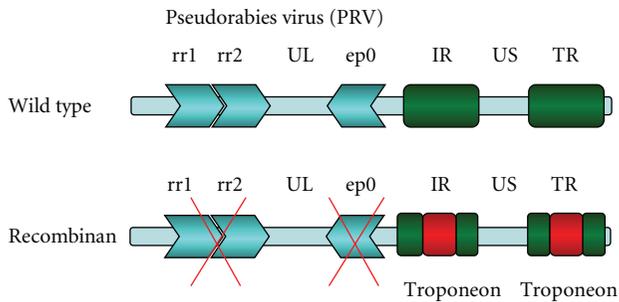


FIGURE 1: This panel shows a part of recombinant herpesvirus genome. To adapt for applicability as virus vector we deleted the two ribonucleotide reductase (rr) and the early protein (ep0) genes from the virus genome by homologue recombination. The rr1 and rr2 genes are in charge for production of raw material of viral DNA. The recombinant virus is not able to productively infect postmitotic cells without rr genes, while ep0 gene is responsible for virus reactivation from latency. Troponin calcium sensor gene was inserted into the antisense promoter region in two copies.

enzyme followed by attaching an *EcoRI* linker to the blunt DNA ends. As a final step, a lacZ gene expression cassette bracketed by *EcoRI* sites was subcloned to the *EcoRI* site of this plasmid.

**2.3.2. Early Protein 0 (EP0) Gene.** Generation of EP0 gene targeting vector was described previously [19]. Briefly, the 9.4-kb *KpnI*-F DNA fragment containing the EP0 gene was subcloned into pRL425; followed by cleaving with *Bam*HI restriction endonuclease, which resulted in the deletion of a 1390-bp sequence, including almost the entire EP0 gene. Free DNA ends were converted to *EcoRI* sites via linker insertion and then religated. The same lacZ gene expression cassette as above was subcloned to the *EcoRI* site of this plasmid.

**2.3.3. Putative Antisense Promoter (ASP) Region.** The 4.9-kb *Bam*HI-8' PRV DNA fragment was isolated and subcloned to pRL525 cloning vector [18, 20]. The *Dra*I site located in the putative TATAA box of ASP was used for the insertion of *EcoRI* thereby destroying its potential function. The fluorescent marker gene expression cassettes (GFP, troponin) were inserted to the *EcoRI* site of this plasmid.

**2.4. Generation and Isolation of Recombinant Viruses.** Recombinant viruses were generated by means of homologous recombination between parent PRV genomes and targeting plasmids containing reporter genes bracketed by virus sequences. Viral DNA was transfected to actively growing PK-15 cells along with the appropriate targeting plasmid using lipid-mediated gene delivery (Lipofectamine 2000 Reagent, Invitrogen). Viral DNA for the transfection was prepared from virions purified from the medium of infected cells showing total cytopathic effects by isopycnic centrifugation on a discontinuous gradient, as described previously [12]. The lacZ gene-expressing viruses were screened based on their blue plaque appearance in the presence

of 5-bromo-4-chloro-3-indolyl-*b*-D-galactopyranoside (X-Gal), the chromogenic substrate of  $\beta$ -galactosidase. Blue plaques were picked and plaque purified to complete homogeneity. Plaques formed by recombinant viruses carrying the fluorescent markers were detected visually on the basis of their fluorescence. Recombinant viruses were isolated by 6–15 cycles of plaque purification procedure using a fluorescence microscope (Olympus IX-71).

**2.5. Isolation of Adult Canine Cardiomyocytes.** All experiments were conducted in compliance with the *Guide for the Care and Use of Laboratory Animals* (USA NIH publication No. 85-23, revised 1985). The protocols were approved by the review board of the Committee on Animal Research (CAR) of the Albert Szent-Györgyi Medical University. The modified protocol for cell isolation was based on established procedures described earlier in detail [21].

Canine ventricular myocytes were enzymatically dissociated as follows. A portion of the left ventricular wall containing an arterial branch large enough to cannulate was perfused in a modified Langendorff apparatus with solutions in the following sequence:

(1) normal Tyrode's solution (10 minutes), (2)  $\text{Ca}^{2+}$ -free Tyrode solution (10 minutes), and (3)  $\text{Ca}^{2+}$ -free Tyrode solution containing collagenase (type I, 0.66 mg/mL) and bovine serum albumin (fraction V, fatty acid free, 2 mg/mL) (15 minutes). Protease (type XIV, 0.12 mg/mL) was added to the final perfusate while and another 15–30 minutes of digestion was allowed. Cells were stored in KB solution. The composition of solutions were (in mM): (a) *normal Tyrode's solution*—NaCl 135, KCl 4.7,  $\text{KH}_2\text{PO}_4$  1.2,  $\text{MgSO}_4$  1.2, HEPES 10,  $\text{NaHCO}_3$  4.4, glucose 10, and  $\text{CaCl}_2$  1.0 (pH 7.2 adjusted with NaOH); (b)  *$\text{Ca}^{2+}$ -free Tyrode solution*—NaCl 135, KCl 4.7,  $\text{KH}_2\text{PO}_4$  1.2,  $\text{MgSO}_4$  1.2, HEPES 10,  $\text{NaHCO}_3$  4.4, Glucose 10, and taurine 20 (pH 7.2 adjusted with NaOH); (c) *KB solution*—KOH 90, L-glutamic acid 70, taurine 15, KCl 30,  $\text{KH}_2\text{PO}_4$  10,  $\text{MgCl}_2$  0.5, HEPES 10, glucose 11, and EGTA 0.5 (pH 7.3 adjusted with KOH). All chemicals used in this method were purchased from Sigma Chemical Co.

**2.6. Culture and Infection of Myocytes.** The entire culture procedure was performed in a class II flow hood. The freshly isolated myocytes were centrifuged five times for 1 minute at 50 g in sterile 10% PBS. The supernatant was replaced first by 500  $\mu\text{M}$  then by 1 mM  $\text{Ca}^{2+}$  containing PBS solution. The mild centrifugation steps removed the majority of bacterial cells, most nonmyocytes and nonfunctioning myocytes. Precipitated cells were resuspended in culture medium and plated on laminin coated (1  $\mu\text{g}/\text{cm}^2$ ) sterile cover glass at densities of up to  $10^3$  rod-shaped cells  $\text{cm}^{-2}$ . Cells were left 4 hours to attach to the plate and after this time period non attached cells were removed. Following the first medium change, subsequent medium changes were carried out every day. Culture medium consisted of serum-free medium 199 (M199) supplemented with 25 mM  $\text{NaHCO}_3$ , 5 mM ceratine, 2 mM L-carnitine, 5 mM taurine, 100 units/mL insulin (CCTI supplemented medium) and

50  $\mu\text{g}/\text{mL}$  gentamycin. All chemicals used in this procedure were purchased from Sigma. Cells were maintained at 37°C under sterile conditions in an incubator ventilated with 5%  $\text{CO}_2$  and 95% air. After 4 hours, plate attached cells were infected. Freshly isolated canine cardiomyocytes were first washed with PBS, followed by low-speed centrifugation and resuspension in culture medium. Subsequently, after a 4-hour incubation at 37°C in a  $\text{CO}_2$  incubator, cells were infected with various titers of recombinant viruses for 12 hours then washed and the culture medium was changed. Infected cells were used for analysis at various time points.

*2.7. Evaluation of Infection Efficacy and Morphological Changes in Cultured Cardiomyocytes.* Cell shape and morphology are closely linked with some aspects of cell function such as excitation-contraction coupling. Therefore, monitoring these properties may give indications of physiological changes that are occurring. Morphological changes of cells were observed by light microscope on a daily basis parallel with physiological measurements. Troponeon-positive cells were examined by fluorescence microscopy from one to three days following isolation at standard titer of viruses. Infection efficacy was determined separately for infected cells by manual cell counting using a fluorescent microscope (Olympus IX-71).

