

Excitotoxic mechanisms in non-motor dysfunctions and levodopa-induced dyskinesia in Parkinson's disease: the role of the interaction between dopaminergic and the kynurenine system

Journal:	Current Medicinal Chemistry	
Manuscript ID	CMC-2015-0247.R1	
Manuscript Type:	Review	
Date Submitted by the Author:	n/a	
Complete List of Authors:	Majláth, Zsófia; University of Szeged, Department of Neurology Toldi, József; University of Szeged, Department of Physiology, Anatomy and Neuroscience; MTA-SZTE Neuroscience Research Group of the Hungarian Academy of Sciences and University of Szeged Fülöp, Ferenc; Department of Pharmaceutical Chemistry, Faculty of Pharmacy, University of Szeged Vécsei, László; University of Szeged, Department of Neurology; MTA-SZTE Neuroscience Research Group of the Hungarian Academy of Sciences and University of Szeged	
Keywords:	Parkinson's disease, kynurenic acid, glutamate, neuroprotection, NMDA receptor, levodopa-induced dyskinesia, excitotoxicity	
Note: The following files were submitted by the author for peer review, but cannot be converted to PDF. You must view these files (e.g. movies) online.		
Schemes.cdx		

SCHOLARONE[™] Manuscripts

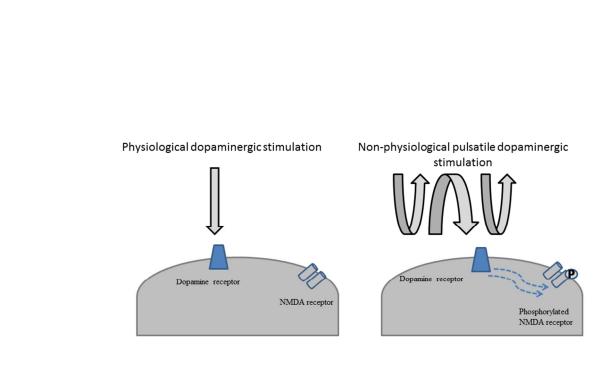
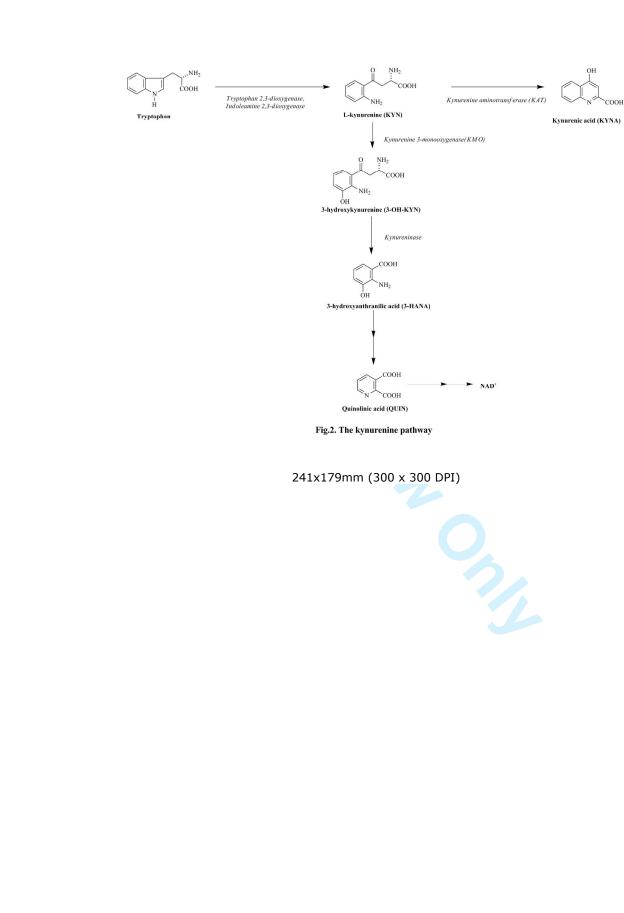
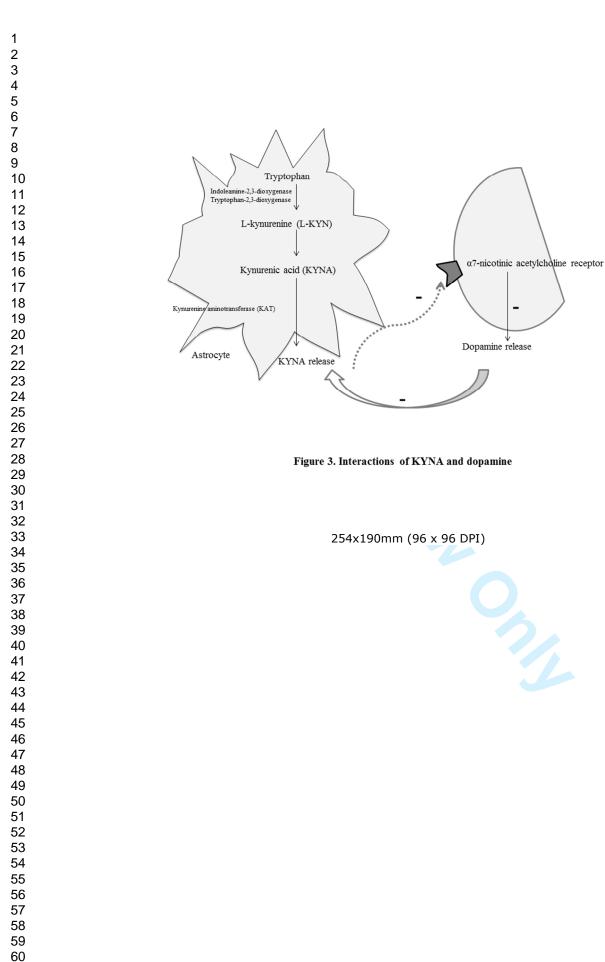


Figure 1. Alterations of the glutamatergic neurotransmission in PD

254x190mm (96 x 96 DPI)





Excitotoxic mechanisms in non-motor dysfunctions and levodopa-induced dyskinesia in Parkinson's disease: the role of the interaction between the dopaminergic and the kynurenine system

Abstract: Parkinson's disease is a common progressing neurodegenerative disorder presenting with characteristic motor symptoms. Non-motor dysfunctions and therapy-related complications frequently develop, but are often underdiagnosed and undertreated. Levodopa-induced dyskinesia and impulse control disorders are suggested to share pathophysiological processes and be related to alterations of the glutamatergic neurotransmission. Anti-glutamatergic interventions are therefore worth considering: several lines of evidence already indicate their beneficial effect. The kynurenine pathway offers the endogenous glutamate receptor antagonist kynurenic acid, which may provide a promising candidate for future drug development with the aim of assessment of the motor symptoms and therapy-related complications of Parkinson's disease.

Keywords: Parkinson's disease, kynurenic acid, neuroprotection, NMDA receptor, glutamate, levodopa-induced dyskinesia, excitotoxicity

Introduction

Parkinson's disease (PD) is a chronic progressive neurodegenerative disorder presenting with characteristic motor symptoms: rigidity, resting tremor, bradykinesia and postural instability ¹. Epidemiological data indicate that the overall prevalence of PD is around 1.5% with no gender differences; the prevalence increases with the age ^{2, 3}. The neuropathological background of PD is the progressive selective degeneration of dopaminergic neurons in the substantia nigra ⁴. However, as our understanding of the pathophysiological process of PD has improved, it has become evident that, besides the dopaminergic system, the cholinergic, serotoninergic, noradrenergic and glutamatergic systems are also affected. Glutamate is the main excitatory neurotransmitter in the human brain, but the excessive stimulation of glutamatergic receptors may lead to neuronal damage, a process known as excitotoxicity.

