Tryptophan catabolites and migraine

Zsuzsanna Bohár^{a,b}, Árpád Párdutz^b, László Vécsei^{*a,b}

^aMTA-SZTE Neuroscience Research Group, University of Szeged, Szeged, Hungary; ^bDepartment of Neurology, Faculty of Medicine, Albert Szent-Györgyi Clinical Center, University of Szeged, Szeged, Hungary

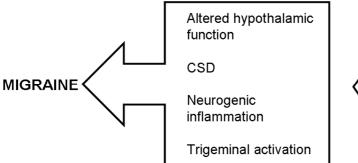
*Corresponding author: Prof. László Vécsei Director, Department of Neurology University of Szeged, Faculty of Medicine Albert Szent-Györgyi Clinical Center Semmelweis u. 6. H-6725 Szeged, Hungary Phone: +36 62 545348 Fax: +36 62 545597 E-mail: <u>vecsei.laszlo@med.u-szeged.hu</u>

Abstract:

Migraine is a highly disabling neurological condition affecting around 15% of the population worldwide. Decades of intensive research shed some light on diseases pathomechanism, but information is still missing about the initiation of the attack. In the past century, serotonin emerged as the main target of both basic and therapeutic research. As a result, the triptans, the only approved migraine specific drugs were developed. The involvement of glutamatergic mechanism in migraine headache development such as cortical hyperexcitability, and cortical spreading depression as the pathological correlate of migraine aura called the attention to the kynurenine pathway in migraine pathomechanism. The serotonin and kynurenine pathways are closely connected, as they both are the metabolic routes of the amino acid tryptophan. Kynurenine catabolites are important participants in glutamatergic neurotransmission, regulation also nociceptive processing of the trigeminal system. The current work attempts to collect recent data on both serotonin and kynurenine research related to migraine and emphasizes the importance of further research on this topic.

Keywords: migraine, kynurenine pathway, cortical spreading depression, serotonin, hyperexcitability, glutamate.

Graphical abstract



Tryptophan catabolites

1. INTRODUCTION

Migraine is a common neurological disorder with a one-year prevalence of 14.7% [1]. Despite intensive research the exact pathomechanism of the disease is still not completely understood. The importance of migraine research is clearly shown by the fact that migraine is the sixth highest cause of disability worldwide, and together the headache disorders are the third on the list of causes of disability [2].

Based on data so far, migraine can be considered as a painful multifactorial disorder [3], involving autonomic, affective, cognitive and sensory functions. Glutamate and serotonin mediated neurotransmission both play an important role in the evolution of migraine headaches [4]. The kynurenine pathway (KP), the main route of tryptophan metabolism, is closely involved with both glutamatergic and serotonergic mechanisms placing the catabolites of this pathway under spotlight in migraine research [5]. The scope of our review is to highlight the possible connections of the KP with processes involved in migraine pathomechanism.

2. METABOLISM OF TRYPTOPHAN

In mammals, the excess of tryptophan not necessary for protein synthesis is utilized either for the synthesis of the important neurotransmitters serotonin (5-hydroxytryptamine, 5HT) and melatonin (N-acetyl-5-methoxytrypamine) or is oxidized via the KP. In humans more than 90% of dietary tryptophan is metabolized via the KP at the periphery [6]. Both metabolic routes include important neuroactive catabolites with a possible role in the pathomechanism of migraine headache.

2.1 The Kynurenine Pathway

The first, rate-limiting step of the kynurenine pathway (KP) is the irreversible cleavage of the indole ring of L-tryptophan (TRP) either by indoleamine 2,3-dioxygenase 1 and 2 (IDO1 and 2), or by tryptophan 2,3-dioxygenase (TDO) (Figure 1).

IDO1 can be found throughout the body, and functions as a monomeric protein. Besides TRP, it can also utilize D-tryptophan, 5-hydroxy-L-kynurenine, 5-hydroxy-D-kynurenine, melatonin [7] and serotonin [8].