## 2.8. Evaluation of the Physiological State of the Virus-Infected Cells

*2.8.1. Evaluation of the Electrophysiological State of the Cells by  $I_{\text{to}}$  Analysis.*  $I_{\text{to}}$  currents were measured at 37°C using the whole-cell configuration of the patch-clamp technique. Measurements were performed daily starting with day 0, when only freshly isolated noninfected myocytes were investigated. In the following days control and infected cells were studied separately by placing cover glasses with the attached cells in the recording chamber mounted on the stage of an inverted microscope equipped with epifluorescence assembly (Olympus IX50, Tokyo, Japan). Only rod-shaped cells with clear cross-striations and relatively strong GFP signal were used. HEPES-buffered Tyrode's solution containing (in mM): NaCl 144,  $\text{NaH}_2\text{PO}_4$  0.33, KCl 4.0,  $\text{CaCl}_2$  1.8,  $\text{MgCl}_2$  0.53, glucose 5.5, and HEPES 5.0 at pH of 7.4 (by NaOH) served as normal superfusate. Cell capacitance ( $199.3 \pm 13.7$  pF,  $n = 69$ ) was determined by applying a 10 mV hyperpolarizing pulse from a holding potential of  $-10$  mV. Cell capacity was calculated as the integral of the capacitive transient divided by the amplitude of the voltage step. Patch-clamp micropipettes were fabricated from borosilicate glass capillaries (Clark, Reading, UK) using a P-97 Flaming/Brown micropipette puller (Sutter Co, USA). These electrodes had resistances between 1.5 and 2.5 Mohms when filled with pipette solution containing (in mM): K-aspartate 100, KCl 45,  $\text{K}_2\text{ATP}$  3,  $\text{MgCl}_2$  1, EGTA 10, and HEPES 5. The pH of this solution was adjusted to 7.2 by KOH. Nisoldipine ( $1 \mu\text{M}$ ) (obtained as a gift from the Bayer AG, Leverkusen, Germany) was used in the external solution to eliminate inward  $\text{Ca}^{2+}$  current

( $I_{\text{Ca}}$ ). Membrane currents were recorded with Axopatch-200B patch-clamp amplifiers (Axon Instruments, USA). After establishing a high (1–10 Gohm) resistance seal by gentle suction, the cell membrane beneath the tip of the electrode was disrupted by suction or by application of 1.5 V electrical pulses for 1–5 milliseconds. The series resistance was typically 4–8 Mohm before compensation (50–80%, depending on the voltage protocols). Experiments where the series resistance was high, or substantially increased during measurement, were discarded. Membrane currents were digitized using a 333 kHz analogue-to-digital converter (Digidata 1200, Axon Instruments) under software control (pClamp 8.0, Axon Instruments). Analyses were performed using pClamp 8.0 (Axon Instruments) software after low-pass filtering at 1 kHz. All patch-clamp data were collected at 37°C.

*2.8.2. Evaluation of Cell Function by Monitoring  $[\text{Ca}^{2+}]_i$  Transient and Cell Shortening.* 24–72 hours following viral infection cultured myocytes were loaded by incubation for 20 minutes with the acetoxymethyl ester (AM) form of a single wavelength calcium-sensitive fluorescent dye (Fluo-4, Molecular Probes Inc. 1–2  $\mu\text{M}$  from a stock of 1 mM in DMSO + 20% pluronic acid) at room temperature. The technique for calcium transient detection was based on established procedures described earlier [22]. After this incubation period the cover glass with attached cultured myocytes was mounted in a low volume imaging chamber (RC47FSLP, Warner Instruments, USA) and placed on the stage of an inverted fluorescent microscope (IX71, Olympus, Japan). The cells were superfused with normal Tyrode solution at 37°C (1 mL/min). Myocytes were stimulated at a constant frequency of 1 Hz through a pair of platinum electrodes by an electronic stimulator. The dye was excited at 480 nm, fluorescence emission was recorded at 535 nm (Chroma, USA). Optical signals were recorded by a photon counting photomultiplier module (H7828, Hamamatsu, Japan), and sampled at 1 kHz. Measurements were performed and data were analyzed using the Isosys software (Experimetria Ltd, Hungary). Cell shortening from both ends was determined by a video edge detection system (VED-105, Crescent Electronics, USA). All experiments were performed at 37°C, using an automatic temperature controller (TC-344B, Warner Instruments, USA).

*2.8.3. Evaluation of the Functionality of the Transferred Gene by FRET Measurement.* For testing the functionality of the transferred gene, following the incubation period the cover glass with the attached primary cultured cardiomyocytes was mounted in a low volume imaging chamber (RC47FSLP, Warner Instruments, USA) on the stage of an Olympus IX71 inverted microscope outfitted with a 75 W Xe arc lamp for epi- and a 100 W tungsten-halogen lamp for transillumination. The myocytes in the chamber were superfused with normal Tyrode solution (1 mL/min) and stimulated at a constant frequency of 1 Hz by an electronic stimulator (Experimetria) through a pair of platinum electrodes. All measurements were performed at

37°C, by using an automatic temperature controller (TC-344B, Warner Instruments, USA). The optical signals were typically monitored 20–36 hours after infection in selected myocytes expressing the genetically encoded FRET-based sensor troponin (csTn-C-L15). Cells with medium-high level of fluorescence were selected for measurement. Though often troponin fluorescence is already visible 16 hours following infection, optical signals were found to be optimal for recording after 24 hours.

Fluorescent optical signals were monitored by a dual channel photon counting system. An Olympus filter cube containing a CFP excitation filter (436/20 x) and a CFP dichroic mirror (455DCLP) but no emission filter was inserted into the light path of the microscope. A connector box for dual emission fluorescence with two interchangeable dichroic mirror holders and three detector ports (P226/0II/011, Cairn Research Ltd, UK) equipped with two photons counting photomultiplier modules (H7828, Hamamatsu, Japan) was attached to the left side port of the microscope. The fluorescent signal from the microscope was splitted by a YFP dichroic mirror (515LP). A second dichroic mirror (565DCLP) was used to separate the longer wavelength (FRET) fluorescence from the red filtered (> 580 nm) transillumination light used for visual observation of the cell by a video camera mounted on the 3rd back port of the connector box. The splitted fluorescence components were band pass filtered (480/30 m and 535/30 m for CFP and citrine, resp.), integrated with 1 milliseconds time resolution by two photons to voltage converter units (Ionoptics, US) and collected for data processing. All interference filters and dichroic mirrors used are available in filter sets 31036V2 and 41028 from Chroma Technology Corporation (US).

Changes in  $[Ca^{2+}]_i$  levels were characterized by the ratio of the emitted fluorescence intensities obtained at 480 and 535 nm wavelengths, ( $F_{FRET\_CITRINE} 535 / F_{CFP} 480$ ) following optical signal correction steps for photobleaching and non-specific background fluorescence. Photobleaching leads to a steady decrease in the FRET ratio over time, because citrine is less photostable than CFP. To correct for bleaching, we multiplied the intensity of the CITRINE channel by a correction factor calculated from the intensity shift between two time points. For background correction we subtracted from the optical signals the nonspecific background fluorescence determined at both wavelengths by moving off the cell from the light path:  $F_{RATIO\_CORR} = (F - F_0)_{CITRINE} / (F - F_0)_{CFP}$ . The background-corrected fluorescence ratio versus time curve can be considered as a representation of the intracellular  $Ca^{2+}$  transients Figure 7.

**2.9. Data Analysis.** All patch-clamp results and optical measurements were compared using Student's *t*-tests for paired and unpaired data and expressed as mean  $\pm$  SEM values. Statistical significance of differences obtained between control and virus infected preparations was evaluated with Student's *t*-test for paired or unpaired data, as relevant. Differences were considered significant when  $P < .05$ .

TABLE 1: This table shows the survival of noninfected and virally infected cells from day 0 to day 5. It can be seen that virally infected cells exhibit slightly better survival than noninfected cells, for which the reason remains to be ascertained.

		Day 0	Day 1	Day 2	Day 3	Day 4	Day 5
Control	Average (%):	100	60,0	44,9	4,3	2,8	2,2
	Standard error:		6,0	8,4	0,5	0,3	0,3
Virus infected	Average (%):	100	85,5	69,2	12,8	3,1	1,3
	Standard error:		4,7	9,3	3,7	1,5	0,6

### 3. Results

#### 3.1. Morphological Changes of Cultured Cardiomyocytes.

Using the described method for isolation of adult dog left ventricular myocytes, we routinely obtained a high yield (more than 80%) and high (more than 80%) percentage of rod-shaped myocytes that were suitable not only for acute functional studies but, more importantly, for short-term culture and gene transfer. Figure 2(a) shows a transillumination image of freshly isolated and 1-day-old cultured cardiomyocytes from the left ventricle. To establish optimal surviving conditions several culture conditions were tested based on microscopic evaluation of changes in cellular morphology during the four days of culture. Cultured cells were used 1–3 days after isolation. During this period, visible small-scale changes in cell shape and cross-striation could be observed. Figure 2(b) shows representative photographs of canine myocytes over time in culture. Features typical of acutely isolated (Day 0) cells were the rod shape with rectangular stepped ends and clear cross-striations. After 1 day (Day 1) in culture, the cells were still rod-shaped with clear cross-striations; however, the ends of the cells started to become slightly rounded in appearance. After 3 days (Day 3) in culture, cells remained rod-shaped and cross-striated, and the main change was that cell ends became progressively more rounded (see Table 1 for cell survivals).