Excitotoxicity has been implicated in the pathological mechanisms of several neurodegenerative disorders, including PD $^{5, 6}$.

The pathomechanism of PD has still not yet been fully clarified despite extensive research, but genetic and environmental factors have both been suggested to contribute. Most PD cases are sporadic, but a minority of the patients have a clear familial disease. The role of hereditary factors has been strengthened by epidemiological studies indicating a significantly higher risk of developing the disease among first-degree relatives of PD patients ⁷⁻⁹. Genetic investigations in families with inherited forms of PD have identified several gene mutations; these data also promoted the understanding of the pathogenesis of the disorder ^{10, 11}. Later, studies suggested that genetic factors may contribute to a higher risk of PD in the general population as well, and environmental factors combined with a genetic predisposition may result together in the development of PD ¹².

The gold standard of PD therapy currently remains dopamine (DA) replacement with oral levodopa, which offers good symptomatic relief for the motor symptoms, but can be associated with therapy-related complications such as levodopa-induced dyskinesia (LID) or impulse control disorders (ICDs). Non-motor symptoms (NMs) of PD are often unrecognized and undertreated, although they may have a serious impact on the quality of life. They often precede the development of motor symptoms and their frequency increases in parallel with the progression of PD ^{13, 14}. NMs can be classified into four main groups: autonomic symptoms, sensory symptoms, sleep disorders and neuropsychiatric symptoms (Table 1.).

• · • •	
Autonomic symptoms	orthostatic hypotension
	gastrointestinal dysfunctions
	nausea and vomiting
	sexual dysfunctions
	bladder dysfunction
Sensory disturbances	anosmia
	paraesthesias
	pain
	taste deficits
Sleep disorders	rapid eye movement disorder
	restless leg syndrome
	excessive daytime sleepiness
	vivid dreaming
	insomnia
Neuropsychiatric disorders	psychosis
	depression

https://mc04.manuscriptcentral.com/crmc

cognitive impairment or dementia anxiety dysphoria impulse control disorders

ICDs include pathological gambling, hypersexuality, compulsive shopping and binge eating. Gambling and hypersexuality show a male predominance, while compulsive shopping and eating are more frequent among female PD patients^{15, 16}. The development of ICDs is associated with the use of DA agonists¹⁷. ICDs are common complications of PD, although they are often underdiagnosed. Epidemiological data indicate the prevalence of ICDs as being between 6 and 18% of PD patients^{16, 18, 19}. There are several risk factors which have been associated with the development of ICDs: a younger age, DA agonists or a higher levodopa dosage, a family history of gambling or alcohol problems, impulsivity and depression²⁰. The exact pathological mechanisms of ICDs are not yet fully clarified.

The role of interactions of the dopaminergic and glutamatergic systems in the development of therapy-related complications of PD

It has recently been suggested that the pathophysiological background of LID and ICDs is common, and involves alterations of the glutamatergic neurotransmission²⁰⁻²². Presynaptic changes affecting DA release and postsynaptic changes affecting DA receptors have been described in LID, and are hypothesized to contribute to the development of ICDs too. DA exerts its effect mainly on the medium spiny neurons of the caudate and putamen, which are rich in glutamate receptors. Interactions of DA and glutamate receptors play an important role in the normal function of the basal ganglia, and this fine balance is disrupted in PD ²³⁻²⁵. DA replacement therapy results in a chronic intermittent, non-physiological stimulation of DA receptors instead of the physiological tonic stimulation, which leads to changes in the glutamate receptors on the striatal spiny neurons. Experimental data from both animal models and humans indicate that the phosphorylation of glutamate receptors undergoes a change. **Phosphorylation of the NMDA receptors is an important mechanism in the regulation of NMDA receptor function. Several kinases have been identified which are able to phosphorylate ionotropic glutamate receptors, including protein kinase A, protein kinase C, Ca(2+)/calmodulin-dependent protein kinase II, Src/Fyn non-receptor tyrosine**

kinases, and cyclin dependent kinase-5²⁶. In PD, different signal transduction cascades - the cyclic adenosine monophosphate (cAMP)-protein kinase A-mediated and the calciumcalmodulin-dependent kinase II pathways - are activated, consequently the subunit composition changes and NMDA receptors become phosphorylated ^{27, 28}. Serin phosphorylation of the NR1 subunit and tyrosine phosphorylation of the NR2 subunit of the NMDA receptors have been described after long-term dopaminergic treatment ²⁹. The hyperphosphorylation of the NMDA receptors leads to long-term potentiation of their sensitivity ³⁰ (Fig.1.). Another important kinase involved in the phosphorylation of NMDA receptors and the mediation of dopamine-dependent signaling processes is glycogen synthase kinase 3-beta (GSK3 β). Activation of GSK3 β has been proved to inhibit presynaptic glutamate release, alter the expression of NMDA receptors and under hyperdopaminergic conditions, affect synaptic NMDA-mediated currents³¹⁻³³. GSK3 β has been described to be involved in several pathogenic processes of PD, including oxidative stress, protein aggregation and neuroinflammation³⁴.

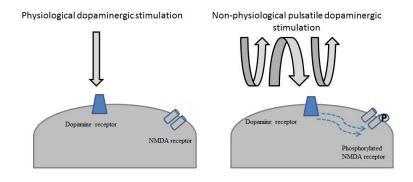


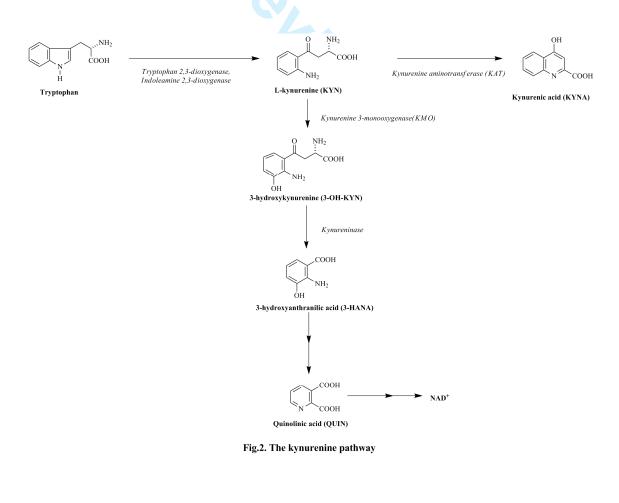
Figure 1. Alterations of the glutamatergic neurotransmission in PD

These alterations result in hypersensitivity towards DA and the development of motor fluctuations. The possible role of the connection of DA and glutamate receptors in the pathogenesis of ICDs has been strengthened by a genetic study which found an association of the GRIN2B gene with ICDs ³⁵. GRIN2B is responsible for the synthesis of the NR2B subunit of NMDA receptors, which is the predominant form of the striatal NMDA receptors.

The kynurenine pathway and its interaction with the dopaminergic system

The kynurenine pathway (KP) is the main route of the tryptophan (Trp) metabolism, responsible for more than 95% of Trp degradation (Fig. 2.) ^{36, 37}. **It has been recognized** a**lready in the 1980s that the** enzymatic cascade of the KP generates several neuroactive compounds^{38, 39}. The first, rate-limiting step of the KP is the enzymatic degradation of Trp, this step catalyzed by indoleamine-2,3-dioxygenase (IDO). The KP divides into two main branches at a central intermediate, L-kynurenine (KYN). The first arm of the pathway is the conversion of KYN into kynurenic acid (KYNA). KYNA synthesis can be attributed to the action of kynurenine-aminotransferases (KATs), so far 4 subtypes of this enzyme are known, which have slightly different biochemical profile ⁴⁰.

The other branch of the KP begins with the conversion osf KYN by kynurenine-3monooxygenase (KMO) into 3-hydroxy-kynurenine. This is further metabolized by multiple enzymatic steps, gives rise to the neurotoxic quinolinic acid (QUIN) and finally ends at the production of the essential coenzyme nicotinamide adenine dinucleotide (NAD).



KYNA is a broad-spectrum endogenous inhibitor of ionotropic excitatory amino acid receptors. It has the highest affinity for the N-methyl-D-aspartate (NMDA) type glutamate receptors, exerting its effect mainly by binding to the strychnine-insensitive glycine-binding site, but at a higher concentration it can also bind to the glutamate-binding site ^{41, 42}. On another type of glutamate receptors, the α -amino-3-hydroxy-5-methyl-4-isoxazolepropionic acid (AMPA) receptors, KYNA exerts a concentration-dependent dual effect: in a higher, micromolar concentration it is an inhibitor, while in a nanomolar concentration it acts as a facilitator ^{43, 44}. KYNA has also been described to be a non-competitive inhibitor of the presynaptic a7 nicotinic acetylcholine receptors thereby reducing presynaptic glutamate release ^{45, 46}. Later investigations could not confirm this inhibitory potential ⁴⁷. However, KAT-deletion and consequently a loss of KYNA resulted in an increased sensitivity of α7 nicotinic acetylcholine receptors, suggesting a regulatory role of KYNA on them⁴⁸. Currently, the possible role of KYNA on α 7 nicotinic acetylcholine receptors is not clearly understood and further research is needed to describe any potential effect ⁴⁹. It was subsequently confirmed that KYNA is an agonist of the previously orphan G-protein coupled receptor GPR35, which has also been suggested to contribute to the reduction of

coupled receptor GPR35, which has also been suggested to contribute to the reduction of extracellular glutamate in the brain ^{50, 51}. The complex modes of actions suggest that KYNA has an important neuromodulatory role in the central nervous system and it is involved in the regulation of glutamatergic and cholinergic neurotransmission (Table 2.).

Table 2. Receptor targets of KYNA and its effects on them	
NMDA receptors	antagonist
AMPA receptors	facilitation in nanomolar, inhibition in micromolar concentration
a7-nicotinic acetylcholine receptors	antagonist
GPR35 receptor	agonist

Increasing evidence confirms the neuroprotective capacity of KYNA in different conditions, such as QUIN, NMDA and glutamate-induced toxicity or ischaemia ⁵²⁻⁵⁵. On the other hand, QUIN is a neurotoxic compound which exerts its deteriorating effect mainly via NMDA agonism, but also contributes to the generation of free radicals, promotes lipid peroxidation and decreases antioxidant capacity⁵⁶⁻⁶⁰. Interestingly, in contrast with KYNA, QUIN enhances glutamate release ⁶¹.

An important aspect regarding the effects of KYNA is its close connection with the dopaminergic system. Elevations of KYNA levels result in the inhibition of dopamine release. This effect is suggested to be mediated predominantly by the α 7-nicotinic acetylcholine receptors, but at higher concentrations this effect may be due to the inhibition of glutamate receptors ^{62, 63}. Conversely, levodopa substitution has also been confirmed to reduce KYNA levels ⁶⁴. (Fig. 3.)

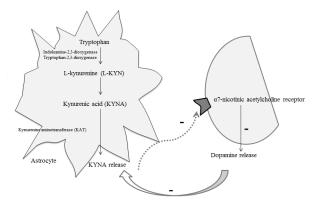


Figure 3. Interactions of KYNA and dopamine

These results further strengthen the concept that KYNA has an important role in the regulation of glutamatergic, cholinergic and dopaminergic neurotransmission, which may also have important implications for the pathophysiology of PD. Alterations of KYNA are able to modulate DA levels, while reduced KYNA levels may also increase vulnerability towards excitotoxic effects by enhancing glutamatergic influences.⁶⁵ Alterations in the delicate balance of neuroactive kynurenines have been confirmed in several neurodegenerative diseases, including Alzheimer's disease, multiple sclerosis and Huntington's disease ⁶⁶⁻⁶⁹. KP changes have also been described in PD. Kynurenine/Trp ratios were increased, while serum Trp levels were decreased in PD patients as compared with healthy controls ⁷⁰. Ogawa et al. have measured reduced concentrations of KYN and KYNA in several brain regions of PD patients, including the frontal cortex, putamen and substantia nigra ⁷¹. They also confirmed a significantly increased 3-OH-KYN levels in the putamen and substantia nigra of PD brains; this compound may contribute to the oxidative stress and consequently to the neuronal damage ⁷¹. KP alterations have also been measured at the periphery: in the plasma of PD patients, the KYNA level and the KAT activity have been found to be decreased ⁷².

Therapeutic opportunities by glutamate antagonism

Memantine, a weak NMDA antagonist, has been revealed to have beneficial symptomatic effects for dementia associated with PD and, interestingly, it may additionally improve motor symptoms ⁷³⁻⁷⁶. Case reports suggested that memantine may also improve LID ^{77, 78}. So far there is no data concerning its therapeutic effect on non-motor symptoms or ICDs.

Another anti-glutamatergic agent, amantadine has been confirmed to improve LID symptoms both as acute and as chronic therapy ⁷⁹⁻⁸¹. It was proved that the beneficial effect of amantadine is long-lasting, and is sustained even after 1 year of continuous therapy ⁸². The beneficial effect of amantadine on LID is suggested to be modulated at least in part by NMDA inhibition ⁸³. Amantadine has been already tested for the therapeutic management of ICDs with promising results. A small study has revealed that amantadine is capable of reducing pathological gambling in PD patients ⁶. The effect of amantadine was recently confirmed by another small-scale study, which described the behavioral background of this effect ⁸⁴. The possible therapeutic effect of amantadine was strengthened by other studies involving patients with pathological gambling without PD ⁸⁵. Case reports have described the beneficial effect of amantadine on punding as well, which is another complication of dopamine-replacement therapy ^{86, 87}.