#### 3.2. Efficacy of Virus Infection in Cultured Cardiomyocytes.

Survival rates were found to be dependent on the isolation procedure, density of the attached myocytes, and the applied virus titer. Intriguingly, a higher total number of viable cells were observed on the laminin-coated surfaces after plating in CCTI supplemented medium. As respective panels of Figures 2 and 3 (upper panel) show, even after three days, the cell culture contained a substantial number of good quality cells both in the control and virus-infected groups. Surprisingly, a moderately but consistently higher cell survival rate was found in virus-infected groups as compared to noninfected groups (not shown). The infection efficiency was found to be 100%, that is 24 hours postinfection every surviving cells emitted fluorescent signals provided that high dose of virus was used for the infection (Figure 3 bottom panel).

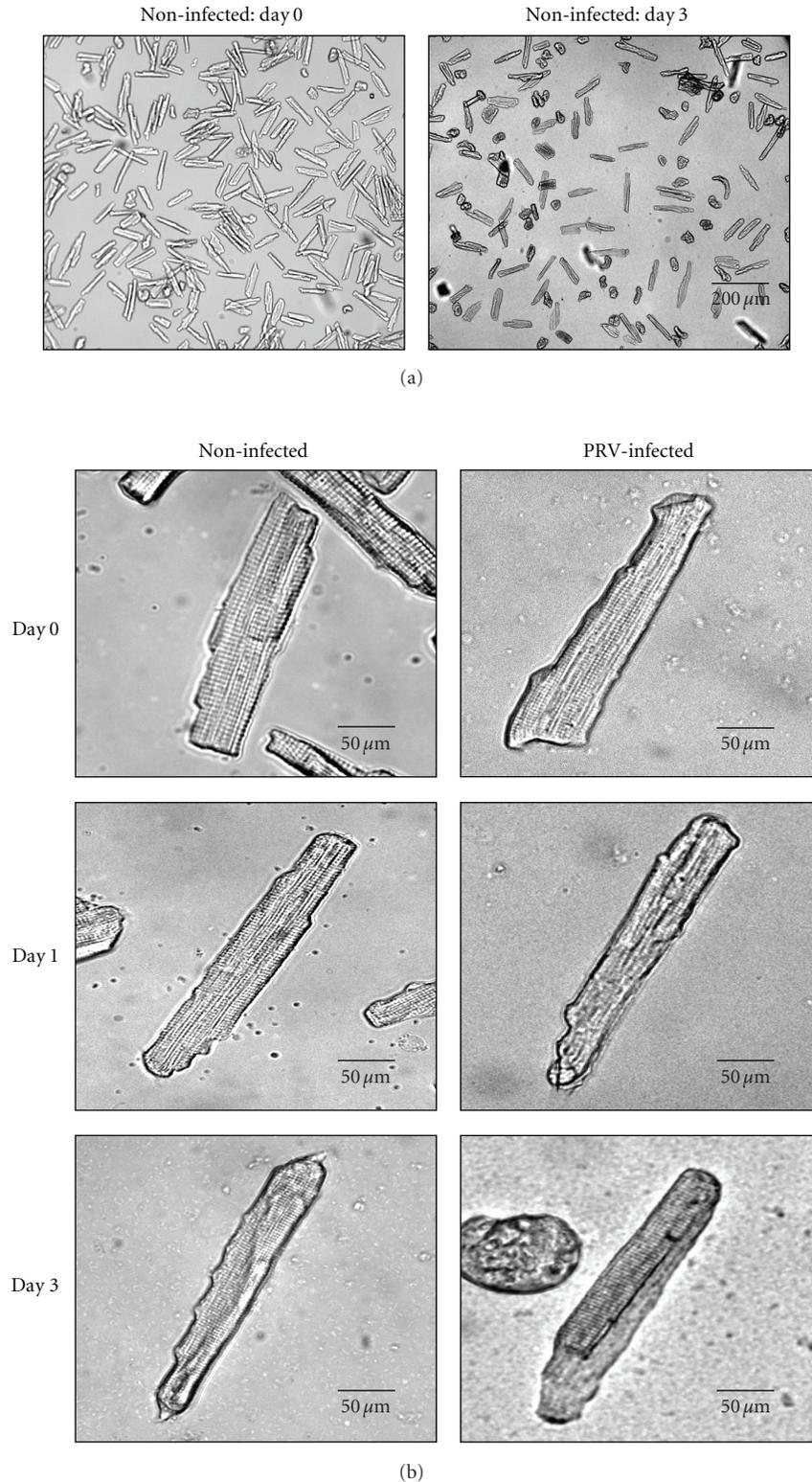


FIGURE 2: Representative low- and high-magnification transillumination of adult dog myocytes after isolation (day 0) and 1–3 days of culture (day 1, day 3). (a) shows the yield of the myocytes before and one day after culturing and infection. The high-magnification transillumination (b) shows the morphology of adult dog myocytes after isolation (day 0) and 1–3 day of culture (day 1, day 3) and the morphological changes of living cells, versus the culturing time from noninfected (right) and the virus-infected groups (left). After one day plated myocytes more than 80% displayed a rod-shaped morphology and healthy cross-striation. After 3 days (day 3) in culture, cells remained rod-shaped and partially cross-striated, and the main change was that cell ends became progressively more rounded.

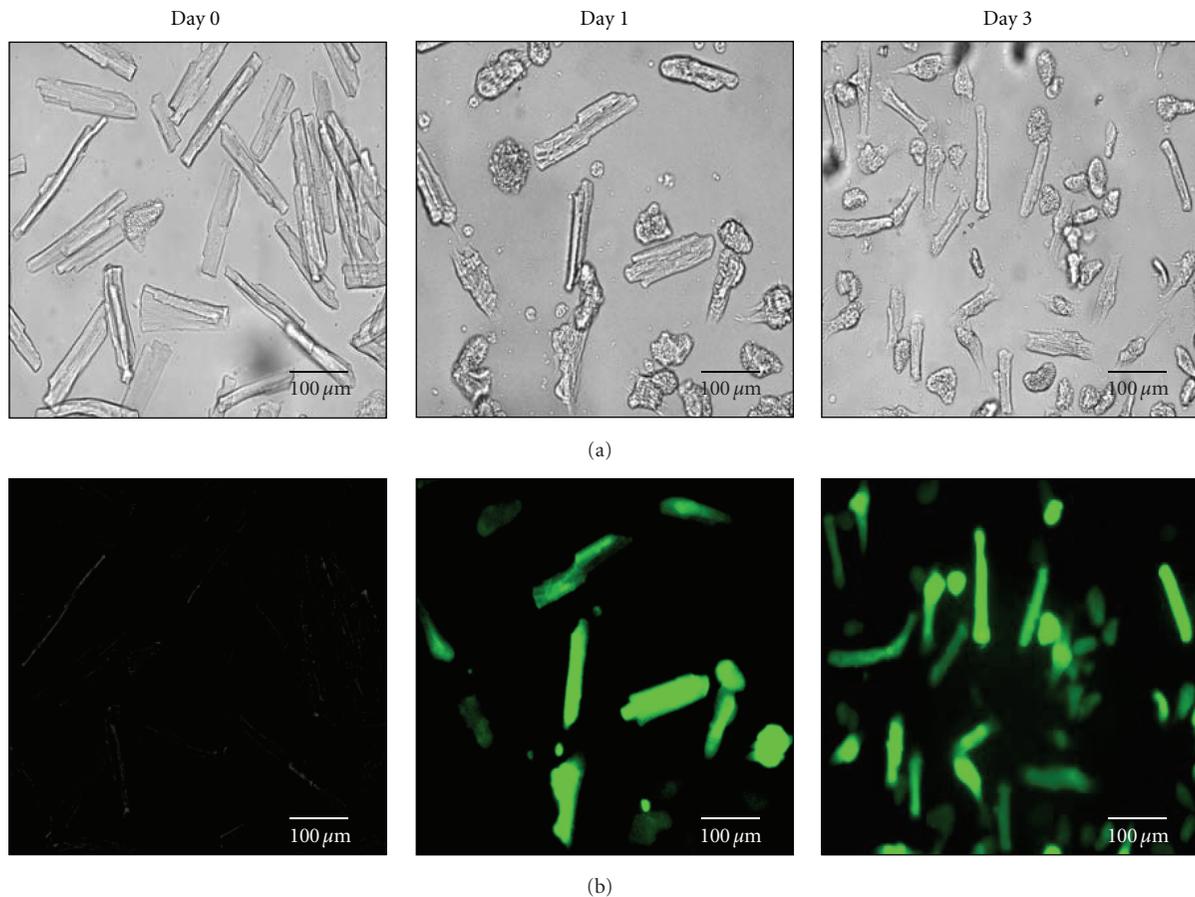


FIGURE 3: (a) Representative light and (b) fluorescent microscopic images of cell culture, before and after 1 day and 3 days of virus infection. Expression of recombinant pseudorabies-viral troponin transgenes appeared on high level in cultured adult dog ventricular myocytes. The isolation of adult dog left ventricular myocytes yielded more than 80% living cells. After three days the troponin expression and the physiological features of survival myocytes was appropriate for the physiological studies.