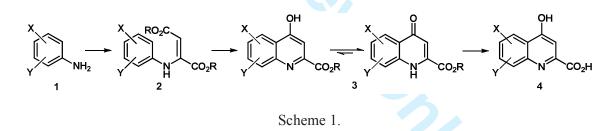
These results suggest that anti-glutamatergic therapies may be valuable options for the management of not only the motor symptoms, but also of the therapy-related complications of PD. KYNA is the only known endogenous NMDA antagonist, which has been suggested to provide symptomatic treatment and also neuroprotection^{88, 89}. In an experimental PD model, KYNA was able to alleviate parkinsonian motor symptoms⁸⁹. There is growing evidence indicating that the hypersensitivity of NMDA receptors plays an important role in the development of LID and other complications of PD, and the NMDA antagonist KYNA may therefore be beneficial for the treatment of these complications too. Elevation of the brain KYNA levels has been already examined in experimental PD models. KYNA itself can cross the blood-brain barrier poorly, but the peripheral administration of its precursor, KYN, together with the organic acid transport inhibitor probenecid, results in significant brain KYNA content elevations. This treatment proved to be neuroprotective in a 6hydroxydopamine model of PD⁸⁸. Another possibility is the synthesis of kynurenine derivatives, in another animal model of PD, the neuroprotective effects of several synthetic kynurenines were confirmed ⁹⁰. Another option to achieve a KYNA elevation would be to influence the enzymatic machinery of the KP; accordingly, KMO inhibition results in a shift

towards the production of KYNA. This method resulted in the improvement of LID, while the antiparkinsonian effect of levodopa was not diminished ^{91,92}.

There are not yet data regarding the effects of KYNA on the non-motor complications of PD, but the promising results with amantadine, and the fact that KYNA is able to improve LID, suggest that KYNA may be of therapeutic value for these conditions too. Further investigations are worth considering to assess the potential therapeutic benefits of KYNA in PD. From this aspect it is of interest to produce synthethic KNYA derivatives which may have more favourable pharmacokinetic properties than those of KYNA, such as an improved receptor selectivity or an improved blood-brain barrier penetration. Several synthethic KYNA analogues have already been developed to achieve neuroprotection ⁹³.

Syntheses of kynurenic acid derivatives

The general procedure for the synthesis of 4-hydroxyquinolinic acid can be achieved by a modified Conrad-Limpach method starting from the commercially available substituted aryl amines $1^{93, 94}$. The first step involves enamine bond formation by using dimethyl acetylenedicarboxylate or diethyl acetylenedicarboxylate resulting in **2** (Scheme 1).



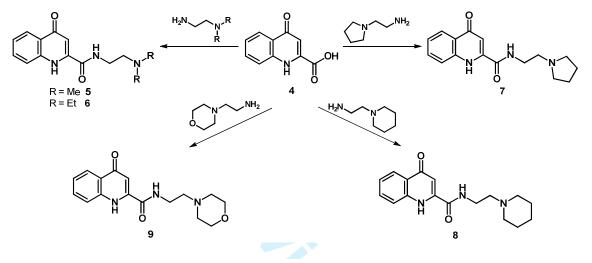
The intermediate fumarate **2** is then cyclized at high temperature, yielding 4-hydroxy-2carboxylates **3** respectively. For 3 the enol-oxo tautomerism is possible. This equilibrium is shifted to the oxo form when C-2 contains an ester function, while the presence of a carboxylic group at C-2 indicate the enol form (Scheme 1). For the transformations of the 4hydroxy-quinoline derivatives, the free acids (KYNA derivatives) are needed. The hydrolysis of the esters is generally performed in methanolic alkali media, followed by acidification with HCl, resulting in kynurenic acid derivatives **4** (Scheme 1).

The transformations of KYNA derivatives can be achieved in different ways: by the transformation of the synthetically active hydroxy group at position 4 or by the conversion of the carboxylic function at position 2^{93} .

Current Medicinal Chemistry

The aim in the last few years has been to introduce a cationic centre in the side-chain so as to have good water solubility and better penetration ability of the products trough the blood-brain barrier ⁹⁴.

The coupling between KYNA and 2-dimethylaminoethylamine was first achieved by using N,N'-diisopropylcarbodiimide in the presence of 1-hydroxybenzotriazole hydrate, yielding **5** (Scheme 2).





The excellent biological activity of **5** led us to prepare a number of analogues $^{93, 94}$. Several of them is shown in Scheme 2. For example, by using 2-diethylaminoethylamine as starting amine, **6** was synthesized as a diethyl analogue of **2** (Scheme 2), and analogues **7**, **8** and **9**, containing the tertiary nitrogen in different ring systems, were prepared by reacting KYNA with 2-pyrrolidinoethylamine, 2-piperidinoethylamine and 2-morpholinoethylamine, respectively.

The synthethic KYNA analogues have already been tested in several animal models of neurological diseases with promising results ^{95, 96}. In an animal model of transient global forebrain ischaemia, KYNA exerted a neuroprotective effect by reducing the hippocampal CA1 pyramidal cell loss. The analogue was effective both as a pre-treatment and as post-treatment ⁹⁶. In a transgenic mice model of Huntington's disease, the same analogue proved not only to ameliorate the motor symptoms but it also significantly, by more than 30%, prolonged the survival of the animals. The neuroprotective effect was also confirmed histologically: the KYNA analogue was able to

prevent neuronal atrophy in the striatum of the transgenic animals ⁹⁵. As KYNA is an NMDA antagonist, an important concern would be the possible interference with the physiological glutamatergic functions. However, behavioral studies with this novel KYNA analogue confirmed, that in the same dose in which it proved to be neuroprotective, it did not induce any cognitive side effects⁹⁷.

Conclusion

The therapeutic management of PD through dopamine replacement is associated with therapyrelated complications, which may have a more severe impact on the quality of life of patients than the parkinsonian motor symptoms. Alterations of the glutamatergic neurotransmission are implicated in the development of both LID and ICDs, and antiglutamatergic interventions may therefore be beneficial for these complications. KYNA as an NMDA antagonist compound, which is also able to influence dopaminergic neurotransmission may be a promising candidate for future drug development.

Acknowledgements

Thanks are due to David Durham for the linguistic corrections of the manuscript.

Conflict of Interest

The authors have no conflict of interest to report.

References

[1] Goetz, C. G. Charcot on Parkinson's disease. Mov Disord, 1986, 1 (1), 27-32. [2] de Rijk, M. C.; Breteler, M. M.; Graveland, G. A.; Ott, A.; Grobbee, D. E.; van der Meche, F. G.; Hofman, A. Prevalence of Parkinson's disease in the elderly: the Rotterdam Study. Neurology, 1995, 45 (12), 2143-6. [3] de Rijk, M. C.; Tzourio, C.; Breteler, M. M.; Dartigues, J. F.; Amaducci, L.; Lopez-Pousa, S.; Manubens-Bertran, J. M.; Alperovitch, A.; Rocca, W. A. Prevalence of parkinsonism and Parkinson's disease in Europe: the EUROPARKINSON Collaborative Study. European Community Concerted Action on the Epidemiology of Parkinson's disease. J Neurol Neurosurg Psychiatry, 1997, 62 (1), 10-5. [4] Gibb, W. R.; Lees, A. J. Anatomy, pigmentation, ventral and dorsal subpopulations of the substantia nigra, and differential cell death in Parkinson's disease. J Neurol Neurosurg *Psychiatry*, **1991**, *54* (5), 388-96. [5] Blandini, F. An update on the potential role of excitotoxicity in the pathogenesis of Parkinson's disease. Funct Neurol, 2010, 25 (2), 65-71. [6] Obal, I.; Majlath, Z.; Toldi, J.; Vecsei, L. Mental disturbances in Parkinson's disease and related disorders: the role of excitotoxins. J Parkinsons Dis, 2014, 4 (2), 139-50. [7] Payami, H.; Larsen, K.; Bernard, S.; Nutt, J. Increased risk of Parkinson's disease in parents and siblings of patients. Ann Neurology, 1994, 36 (4), 659-61. [8] Vieregge, P.; Heberlein, I. Increased risk of Parkinson's disease in relatives of patients. Annals of neurology, 1995, 37 (5), 685. [9] Hattori, N.; Shimura, H.; Kubo, S.; Wang, M.; Shimizu, N.; Tanaka, K.; Mizuno, Y. Importance of familial Parkinson's disease and parkinsonism to the understanding of nigral degeneration in sporadic Parkinson's disease. J Neural Transm Suppl. 2000, (60), 101-16. [10] Lev, N.; Melamed, E. Heredity in Parkinson's disease: new findings. IMAJ, 2001, 3 (6), 435-8. [11] Gasser, T. Genetics of Parkinson's disease. J Neurology, 2001, 248 (10), 833-40. [12] Cheon, S. M.; Chan, L.; Chan, D. K.; Kim, J. W. Genetics of Parkinson's disease - a clinical perspective. J Mov Disord, 2012, 5 (2), 33-41. [13] Barone, P.; Antonini, A.; Colosimo, C.; Marconi, R.; Morgante, L.; Avarello, T. P.; Bottacchi, E.; Cannas, A.; Ceravolo, G.; Ceravolo, R.; Cicarelli, G.; Gaglio, R. M.; Giglia, R. M.; Iemolo, F.; Manfredi, M.; Meco, G.; Nicoletti, A.; Pederzoli, M.; Petrone, A.; Pisani, A.; Pontieri, F. E.; Quatrale, R.; Ramat, S.; Scala, R.; Volpe, G.; Zappulla, S.; Bentivoglio, A. R.; Stocchi, F.; Trianni, G.; Dotto, P. D. The PRIAMO study: A multicenter assessment of nonmotor symptoms and their impact on quality of life in Parkinson's disease. Mov Disord, 2009, 24 (11), 1641-9. [14] Chaudhuri, K. R.; Odin, P.; Antonini, A.; Martinez-Martin, P. Parkinson's disease: the non-motor issues. Parkinsonism Relat Disord, 2011, 17 (10), 717-23. [15] Voon, V.; Hassan, K.; Zurowski, M.; de Souza, M.; Thomsen, T.; Fox, S.; Lang, A. E.; Miyasaki, J. Prevalence of repetitive and reward-seeking behaviors in Parkinson disease. Neurology, 2006, 67 (7), 1254-7. [16] Weintraub, D.; Koester, J.; Potenza, M. N.; Siderowf, A. D.; Stacy, M.; Voon, V.; Whetteckey, J.; Wunderlich, G. R.; Lang, A. E. Impulse control disorders in Parkinson disease: a cross-sectional study of 3090 patients. Arch Neurol, 2010, 67 (5), 589-95. [17] Weintraub, D.; Siderowf, A. D.; Potenza, M. N.; Goveas, J.; Morales, K. H.; Duda, J. E.; Moberg, P. J.; Stern, M. B. Association of dopamine agonist use with impulse control disorders in Parkinson disease. Arch Neurol, 2006, 63 (7), 969-73.

40

41

42

43

44

45

46

47

48 49

50

51

52

53

54

55

56

57

58 59

2	[18] Poletti, M.; Logi, C.; Lucetti, C.; Del Dotto, P.; Baldacci, F.; Vergallo, A.; Ulivi, M.; Del Sarto, S.; Rossi, G.; Ceravolo, R.; Bonuccelli, U. A single-center, cross-sectional prevalence study of impulse control disorders in Parkinson disease: association with dopaminergic drugs. <i>J Clin Psychopharmacol</i> , 2013 , <i>33</i> (5), 691-4.
1	19] Lee, J. Y.; Kim, J. M.; Kim, J. W.; Cho, J.; Lee, W. Y.; Kim, H. J.; Jeon, B. S. Association between the dose of dopaminergic medication and the behavioral disturbances in Parkinson disease. <i>Parkinsonism Relat Disord</i> , 2010 , <i>16</i> (3), 202-7.
[] (20] Voon, V.; Fernagut, P. O.; Wickens, J.; Baunez, C.; Rodriguez, M.; Pavon, N.; Juncos, J. L.; Obeso, J. A.; Bezard, E. Chronic dopaminergic stimulation in Parkinson's disease: from dyskinesias to impulse control disorders. <i>Lancet Neurol</i> , 2009 , <i>8</i> (12), 1140-9.
_	21] Kalivas, P. W. The glutamate homeostasis hypothesis of addiction. <i>Nat Rev Neurosci</i> , 2009 , <i>10</i> (8), 561-72.
[]	22] Linazasoro, G. Dopamine dysregulation syndrome and levodopa-induced dyskinesias in Parkinson disease: common consequences of anomalous forms of neural plasticity. <i>Clin Neuropharmacol</i> , 2009 , <i>32</i> (1), 22-7.
[[23] Kotter, R. Postsynaptic integration of glutamatergic and dopaminergic signals in the striatum. <i>Prog Neurobiol</i> , 1994 , <i>44</i> (2), 163-96.
8	24] Nash, J. E.; Brotchie, J. M. A common signaling pathway for striatal NMDA and adenosine A2a receptors: implications for the treatment of Parkinson's disease. <i>J Neurosci</i> , 2000 , <i>20</i> (20), 7782-9.
[]	25] Calabresi, P.; Giacomini, P.; Centonze, D.; Bernardi, G. Levodopa-induced dyskinesia: a bathological form of striatal synaptic plasticity? <i>Ann Neurol</i> , 2000 , <i>47</i> (4 Suppl 1), S60-8; liscussion S68-9.
Į [[26] Wang, J. Q.; Guo, M. L.; Jin, D. Z.; Xue, B.; Fibuch, E. E.; Mao, L. M. Roles of subunit phosphorylation in regulating glutamate receptor function. <i>Eur J Pharm</i>, 2014, <i>728</i>, 183-7. [27] Chase, T. N. The significance of continuous dopaminergic stimulation in the treatment of Parkinson's disease. <i>Drugs</i>, 1998, <i>55 Suppl 1</i>, 1-9.
[r [S	 [28] Oh, J. D.; Chase, T. N. Glutamate-mediated striatal dysregulation and the pathogenesis of notor response complications in Parkinson's disease. <i>Amino Acids</i>, 2002, <i>23</i> (1-3), 133-9. [29] Dunah, A. W.; Wang, Y.; Yasuda, R. P.; Kameyama, K.; Huganir, R. L.; Wolfe, B. B.; Standaert, D. G. Alterations in subunit expression, composition, and phosphorylation of striatal N-methyl-D-aspartate glutamate receptors in a rat 6-hydroxydopamine model of Parkinson's disease. <i>Mol Pharm</i>, 2000, <i>57</i> (2), 342-52.
	30] Chase, T. N. Levodopa therapy: consequences of the nonphysiologic replacement of dopamine. <i>Neurology</i> , 1998 , <i>50</i> (5 Suppl 5), S17-25.
v C	31] Zhu, L. Q.; Liu, D.; Hu, J.; Cheng, J.; Wang, S. H.; Wang, Q.; Wang, F.; Chen, J. G.; Wang, J. Z. GSK-3 beta inhibits presynaptic vesicle exocytosis by phosphorylating P/Q-type calcium channel and interrupting SNARE complex formation. J <i>Neurosci</i> , 2010 , <i>30</i> (10), 3624-33.
[2 (32] Chen, P.; Gu, Z.; Liu, W.; Yan, Z. Glycogen synthase kinase 3 regulates N-methyl-D- aspartate receptor channel trafficking and function in cortical neurons. <i>Mol Pharm</i> , 2007 , <i>72</i> (1), 40-51.
	 [33] Li, Y. C.; Gao, W. J. GSK-3beta activity and hyperdopamine-dependent behaviors. [34] Li, Y. C.; Gao, W. J. GSK-3beta activity and hyperdopamine-dependent behaviors. [34] Li, D. W.; Liu, Z. Q.; Chen, W.; Yao, M.; Li, G. R. Association of glycogen synthase cinase-3beta with Parkinson's disease (review). <i>Mol Med Rep</i>, 2014, <i>9</i> (6), 2043-50. [35] Lee, J. Y.; Lee, E. K.; Park, S. S.; Lim, J. Y.; Kim, H. J.; Kim, J. S.; Jeon, B. S. Association of DRD3 and GRIN2B with impulse control and related behaviors in Parkinson's disease. <i>Mov Disord</i>, 2009, <i>24</i> (12), 1803-10.
	13
	https://mc04.manuscriptcentral.com/crmc