**3.3. Whole-Cell Patch-Clamp Recordings.** The whole-cell configuration of the patch-clamp technique was used to record the transient outward  $I_{to}$  current.  $I_{to}$  was chosen as a physiological assay because it is a large current that can be measured relatively easily in isolated canine ventricular myocytes. The current was activated by 300 milliseconds long depolarizing voltage pulses from the holding potential of  $-90$  mV to test potentials ranging from 0 to  $+60$  mV with a pulse frequency of 0.33 Hz. The amplitude of  $I_{to}$  was measured as the difference between the peak and the sustained current at the end of the voltage pulse. Figure 4(a) shows typical recordings of  $I_{to}$  measured after one (Day 1) and three days (Day 3) of culture either in control (top panels) or virus-infected cells (bottom). Figure 4(b) summarizes all  $I_{to}$  measurements performed after 0–4 days of culture in both groups. As the corresponding panels of Figure 4 show, the amplitude of the  $I_{to}$  was reduced by less than 10% in virus-infected myocytes (VM) compared to control noninfected cells (CM) even after 4 elapsed days. Moreover current amplitude was somewhat larger in PRV-infected cells compared to that of observed in control cells. Mean  $I_{to}$  density (Figure 4(b)) was similar for CM and VM cells after

1 day. The  $I_{to}$  current density in VM changed in four days from  $19.6 \pm 1.4$  to  $24.6 \pm 2.6$  pA/pF ( $n = 10-12$ ), which corresponds well with that observed in CM (from  $19.3 \pm 2.1$  to  $17.1 \pm 1.5$  pA/pF,  $n = 8-10$ ). Although mean  $I_{to}$  density of several-day-old (2, 3, 4) cultured myocytes was significantly larger in VM than CM. The  $I_{to}$  kinetics (activation and inactivation properties) was also not significantly altered by the virus infection.

**3.4. Parameters of Intracellular  $Ca^{2+}$  Transients and Contractility.** Cultured myocytes were stimulated at a constant frequency of 1 Hz through a pair of platinum electrodes. Similarly to  $I_{to}$  measurements, the steady-state  $[Ca^{2+}]_i$  transient and contractile function were measured and compared on a daily basis both in the infected and control cell populations. Original recordings of  $[Ca^{2+}]_i$  transients and cell shortening before and after virus infection are presented in Figure 5. The calcium transient kinetics were significantly distorted in culture, but were also not significantly altered by the virus-infection. As Figure 6 summarizes, we found no statistically significant differences in either the amplitude of

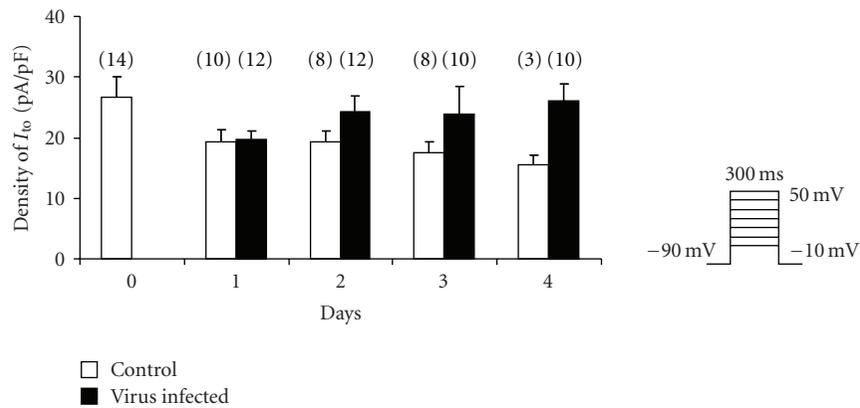
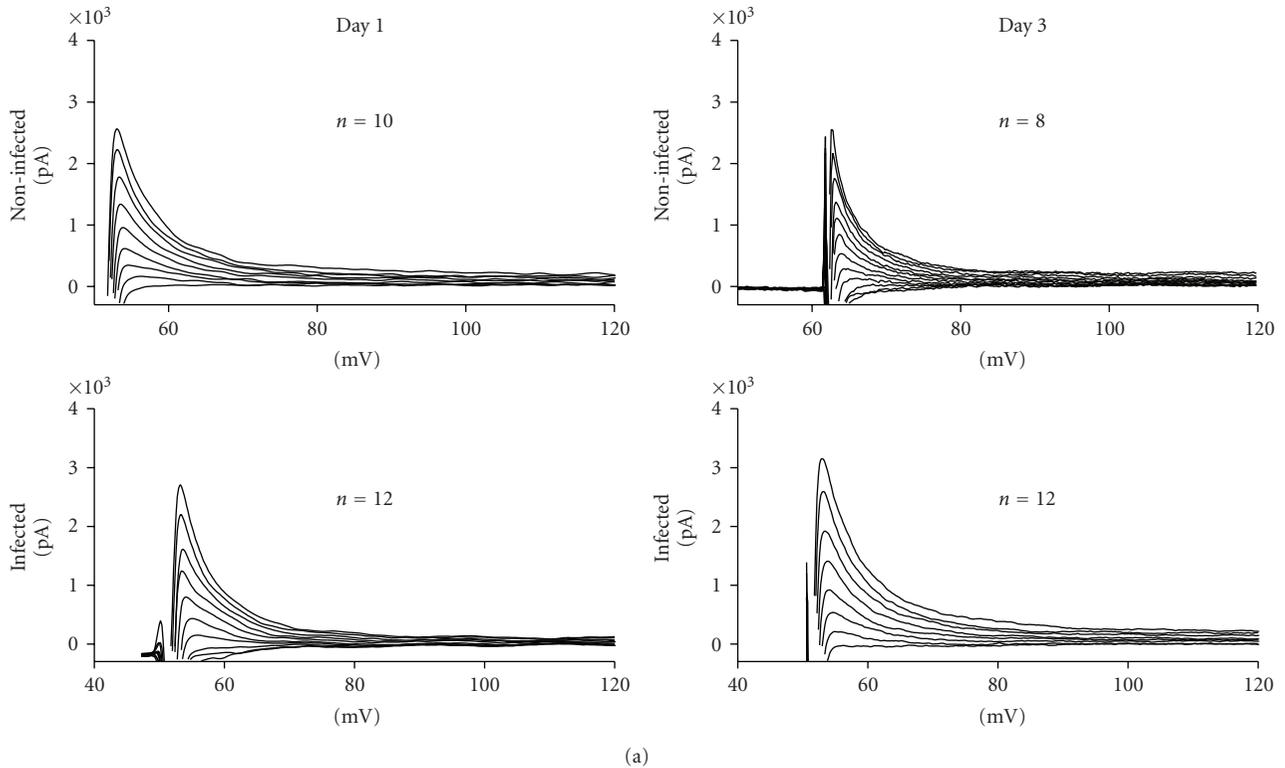


FIGURE 4: (a) Transient outward K current ( $I_{to}$ ) recordings from control and from virus-infected myocytes after (a) 1 and (b) 3 days of culture. Inset shows applied voltage protocol. (b) shows the  $I_{to}$  current densities from control and from virus-infected myocytes after 1 to 4 days long culture. Data represent means  $\pm$  SEM, and  $n$  represents the number of experiments.

calcium transient (A) or diastolic calcium level (B) between the studied groups. Moreover, transient decay time (C) in noninfected control myocytes was significantly larger than in the virus infected group after two and three days in culture. Conversely in virus-infected group's decay time was similar to freshly isolated cells. The cell-shortening measurements (D) represent a significant decline in noninfected cell culture after 1 and 3 days. Data indicate no significant changes in these parameters.

**3.5. Functionality of FRET-Based Calcium Sensor.** Freshly isolated ventricular myocytes were infected with the virus following 4 hours plating and fluorescence could already be detected within 16 hours. The functionality of the transferred gene was verified by monitoring  $[Ca^{2+}]_i$ . Figure 7 shows two typical fluorescence signal emissions measured in troponin-expressing cells obtained at 485 nm and 535 nm excitation (upper panel). The lower panel shows the ratio of the citrine and CFP emitted signals representing the changes of the  $[Ca^{2+}]_i$  levels in the studied cells. This measurement

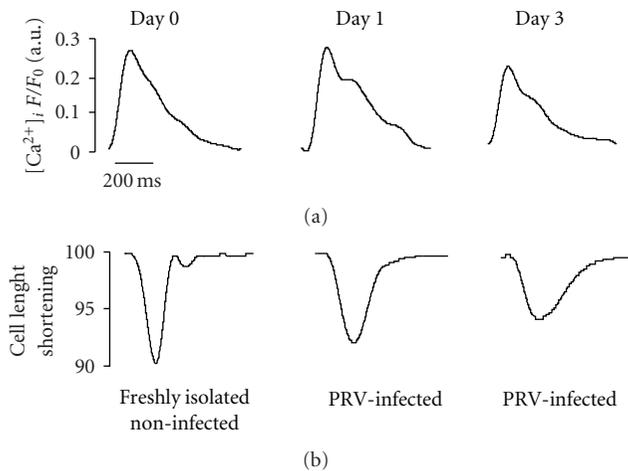


FIGURE 5: Original  $\text{Ca}^{2+}$  transients and cell contraction recording in cardiomyocytes cultured for 1 and 3 days. The top panel shows the intracellular  $\text{Ca}^{2+}$  concentration as  $\text{Ca}^{2+}$  transient fluorescent signal (F535 nm), and bottom panel shows the changes in cell length.

clearly shows that virally-encoded troponin retained its ability of indicating  $[\text{Ca}^{2+}]_i$  levels in cardiomyocytes.

#### 4. Discussion

Recent advances of transgenesis and gene targeting technologies have heralded a new era of cellular physiology to study molecular function using genetically engineered animal models and genetically encoding vector-based gene transfer systems. Practically, the mouse is the only mammalian species in which transgenic technology efficiently works. Several transgenic and gene-targeted models have been generated for the overexpression [23, 24], and genetic ablation of key proteins [25, 26] governing cardiac structure and function.

However, the mouse is not an ideal species for modelling human disease like heart failure, myocardial infarction, and arrhythmia studies, because the mouse heart has distinctly different action potential waveform due to its different underlying ionic current structure [27, 28]. Transducing foreign genes to cardiac cells of living animals is a feasible technique in several animal models; however, performing electrophysiological measurements *in vivo* is not easy therefore *in vitro* electrophysiological techniques are usually used in cultured cells for this purpose. Similar experiments were performed in a recently published work using cultured human atrial myocytes [29] and myocytes isolated from rat, a species from which producing long-term cell culture is much easier.

Gene transfer to cardiac myocytes has been traditionally carried out in neonatal cells. However, these cells undergo differentiation, and as a result, this model is inappropriate for certain experiments since differentiating cells have different currents than that of from adult cells. Therefore, *in vitro* transducing of isolated cardiac myocytes can be a useful

alternative for investigations of cardiovascular cell physiology and diseases.

The structural changes during culturing ventricular myocytes were studied and as well [30, 31]. These changes are associated with culturing procedures associated effects, which may change a number of physiological properties of cells in culture. Therefore, some possible alterations in the  $\text{Ca}^{2+}$  handling and sarcolemmal ionic currents may have been diminished in cultured cells.

Another potential application of our viral gene transfer strategy is to introduce siRNA for silencing pore or auxiliary ionic channel subunits underlying transmembrane ionic currents. In this study we opted for canine model, since the dog unlike the mouse and rat is known to have action potential and ionic current characteristics similar to human [32]. We have chosen the transient outward current for testing the possible effects of viral infection because  $I_{to}$  is a relatively large current and present in all cells. Also it can be relatively easily measured. The  $I_{to}$  gene structure is rather complex. The  $I_{to}$  current in canine myocytes resembles human myocytes and has a large conducting pore forming unit Kv4.3 connected with several auxiliary subunits such as KCHIP2, KCNE2, and DPPX [33]. In the present work, we measured the effect of the viral infection on  $I_{to}$  and we found that the virus did not affect either the density or kinetics of  $I_{to}$  current (Figure 4).

Since their introduction more than twenty years ago of  $\text{Ca}^{2+}$ -sensitive fluorescent dyes have been used to evaluate intracellular free calcium concentration ( $[\text{Ca}^{2+}]_i$ ) [34], enabling investigators to gain unprecedented insights into the mechanisms of cell signalling. The dynamics of changing  $\text{Ca}^{2+}$  concentrations at cellular level and well-defined subcellular spaces in excitable cells, during the course of membrane depolarization can now be understood in the context of disease processes such as cardiac arrhythmias and heart failure. Although many details of excitation-contraction coupling were described previously, novel quantitative fluorescence techniques have significantly improved our understanding of major cell regulatory pathways.

Fluorescent dyes commonly used for  $\text{Ca}^{2+}$  imaging are based on bis(2-aminophenoxy)ethane tetraacetic acid (BAPTA) structure and subsequently on bis(2-aminoethyl ether) tetraacetic acid (EGTA) backbone conjugated to a fluorescent moiety. Fura-2, indo-1, and fluo-4 are the most widely used fluorescent dyes for monitoring intracellular  $[\text{Ca}^{2+}]_i$ ; in their acetoxymethyl ester form they can relatively easily be loaded into cells, are relatively photostable, and are well suited for reliable detection of changes in  $\text{Ca}^{2+}$  concentration under physiological conditions [35]. These  $\text{Ca}^{2+}$  indicators with low molecular weight have some important advantages as great dynamic range, increased sensitivity, high fluorescent intensity, and rapid response kinetics, but also have some limitations, being not targetable and subcellular localization of dye cannot be controlled [36]. Recently developed genetically encoded  $\text{Ca}^{2+}$  indicators (GECIs) are defined as optical  $\text{Ca}^{2+}$  sensors produced by the expression of specific genes. These proteins contain a light-emitting fluorescent protein unit and a  $\text{Ca}^{2+}$ -responsive unit. Troponin, a typical member of the GECI family is