https://mc04.manuscriptcentral.com/crmc

 [36] Wolf, H. The effect of hormones and vitamin B6 on urinary excretion of metabolites of the kynurenine pathway. <i>Scand J Clin Lab Invest Suppl</i>, 1974, <i>136</i>, 1-186. [37] Schwarcz, R. Metabolism and function of brain kynurenines. <i>Biochem Soc Trans</i>, 1993, <i>21</i> (1), 77-82.
 [38] Stone, T. W.; Perkins, M. N. Actions of excitatory amino acids and kynurenic acid in the primate hippocampus: a preliminary study. <i>Neurosci Lett</i>, 1984, <i>52</i> (3), 335-40. [39] Perkins, M. N.; Stone, T. W. Actions of kynurenic acid and quinolinic acid in the rat
hippocampus in vivo. <i>Exp Neurol</i> , 1985 , <i>88</i> (3), 570-9.
 [40] Han, Q.; Cai, T.; Tagle, D. A.; Li, J. Structure, expression, and function of kynurenine aminotransferases in human and rodent brains. <i>Cell Mol Life Sci</i>, 2010, <i>67</i> (3), 353-68. [41] Birch, P. J.; Grossman, C. J.; Hayes, A. G. Kynurenic acid antagonises responses to NMDA via an action at the strychnine-insensitive glycine receptor. <i>Eur J Pharmacol</i>, 1988,
154 (1), 85-7.[42] Kessler, M.; Terramani, T.; Lynch, G.; Baudry, M. A glycine site associated with N-
methyl-D-aspartic acid receptors: characterization and identification of a new class of antagonists. <i>J Neurochem</i> , 1989 , <i>52</i> (4), 1319-28.
[43] Prescott, C.; Weeks, A. M.; Staley, K. J.; Partin, K. M. Kynurenic acid has a dual action on AMPA receptor responses. <i>Neurosci Lett</i> , 2006 , <i>402</i> (1-2), 108-12.
[44] Rozsa, E.; Robotka, H.; Vecsei, L.; Toldi, J. The Janus-face kynurenic acid. <i>J Neural</i>
<i>Transm</i> , 2008 , <i>115</i> (8), 1087-91. [45] Hilmas, C.; Pereira, E. F.; Alkondon, M.; Rassoulpour, A.; Schwarcz, R.; Albuquerque,
E. X. The brain metabolite kynurenic acid inhibits alpha7 nicotinic receptor activity and increases non-alpha7 nicotinic receptor expression: physiopathological implications. <i>J</i>
<i>Neurosci</i> , 2001 , <i>21</i> (19), 7463-73. [46] Carpenedo, R.; Pittaluga, A.; Cozzi, A.; Attucci, S.; Galli, A.; Raiteri, M.; Moroni, F.
Presynaptic kynurenate-sensitive receptors inhibit glutamate release. <i>Eur J Neurosci</i> , 2001 , <i>13</i> (11), 2141-7.
[47] Dobelis, P.; Staley, K. J.; Cooper, D. C. Lack of modulation of nicotinic acetylcholine
alpha-7 receptor currents by kynurenic acid in adult hippocampal interneurons. <i>PloS one</i> , 2012, 7 (7), e41108.
[48] Alkondon, M.; Pereira, E. F.; Yu, P.; Arruda, E. Z.; Almeida, L. E.; Guidetti, P.; Fawcett, W. P.; Sapko, M. T.; Randall, W. R.; Schwarcz, R.; Tagle, D. A.; Albuquerque, E. X.
Targeted deletion of the kynurenine aminotransferase ii gene reveals a critical role of endogenous kynurenic acid in the regulation of synaptic transmission via alpha7 nicotinic
receptors in the hippocampus. <i>J Neurosci</i> , 2004 , <i>24</i> (19), 4635-48. [49] Stone, T. W.; Darlington, L. G. The kynurenine pathway as a therapeutic target in
cognitive and neurodegenerative disorders. <i>Br J Pharmacol</i> , 2013 , <i>169</i> (6), 1211-27.
[50] Berlinguer-Palmini, R.; Masi, A.; Narducci, R.; Cavone, L.; Maratea, D.; Cozzi, A.; Sili, M.; Moroni, F.; Mannaioni, G. GPR35 activation reduces Ca2+ transients and contributes to the kynurenic acid-dependent reduction of synaptic activity at CA3-CA1 synapses. <i>PLoS One</i> ,
 2013, 8 (11), e82180. [51] Wang, J.; Simonavicius, N.; Wu, X.; Swaminath, G.; Reagan, J.; Tian, H.; Ling, L. Kynurenic acid as a ligand for orphan G protein-coupled receptor GPR35. <i>J Biol Chem</i>, 2006,
<i>281</i> (31), 22021-8.
[52] Sas, K.; Robotka, H.; Rozsa, E.; Agoston, M.; Szenasi, G.; Gigler, G.; Marosi, M.; Kis, Z.; Farkas, T.; Vecsei, L.; Toldi, J. Kynurenine diminishes the ischemia-induced histological
and electrophysiological deficits in the rat hippocampus. <i>Neurobiol Dis</i> , 2008 , <i>32</i> (2), 302-8.
[53] Nozaki, K.; Beal, M. F. Neuroprotective effects of L-kynurenine on hypoxia-ischemia and NMDA lesions in neonatal rats. <i>J Cereb Blood Flow Metab</i> , 1992 , <i>12</i> (3), 400-7.
14