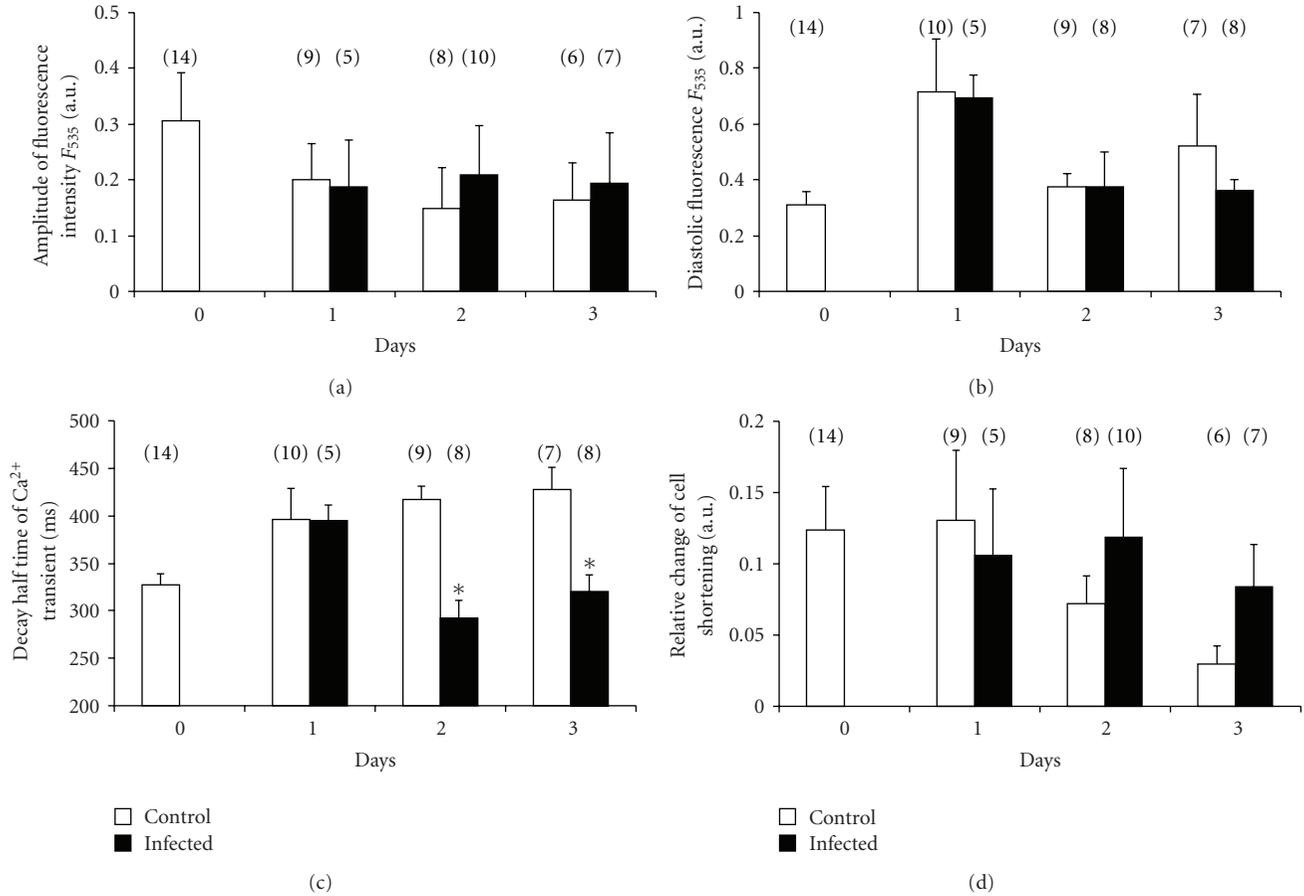


FIGURE 6: This figure shows several parameters of intracellular free calcium transient such as (a) amplitude of calcium transients, (b) diastolic calcium levels, (c) changes of calcium transient decay constant, and (d) cell-shortening measurements from recordings from control and from virus infected myocytes after 1 to 3 days long culture. Bars represent means  $\pm$  SEM, and  $n$  represents the number of experiments.

composed of a calcium-binding domain carrying a variant of troponin C protein and a pair of mutant green fluorescent proteins (CFP and Citrine) engineered for fluorescence resonance energy transfer (FRET) [15]. A significant difficulty in using these new  $Ca^{2+}$  sensors is their low efficiency of transfection into several cell types, cardiomyocytes included. On the other hand, the strengths of GECIs include the following: localization can be controlled by a custom signal sequence; it can be genetically fused to a protein of interest. GECIs can be maintained within cells over days to weeks and expression level/concentration of GECIs can be well controlled by incorporating a promoter [37].

At present, in spite of permanent improvements, techniques for introducing foreign genes to cultured adult cardiomyocytes suffer from substantial limitations, such as relatively low infection efficacy and/or cell surviving rate for the integration of transgenes be delivered [38, 39]. In addition, a number of studies demonstrated that vector associated cytotoxic effects directly affect a number of (electro) physiological properties of the cells [1].

In this study, we demonstrated that pseudorabies virus vectors can effectively penetrate cultured dog cardiomyocytes and that the transferred foreign gene (troponin) could be

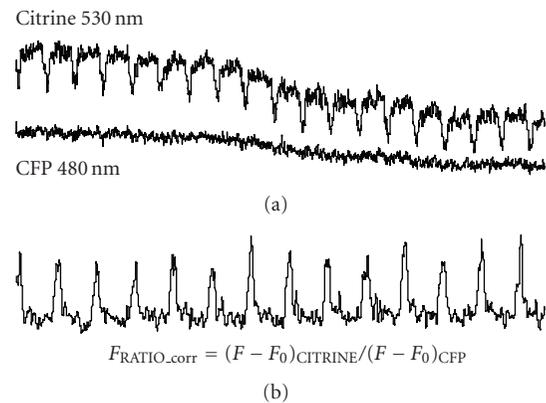


FIGURE 7: (a) Original recordings from cells expressed troponin at the excitation wavelength of (a) 485 and 535 nm and (b) the characterized calcium transient by ratio of the fluorescence intensity (FCFP 485 / FCITRINE 535) from one day after infection.

detected already at 16 hours following infection and until at least four days postinfection. Furthermore, we showed that infected cardiomyocytes tolerated the presence of PRV

vector since their electrophysiological properties were not fundamentally changed by the virus. The survival of the cells suitable for electrophysiological studies even after 4 days was high enough, proving that the virus entered the cells did not cause any observable substantial cytotoxic effects; moreover the cells displayed unaltered electrophysiological properties. This was analyzed by (1) measuring the properties of a specific transmembrane ionic current, the transient outward current ( $I_{to}$ ), which is known to be ubiquitously present in all ventricular myocytes; (2) analyzing the intracellular  $Ca^{2+}$  transient by FRET. In these measurements we found that  $I_{to}$  was present in all cells even after 72 hours of viral infection, and neither the density nor the kinetics was different than that of observed in control cells (Figure 4). On the other hand we found that the kinetics of the intracellular  $Ca^{2+}$  transient was not significantly different between infected and noninfected cells (Figures 5 and 6). Specifically, we found that infected adult dog myocytes had a lower rate of physiological degradation than that of the noninfected control cell culture in every parameter. One possible explanation for this unexpected result is that delayed apoptosis was induced by the virus infection probably by inactivating the caspase system [40]. Recent studies have suggested that the latency-associated transcript (LAT) region of herpes simplex virus type 1 (HSV-1) is effective at blocking virus-induced apoptosis in vitro in various cell types [41, 42]. Alternatively, since antiapoptotic effects depend on intracellular  $Ca^{2+}$  level [43], it is possible that  $Ca^{2+}$  level was decreased by binding a portion of  $Ca^{2+}$  to troponin therefore the lower  $Ca^{2+}$  level inhibited cell death. However, the verification of this effect needs further investigation.

## 5. Conclusions

We have developed a PRV-based vector for delivery of genetically encoded activity sensors to cultured canine cardiomyocytes. This system has several advantageous features: the virus enters to cells without destroying the intact physiological properties of the cells for a prolonged period; the virus does not change the measured physiological properties. We tested the survival rate, physiological changes on ionic current, contractile function, and  $Ca^{2+}$  signalling under effect of virus-mediated gene transfer in cultured adult canine cardiomyocytes. Our results show that novel herpesvirus-based vectors can transduce genes efficiently into nondividing cardiac myocytes. Future studies are required to evaluate, whether PRV will be effective for in vivo infection for physiological research or gene therapeutic application.

## Acknowledgments

This work was supported by Grants from the Hungarian Scientific Research Fund (OTKA NI-61902, F-67879, K-68911 to A. Varró and by: T049171 to Z. Boldogkői), Human Frontiers Science Program Young Investigator Grant to Z.B.; Hungarian Ministry of Health

(T-353/2006, T-483/2006 and T-542/2006), the National Office for Research and Technology—Ányos Jedlik Programme TECH\_08\_A1\_CARDIO08), European Community (EU FP6 Grant LSHM-CT-2005-018833, EUGeneHeart; EU FP6 Grant LSHM-CT-2006-018676, NORMACOR; EU FP7 Grant ICT-2008-224381, preDiCT) and by the Hungarian Academy of Sciences, and GVOP. The authors would like to thank Dr. Oliver Griesbeck (Max Planck Institute of Neurobiology, Martinsried, Germany) for providing TN-L15.

## References

- [1] L. G. Melo, A. S. Pachori, D. Kong, et al., "Gene and cell-based therapies for heart disease," *The FASEB Journal*, vol. 18, no. 6, pp. 648–663, 2004.
- [2] H. Ly, Y. Kawase, R. Yoneyama, and R. J. Hajjar, "Gene therapy in the treatment of heart failure," *Physiology*, vol. 22, no. 2, pp. 81–96, 2007.
- [3] R. J. Guzman, P. Lemarchand, R. G. Crystal, S. E. Epstein, and T. Finkel, "Efficient gene transfer into myocardium by direct injection of adenovirus vectors," *Circulation Research*, vol. 73, no. 6, pp. 1202–1207, 1993.
- [4] J. K. Donahue, K. Kikkawa, D. C. Johns, E. Marban, and J. H. Lawrence, "Ultraprapid, highly efficient viral gene transfer to the heart," *Proceedings of the National Academy of Sciences of the United States of America*, vol. 94, no. 9, pp. 4664–4668, 1997.
- [5] A. H. Schulick, K. D. Newman, R. Virmani, and D. A. Dichek, "In vivo gene transfer into injured carotid arteries: optimization and evaluation of acute toxicity," *Circulation*, vol. 91, no. 9, pp. 2407–2414, 1995.
- [6] W. Poller, H. Fechner, M. Noutsias, C. Tschöpe, and H.-P. Schultheiss, "Highly variable expression of virus receptors in the human cardiovascular system: implications for cardiotropic viral infections and gene therapy," *Zeitschrift für Kardiologie*, vol. 91, no. 12, pp. 978–991, 2002.
- [7] J. Nalbantoglu, G. Pari, G. Karpati, and P. C. Holland, "Expression of the primary coxsackie and adenovirus receptor is downregulated during skeletal muscle maturation and limits the efficacy of adenovirus-mediated gene delivery to muscle cells," *Human Gene Therapy*, vol. 10, no. 6, pp. 1009–1019, 1999.
- [8] Y. Maeda, U. Ikeda, M. Shimpo, et al., "Efficient gene transfer into cardiac myocytes using adeno associated virus (AAV) vectors," *Journal of Molecular and Cellular Cardiology*, vol. 30, no. 7, pp. 1341–1348, 1998.
- [9] L. Du, M. Kido, D. V. Lee, et al., "Differential myocardial gene delivery by recombinant serotype-specific adeno-associated viral vectors," *Molecular Therapy*, vol. 10, no. 3, pp. 604–608, 2004.
- [10] T. Sakoda, N. Kasahara, Y. Hamamori, and L. Kedes, "A high-titer lentiviral production system mediates efficient transduction of differentiated cells including beating cardiac myocytes," *Journal of Molecular and Cellular Cardiology*, vol. 31, no. 11, pp. 2037–2047, 1999.
- [11] J. Zhao, G. J. Pettigrew, J. Thomas, et al., "Lentiviral vectors for delivery of genes into neonatal and adult ventricular cardiac myocytes in vitro and in vivo," *Basic Research in Cardiology*, vol. 97, no. 5, pp. 348–358, 2002.
- [12] Z. Boldogkői, A. Szabó, G. Vrbová, and A. Nógrádi, "Pseudorabies virus-based gene delivery to rat embryonic spinal

- cord grafts," *Human Gene Therapy*, vol. 13, no. 6, pp. 719–729, 2002.
- [13] Z. Boldogkői, A. Sík, Á. Dénes, et al., "Novel tracing paradigms—genetically engineered herpesviruses as tools for mapping functional circuits within the CNS: present status and future prospects," *Progress in Neurobiology*, vol. 72, no. 6, pp. 417–445, 2004.
- [14] Z. Boldogkői, K. Balint, G. B. Awatramani, et al., "Genetically timed, activity-sensor and rainbow transsynaptic viral tools," *Nature Methods*, vol. 6, no. 2, pp. 127–130, 2009.
- [15] N. Heim and O. Griesbeck, "Genetically encoded indicators of cellular calcium dynamics based on troponin C and green fluorescent protein," *Journal of Biological Chemistry*, vol. 279, no. 14, pp. 14280–14286, 2004.
- [16] A. S. Kaplan and A. E. Vatter, "A comparison of herpes simplex and pseudorabies viruses," *Virology*, vol. 7, no. 4, pp. 394–407, 1959.
- [17] Z. Boldogkői, A. Braun, J. Antal, and I. Fodor, "A restriction cleavage and transfection system for introducing foreign DNA sequences into the genome of a herpesvirus," *Research in Virology*, vol. 149, no. 2, pp. 87–97, 1998.
- [18] J. Elhai and C. P. Wolk, "A versatile class of positive-selection vectors based on the nonviability of palindrome-containing plasmids that allows cloning into long polylinkers," *Gene*, vol. 68, no. 1, pp. 119–138, 1988.
- [19] Z. Boldogkői, A. Braun, and I. Fodor, "Replication and virulence of early protein 0 and long latency transcript deficient mutants of the Aujeszky's disease (pseudorabies) virus," *Microbes and Infection*, vol. 2, no. 11, pp. 1321–1328, 2000.
- [20] Z. Boldogkői, F. Erdélyi, and I. Fodor, "A putative latency promoter/enhancer (P(LAT2)) region of pseudorabies virus contains a virulence determinant," *Journal of General Virology*, vol. 81, no. 2, pp. 415–420, 2000.
- [21] A. Varró, B. Baláti, N. Iost, et al., "The role of the delayed rectifier component IKs in dog ventricular muscle and Purkinje fibre repolarization," *Journal of Physiology*, vol. 523, no. 1, pp. 67–81, 2000.
- [22] P. Birinyi, A. Tóth, I. Jóna, et al., "The Na<sup>+</sup>/Ca<sup>2+</sup> exchange blocker SEA0400 fails to enhance cytosolic Ca<sup>2+</sup> transient and contractility in canine ventricular cardiomyocytes," *Cardiovascular Research*, vol. 78, no. 3, pp. 476–484, 2008.
- [23] S. Adachi-Akahane, L. Lu, Z. Li, J. S. Frank, K. D. Philipson, and M. Morad, "Calcium signaling in transgenic mice overexpressing cardiac Na<sup>+</sup>-Ca<sup>2+</sup> exchanger," *Journal of General Physiology*, vol. 109, no. 6, pp. 717–729, 1997.
- [24] N. Chossat, F. Griscelli, P. Jourdon, et al., "Adenoviral SERCA1a gene transfer to adult rat ventricular myocytes induces physiological changes in calcium handling," *Cardiovascular Research*, vol. 49, no. 2, pp. 288–297, 2001.
- [25] L. Pohlmann, I. Kröger, N. Vignier, et al., "Cardiac myosin-binding protein C is required for complete relaxation in intact myocytes," *Circulation Research*, vol. 101, no. 9, pp. 928–938, 2007.
- [26] A. Rinne, C. Littwitz, M.-C. Kienitz, et al., "Gene silencing in adult rat cardiac myocytes in vitro by adenovirus-mediated RNA interference," *Journal of Muscle Research and Cell Motility*, vol. 27, no. 5–7, pp. 413–421, 2006.
- [27] H. Xu, W. Guo, and J. M. Nerbonne, "Four kinetically distinct depolarization-activated K<sup>+</sup> currents in adult mouse ventricular myocytes," *Journal of General Physiology*, vol. 113, no. 5, pp. 661–677, 1999.
- [28] H. M. Himmel, E. Wettwer, Q. Li, and U. Ravens, "Four different components contribute to outward current in rat ventricular myocytes," *American Journal of Physiology*, vol. 277, no. 1, pp. H107–H118, 1999.
- [29] X. Liu, J. Yang, F. Shang, et al., "Silencing GIRK4 expression in human atrial myocytes by adenovirus-delivered small hairpin RNA," *Molecular Biology Reports*, pp. 1–8, 2008.
- [30] P. Lipp, J. Hüser, L. Pott, and E. Niggli, "Spatially non-uniform Ca<sup>2+</sup> signals induced by the reduction of transverse tubules in citrate-loaded guinea-pig ventricular myocytes in culture," *Journal of Physiology*, vol. 497, no. 3, pp. 589–597, 1996.
- [31] C. Viero, U. Kraushaar, S. Ruppenthal, L. Kaestner, and P. Lipp, "A primary culture system for sustained expression of a calcium sensor in preserved adult rat ventricular myocytes," *Cell Calcium*, vol. 43, no. 1, pp. 59–71, 2008.
- [32] N. Jost, L. Virág, M. Bitay, et al., "Restricting excessive cardiac action potential and QT prolongation: a vital role for IKs in human ventricular muscle," *Circulation*, vol. 112, no. 10, pp. 1392–1399, 2005.
- [33] S. Radicke, D. Cotella, E. M. Graf, et al., "Functional modulation of the transient outward current I<sub>to</sub> by KCNE β-subunits and regional distribution in human non-failing and failing hearts," *Cardiovascular Research*, vol. 71, no. 4, pp. 695–703, 2006.
- [34] G. Grynkiewicz, M. Poenie, and R. Y. Tsien, "A new generation of Ca<sup>2+</sup> indicators with greatly improved fluorescence properties," *Journal of Biological Chemistry*, vol. 260, no. 6, pp. 3440–3450, 1985.
- [35] H. J. Knot, I. Laher, E. A. Sobie, et al., "Twenty years of calcium imaging: cell physiology to dye for," *Molecular Interventions*, vol. 5, no. 2, pp. 112–127, 2005.
- [36] A. E. Palmer and R. Y. Tsien, "Measuring calcium signaling using genetically targetable fluorescent indicators," *Nature Protocols*, vol. 1, no. 3, pp. 1057–1065, 2006.
- [37] J. E. McCombs and A. E. Palmer, "Measuring calcium dynamics in living cells with genetically encodable calcium indicators," *Methods*, vol. 46, no. 3, pp. 152–159, 2008.
- [38] C. Communal, F. Huq, D. Lebeche, C. Mestel, J. K. Gwathmey, and R. J. Hajjar, "Decreased efficiency of adenovirus-mediated gene transfer in aging cardiomyocytes," *Circulation*, vol. 107, no. 8, pp. 1170–1175, 2003.
- [39] Z. Li, R. V. Sharma, D. Duan, and R. L. Davisson, "Adenovirus-mediated gene transfer to adult mouse cardiomyocytes is selectively influenced by culture medium," *Journal of Gene Medicine*, vol. 5, no. 9, pp. 765–772, 2003.
- [40] C. Communal, M. Sumandea, P. de Tombe, J. Narula, R. J. Solaro, and R. J. Hajjar, "Functional consequences of caspase activation in cardiac myocytes," *Proceedings of the National Academy of Sciences of the United States of America*, vol. 99, no. 9, pp. 6252–6256, 2002.
- [41] M. Ahmed, M. Lock, C. G. Miller, and N. W. Fraser, "Regions of the herpes simplex virus type 1 latency-associated transcript that protect cells from apoptosis in vitro and protect neuronal cells in vivo," *Journal of Virology*, vol. 76, no. 2, pp. 717–729, 2002.
- [42] M. Inman, G.-C. Perng, G. Henderson, et al., "Region of herpes simplex virus type 1 latency-associated transcript sufficient for wild-type spontaneous reactivation promotes cell survival in tissue culture," *Journal of Virology*, vol. 75, no. 8, pp. 3636–3646, 2001.
- [43] X. Chen, X. Zhang, H. Kubo, et al., "Ca<sup>2+</sup> influx-induced sarcoplasmic reticulum Ca<sup>2+</sup> overload causes mitochondrial-dependent apoptosis in ventricular myocytes," *Circulation Research*, vol. 97, no. 10, pp. 1009–1017, 2005.