[54] Miranda, A. F.; Boegman, R. J.; Beninger, R. J.; Jhamandas, K. Protection against quinolinic acid-mediated excitotoxicity in nigrostriatal dopaminergic neurons by endogenous kynurenic acid. Neuroscience, 1997, 78 (4), 967-75. [55] Kumar, A.; Babu, G. N. In vivo neuroprotective effects of peripheral kynurenine on acute neurotoxicity induced by glutamate in rat cerebral cortex. Neurochem Res, 2010, 35 (4), 636-44. [56] Stone, T. W.; Perkins, M. N. Quinolinic acid: a potent endogenous excitant at amino acid receptors in CNS. Eur H Pharmacol, 1981, 72 (4), 411-2. [57] Perkins, M. N.; Stone, T. W. Pharmacology and regional variations of quinolinic acidevoked excitations in the rat central nervous system. J Pharm Exp Ther. 1983. 226 (2), 551-7. [58] Rodriguez-Martinez, E.; Camacho, A.; Maldonado, P. D.; Pedraza-Chaverri, J.; Santamaria, D.; Galvan-Arzate, S.; Santamaria, A. Effect of quinolinic acid on endogenous antioxidants in rat corpus striatum. Brain Res, 2000, 858 (2), 436-9. [59] Behan, W. M.; McDonald, M.; Darlington, L. G.; Stone, T. W. Oxidative stress as a mechanism for quinolinic acid-induced hippocampal damage: protection by melatonin and deprenyl. Br J Pharmacol, 1999, 128 (8), 1754-60. [60] Rios, C.; Santamaria, A. Quinolinic acid is a potent lipid peroxidant in rat brain homogenates. Neurochem Res, 1991, 16 (10), 1139-43. [61] Tavares, R. G.; Tasca, C. I.; Santos, C. E.; Alves, L. B.; Porciuncula, L. O.; Emanuelli, T.; Souza, D. O. Quinolinic acid stimulates synaptosomal glutamate release and inhibits glutamate uptake into astrocytes. Neurochem Int, 2002, 40 (7), 621-7. [62] Rassoulpour, A.; Wu, H. O.; Ferre, S.; Schwarcz, R. Nanomolar concentrations of kynurenic acid reduce extracellular dopamine levels in the striatum. J Neurochem, 2005, 93 (3), 762-5.[63] Erhardt, S.; Oberg, H.; Engberg, G. Pharmacologically elevated levels of endogenous kynurenic acid prevent nicotine-induced activation of nigral dopamine neurons. Naunyn Schmiedebergs Arch Pharmacol, 2001, 363 (1), 21-7. [64] Wu, H. Q.; Rassoulpour, A.; Schwarcz, R. Effect of systemic L-DOPA administration on extracellular kynurenate levels in the rat striatum. J Neural Transm, 2002, 109 (3), 239-49. [65] Poeggeler, B.; Rassoulpour, A.; Guidetti, P.; Wu, H. Q.; Schwarcz, R. Dopaminergic control of kynurenate levels and N-methyl-D-aspartate toxicity in the developing rat striatum. Dev Neurosci, 1998, 20 (2-3), 146-53. [66] Szalardy, L.; Klivenyi, P.; Zadori, D.; Fulop, F.; Toldi, J.; Vecsei, L. Mitochondrial disturbances, tryptophan metabolites and neurodegeneration: medicinal chemistry aspects. Curr Med Chem, 2012, 19 (13), 1899-920. [67] Stone, T. W.; Forrest, C. M.; Stoy, N.; Darlington, L. G. Involvement of kynurenines in Huntington's disease and stroke-induced brain damage. J Neural Transm, 2012, 119 (2), 261-74. [68] Majlath, Z.; Toldi, J.; Vecsei, L. The potential role of kynurenines in Alzheimer's disease: pathomechanism and therapeutic possibilities by influencing the glutamate receptors. J Neural Transm, 2014, 121 (8), 881-9. [69] Vecsei, L.; Szalardy, L.; Fulop, F.; Toldi, J. Kynurenines in the CNS: recent advances and new questions. Nat Rev Drug Discov, 2013, 12 (1), 64-82. [70] Widner, B.; Leblhuber, F.; Fuchs, D. Increased neopterin production and tryptophan degradation in advanced Parkinson's disease. J Neural Transm, 2002, 109 (2), 181-9. [71] Ogawa, T.; Matson, W. R.; Beal, M. F.; Myers, R. H.; Bird, E. D.; Milbury, P.; Saso, S. Kynurenine pathway abnormalities in Parkinson's disease. Neurology, **1992**, 42 (9), 1702-6. [72] Hartai, Z.; Klivenyi, P.; Janaky, T.; Penke, B.; Dux, L.; Vecsei, L. Kynurenine metabolism in plasma and in red blood cells in Parkinson's disease. J Neurol Sci, 2005, 239 (1), 31-5.

58 59

[73] Aarsland, D.; Ballard, C.; Walker, Z.; Bostrom, F.; Alves, G.; Kossakowski, K.; Leroi, I.; Pozo-Rodriguez, F.; Minthon, L.; Londos, E. Memantine in patients with Parkinson's disease dementia or dementia with Lewy bodies: a double-blind, placebo-controlled, multicentre trial.
Lancet Neurol, 2009 , 8 (7), 613-8. [74] Li, W.; Zhao, J. H.; Sun, S. G.; Zhang, J. W.; Suo, A. Q.; Ma, M. M. [Clinical rehabilitative effect of memantine on cognitive and motor disorders in patients with Parkinson's disease]. <i>Zhonghua Yi Xue Za Zhi</i> , 2011 , <i>91</i> (5), 301-3. [75] Leroi, L: Overshott, P. : Purne, F. L: Daniel, F. : Purns, A. Pandomized controlled trial of
[75] Leroi, I.; Overshott, R.; Byrne, E. J.; Daniel, E.; Burns, A. Randomized controlled trial of memantine in dementia associated with Parkinson's disease. <i>Mov Disord</i> , 2009 , <i>24</i> (8), 1217-21
21. [76] Moreau, C.; Delval, A.; Tiffreau, V.; Defebvre, L.; Dujardin, K.; Duhamel, A.; Petyt, G.; Hossein-Foucher, C.; Blum, D.; Sablonniere, B.; Schraen, S.; Allorge, D.; Destee, A.; Bordet, R.; Devos, D. Memantine for axial signs in Parkinson's disease: a randomised, double-blind, placebo-controlled pilot study. <i>J Neurol Neurosurg Psychiatry</i> , 2013 , <i>84</i> (5), 552-5.
[77] Varanese, S.; Howard, J.; Di Rocco, A. NMDA antagonist memantine improves
levodopa-induced dyskinesias and "on-off" phenomena in Parkinson's disease. <i>Mov Disord</i> , 2010 , <i>25</i> (4), 508-10.
[78] Vidal, E. I.; Fukushima, F. B.; Valle, A. P.; Villas Boas, P. J. Unexpected improvement in levodopa-induced dyskinesia and on-off phenomena after introduction of memantine for treatment of Parkinson's disease dementia. <i>J Am Geriatr Soc</i> , 2013 , <i>61</i> (1), 170-2.
[79] Del Dotto, P.; Pavese, N.; Gambaccini, G.; Bernardini, S.; Metman, L. V.; Chase, T. N.; Bonuccelli, U. Intravenous amantadine improves levadopa-induced dyskinesias: an acute double-blind placebo-controlled study. <i>Mov Disord</i> , 2001 , <i>16</i> (3), 515-20.
[80] Thomas, A.; Iacono, D.; Luciano, A. L.; Armellino, K.; Di Iorio, A.; Onofrj, M. Duration of amantadine benefit on dyskinesia of severe Parkinson's disease. <i>J Neurol Neurosurg</i>
<i>Psychiatry</i> , 2004 , <i>75</i> (1), 141-3.
[81] Verhagen Metman, L.; Del Dotto, P.; van den Munckhof, P.; Fang, J.; Mouradian, M.M.; Chase, T. N. Amantadine as treatment for dyskinesias and motor fluctuations in
Parkinson's disease. <i>Neurology</i> , 1998 , <i>50</i> (5), 1323-6.
[82] Wolf, E.; Seppi, K.; Katzenschlager, R.; Hochschorner, G.; Ransmayr, G.; Schwingenschuh, P.; Ott, E.; Kloiber, I.; Haubenberger, D.; Auff, E.; Poewe, W. Long-term antidyskinetic efficacy of amantadine in Parkinson's disease. <i>Mov Disord</i> , 2010 , <i>25</i> (10),
1357-63.
[83] Paquette, M. A.; Martinez, A. A.; Macheda, T.; Meshul, C. K.; Johnson, S. W.; Berger, S. P.; Giuffrida, A. Anti-dyskinetic mechanisms of amantadine and dextromethorphan in the
6-OHDA rat model of Parkinson's disease: role of NMDA vs. 5-HT1A receptors. <i>Eur J Neurosci</i> , 2012 , <i>36</i> (9), 3224-34.
[84] Cera, N.; Bifolchetti, S.; Martinotti, G.; Gambi, F.; Sepede, G.; Onofrj, M.; Di Giannantonio, M.; Thomas, A. Amantadine and cognitive flexibility: decision making in Parkinson's patients with severe pathological gambling and other impulse control disorders.
 Neuropsychiatr Dis Treat, 2014, 10, 1093-101. [85] Pettorruso, M.; Martinotti, G.; Di Nicola, M.; Onofrj, M.; Di Giannantonio, M.; Conte, G.; Janiri, L. Amantadine in the treatment of pathological gambling: a case report. <i>Front</i>
<i>Psychiatry</i> , 2012 , <i>3</i> , 102. [86] Kashihara, K.; Imamura, T. Amantadine may reverse punding in Parkinson's disease-
observation in a patient. <i>Mov Disord</i> , 2008 , <i>23</i> (1), 129-30.
[87] Fasano, A.; Ricciardi, L.; Pettorruso, M.; Bentivoglio, A. R. Management of punding in Parkinson's disease: an open-label prospective study. <i>J Neurol</i> , 2011 , <i>258</i> (4), 656-60.
[88] Silva-Adaya, D.; Perez-De La Cruz, V.; Villeda-Hernandez, J.; Carrillo-Mora, P.; Gonzalez-Herrera, I. G.; Garcia, E.; Colin-Barenque, L.; Pedraza-Chaverri, J.; Santamaria, A.
16