## Special Issue on Transient Protein-Protein Complexes

### Call for Papers

Formation of transient macromolecular complexes between two or more biological molecules—including proteins, nucleic acids, carbohydrates, as well as small-molecule ligands—is a universal theme underpinning all living processes. While X-ray crystallography and nuclear magnetic resonance (NMR) have revealed a great deal of information about macromolecular complexes at atomic resolution, much remains to be learned. It often happens that the formation of transient complexes is too weak and too unstable to be detected and visualized by established methods. Consequently, the structural information on macromolecular interactions does not grow at the same pace as that of individual components, which constitutes an outstanding problem in molecular biology. Furthermore, formation of macromolecular complexes can be modulated by posttranslational modifications and by the presence of other binding partners. It has also been evidenced that macromolecules can form complexes other than the stereospecific one. The formation of such nonspecific complexes may facilitate macromolecular recognition, but may also lead to inappropriate utilization of biochemical pathways in disease conditions.

We invite authors to present original research as well as review articles that will stimulate the continuing efforts in understanding transient macromolecular complexes. We are particularly interested in manuscripts that report new methods for detecting, analyzing, and visualizing transient protein-protein interactions *in vivo*, *in vitro* or *in silico*, as well as original findings about characterizing such complexes. Reviews that summarize the latest development in the topic area are also welcome. The topics to be considered include, but are not limited to:

- Novel experimental approaches, such as NMR, small-angle neutron, X-ray scattering (SANS and SAXS), and fluorescence (either bulk or single molecule), to detect and visualize transient protein-protein interactions
- Kinetics studies of association and dissociation of transient protein-protein complexes
- Computational methods to predict and depict transient protein-protein complexes

- Native and aberrant formation of protein-protein complexes in essential biological processes such as signal transduction, transcription/translation, and electron transfer
- Theoretical account for the mechanism of transient protein-protein association and its evolutionary origin

Before submission authors should carefully read over the journal's Author Guidelines, which are located at <http://www.hindawi.com/journals/jbb/guidelines.html>. Prospective authors should submit an electronic copy of their complete manuscript through the journal Manuscript Tracking System at <http://mts.hindawi.com/> according to the following timetable:

Manuscript Due	December 1, 2009
First Round of Reviews	March 1, 2010
Publication Date	June 1, 2010

### Lead Guest Editor

**Chun Tang**, Department of Biochemistry, Department of Physics & Informatics Institute, University of Missouri, Columbia, MO 65211-5100, USA; [tangch@missouri.edu](mailto:tangch@missouri.edu)

### Guest Editor

**Juan Fernández-Recio**, Life Sciences Department, Barcelona Supercomputing Center, Jordi Girona 31, 08034 Barcelona, Spain; [juanf@bsc.es](mailto:juanf@bsc.es)

## Special Issue on Advances in Muscle Physiology and Pathophysiology

### Call for Papers

A multitude of important advances have occurred in the field of muscle research during the last decade, concerning striated muscle physiology and pathophysiology. The structure and function of old and new components of the sarcomeric and extrasarcomeric cytoskeleton, the sarcolemma and internal membranes, the extracellular matrix, and the neuromuscular junction have been unraveled using cutting-edge molecular, cellular, biochemical, and biophysical tools combined with high-throughput genomic and proteomic approaches. A number of animal models, ranging from *Drosophila* and *C. elegans* to zebrafish and mouse, have been developed, which in some instances confirmed our predictions about the activities of certain genes, but in others raised questions about their roles. Such studies further underscored the complexity of the muscle cell, and pinpointed the need for development of highly sophisticated and integrated approaches, which will ultimately provide us with more accurate answers.

In this volume, we invite authors to present original research articles as well as review articles that will stimulate the continuing efforts to unravel the intricate web of the skeletal and cardiac muscle cell and their properties in normalcy and disease.

Potential topics include but are not limited to:

- Assembly of the muscle cell; the sarcomeric and extrasarcomeric cytoskeleton
- Sarcolemma, extracellular matrix, and adhesion
- Internal membranes and excitation/contraction coupling
- Muscle biomechanics at organismal, cellular, and single-molecule levels
- Metabolism and energetics
- Models of muscular dystrophy
- Stem cells and muscle regeneration
- Apoptosis and  $Ca^{2+}$

Before submission authors should carefully read over the journal's Author Guidelines, which are located at <http://www.hindawi.com/journals/jbb/guidelines.html>. Prospective authors should submit an electronic copy of their complete

manuscript through the journal Manuscript Tracking System at <http://mts.hindawi.com/> according to the following timetable:

Manuscript Due	November 1, 2009
First Round of Reviews	February 1, 2010
Publication Date	May 1, 2010

### Lead Guest Editor

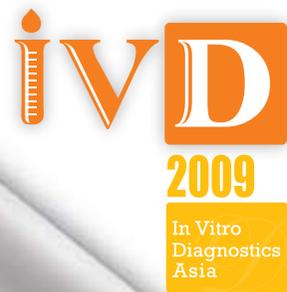
**Aikaterini Kontrogianni-Konstantopoulos**, Department of Biochemistry and Molecular Biology, Baltimore School of Medicine, University of Maryland, Baltimore, MD, USA; [akons001@umaryland.edu](mailto:akons001@umaryland.edu)

### Guest Editors

**Guy Benian**, Departments of Pathology and Cell Biology, Emory University, Atlanta, GA, USA; [pathgb@emory.edu](mailto:pathgb@emory.edu)

**Henk Granzier**, Departments of Physiology and Molecular and Cellular Biology, University of Arizona, Tucson, AZ, USA; [granzier@email.arizona.edu](mailto:granzier@email.arizona.edu)

# In Vitro Diagnostics Asia 2009



Embracing the cutting-edge discoveries and innovative technologies in clinical diagnostics to revolutionize the healthcare arena

[www.ivdindia.com](http://www.ivdindia.com)

Crowne Plaza Hotel, New Delhi, India  
[www.crowneplazadelhi.com](http://www.crowneplazadelhi.com)  
19-20 August 2009

Due to overwhelming responses we received from our last two annual conferences on IVD China. JFPS is proud to announce the 3rd In Vitro Diagnostics conference in Asia. From our past events, we have successfully drawn in more than 400 international leaders and investors to our IVD event. Therefore, this event will be an excellent platform for networking and future dealings.

Organizer

**JFPS**  
WE EMPOWER BUSINESS  
[www.jfpsgroup.com](http://www.jfpsgroup.com)