Protective effect of L-kynurenine and probenecid on 6-hydroxydopamine-induced striatal toxicity in rats: implications of modulating kynurenate as a protective strategy. *Neurotoxicol Teratol*, **2011**, *33* (2), 303-12.

[89] Graham, W. C.; Robertson, R. G.; Sambrook, M. A.; Crossman, A. R. Injection of excitatory amino acid antagonists into the medial pallidal segment of a 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine (MPTP) treated primate reverses motor symptoms of parkinsonism. *Life sciences*, **1990**, *47* (18), PL91-7.

[90] Acuna-Castroviejo, D.; Tapias, V.; Lopez, L. C.; Doerrier, C.; Camacho, E.; Carrion, M. D.; Mora, F.; Espinosa, A.; Escames, G. Protective effects of synthetic kynurenines on 1methyl-4-phenyl-1,2,3,6-tetrahydropyridine-induced parkinsonism in mice. *Brain Res Bull*, **2011**, *85* (3-4), 133-40.

[91] Gregoire, L.; Rassoulpour, A.; Guidetti, P.; Samadi, P.; Bedard, P. J.; Izzo, E.; Schwarcz, R.; Di Paolo, T. Prolonged kynurenine 3-hydroxylase inhibition reduces development of levodopa-induced dyskinesias in parkinsonian monkeys. *Behav Brain Res*, **2008**, *186* (2), 161-7.

[92] Ouattara, B.; Belkhir, S.; Morissette, M.; Dridi, M.; Samadi, P.; Gregoire, L.; Meltzer, L. T.; Di Paolo, T. Implication of NMDA receptors in the antidyskinetic activity of cabergoline, CI-1041, and Ro 61-8048 in MPTP monkeys with levodopa-induced dyskinesias. *J Mol Neurosci*, **2009**, *38* (2), 128-42.

[93] Fulop, F.; Szatmari, I.; Vamos, E.; Zadori, D.; Toldi, J.; Vecsei, L. Syntheses, transformations and pharmaceutical applications of kynurenic acid derivatives. *Curr Med Chem*, **2009**, *16* (36), 4828-42.

[94] Fulop, F.; Szatmari, I.; Toldi, J.; Vecsei, L. Modifications on the carboxylic function of kynurenic acid. *J Neural Transm*, **2012**, *119* (2), 109-14.

[95] Zadori, D.; Nyiri, G.; Szonyi, A.; Szatmari, I.; Fulop, F.; Toldi, J.; Freund, T. F.; Vecsei, L.; Klivenyi, P. Neuroprotective effects of a novel kynurenic acid analogue in a transgenic mouse model of Huntington's disease. *J Neural Transm*, **2011**, *118* (6), 865-75.

[96] Gellert, L.; Fuzik, J.; Goblos, A.; Sarkozi, K.; Marosi, M.; Kis, Z.; Farkas, T.; Szatmari, I.; Fulop, F.; Vecsei, L.; Toldi, J. Neuroprotection with a new kynurenic acid analog in the four-vessel occlusion model of ischemia. *Eur J Pharmacol*, **2011**, *667* (1-3), 182-7.

[97] Gellert, L.; Varga, D.; Ruszka, M.; Toldi, J.; Farkas, T.; Szatmari, I.; Fulop, F.; Vecsei, L.; Kis, Z. Behavioural studies with a newly developed neuroprotective KYNA-amide. *J Neurla Transm*, **2012**, *119* (2), 165-72.

Current Medicinal Chemistry

Excitotoxic mechanisms in non-motor dysfunctions and levodopa-induced dyskinesia in Parkinson's disease: the role of the interaction between dopaminergic and the kynurenine system

Zsófia Majláth¹, József Toldi^{2,4}, Ferenc Fülöp³, László Vécsei*^{1,4}

¹ Department of Neurology, University of Szeged, Semmelweis u. 6, H-6725, Szeged Hungary

² Department of Physiology, Anatomy and Neuroscience, University of Szeged, Közép fasor

52, H-6726 Szeged, Hungary

³ Department of Pharmaceutical Chemistry, Faculty of Pharmacy, University of Szeged, Szeged, Hungary

⁴ MTA-SZTE Neuroscience Research Group of the Hungarian Academy of Sciences and University of Szeged, Semmelweis u. 6, H-6725 Szeged, Hungary

*Corresponding author. E-mail: vecsei.laszlo@med.u-szeged.hu, Tel.: +36-62-545348, Fax: +36-62-545597

Acknowledgements

This work was supported by the MTA-SZTE Neuroscience Research Group of the Hungarian Academy of Sciences and the University of Szeged and by the projects entitled TÁMOP-4.2.2.A-11/1/KONV-2012-0052, OTKA K105077 and the Hungarian Brain Research Programme (NAP, Grant No. KTIA-13-NAP-A-III/9. and KTIA-13-NAP-A-II/17.). Thanks are due to David Durham for the linguistic corrections of the manuscript.

Conflict of Interest

The authors have no conflict of interest to report.