Less is known about IDO2, since it was isolated less than 10 years ago [9]. Data gathered so far indicate that the expression of IDO2 is not as widespread as that of IDO1, and that this enzyme may influences immune regulation differently than IDO1 [10]. TDO functions as a homotetramer and contains a heme B group. It is expressed predominantly in the liver, but can also be found in the brain and placenta [11, 12]. TDO is more substrate specific than IDO 1, claiming only L-tryptophan for metabolism [13].

Reactions catalyzed by either enzymes result in the formation of N-formyl-L-kynurenine, which is further degraded by kynurenine formamidase to L-kynurenine (L-KYN), the central metabolite of the KP. From L-KYN the kynurenine aminotransferases (KATs) form kynurenic acid (KYNA), kynureninase forms anthranilic acid (AA), while kynurenine 3-monooxygenase (KMO) activity results in the formation of 3-hydroxykynurenine (3HK). In mammals four type of KATs are present, KAT I also known as glutamine transaminase K/cysteine conjugate beta-lyase I [14, 15], KAT II/aminoadipate aminotransferase [16, 17], KAT III/cysteine conjugate beta-lyase II [18] and KAT IV/glutamic-oxaloacetic transaminase 2/mitochondrial aspartate aminotransferase [19, 20]. All of the KATs are able to metabolize L-KYN to KYNA, however their substrate specificity, metabolic rate and expression are different [21].

Kynureninase is a pyridoxal-5'-phosphate dependent enzyme, it is able to catalyze two reactions along the KP it converts L-KYN to AA and it promotes the conversion of 3HK to 3-hydroxyanthranilic acid (3HA). The first mechanism is common in prokaryotes while in eukaryotes the latter pathway is preferred, therefore in mammals the conversion of L-KYN proceeds either to the KYNA or to the 3HK route [22].

AA is further metabolized by anthranilae-3-monooxygenase to 3HA, while from 3HK kynureninase forms the same compound. 3HK can be also converted by KATs to xanthurenic acid. 3HA is either subjected to auto oxidation resulting in cinnabarinic acid, or it is converted to 2-amino-3-carboxymuconate semialdehyde (ACMS) by 3-hydroxyanthranilic acid oxygenase. ACMS can be further processed by aminocarboxymuconate-semialdehyde decarboxylase to 2-aminomuconate semialdehyde, followed by a non-enzymatic transformation to picolinic acid. ACMS can also undergo spontaneous pyridine ring closure to form quinolinic acid (QUIN), which can be further processed to form nicotinamide adenine dinucleotide.

2.2 Serotonin pathway

The first rate-limiting step in the synthesis of 5HT is the hydroxylation of L-tryptophan by the enzyme tryptophan hydroxylase (TPH) and results in the formation of 5-hydroxytryptophan. This metabolite is transformed by aromatic amino acid decarboxylase (AADA) to 5HT. Metabolism of 5HT by monoamine oxidase (MAO) to 5-hydroxyindole acetaldehyde and further by aldehyde dehydrogenase, results in the end product 5-hydroxyindoleacetic acid (5HIAA), which is generally used to indirectly detect changes in serotonin levels. In the pineal gland and in the retina 5HT is metabolized by serotonin N-acetyltransferase to N-acetylserotonin, which is further converted by hydroxyindole-O-methyltransferase to melatonin.

2.3 Receptorial and non-receptorial actions of tryptophan catabolites

Several kynurenine catabolites were shown to have direct receptorial actions. L-KYN, KYNA and cinnabarinic acid are ligands of aryl hydrocarbon receptors (AHRs) [23-25]. AHR is a nuclear transcription factor regulating the expression of genes of xenobiotic metabolizing enzymes, such as cytochrome P450 protein and glutathione transferase. This receptor possesses important functions in immune regulation, and T-cell development [26].

KYNA is an endogenous ionotropic glutamate receptor antagonist, acting on N-methyl-D-aspartate (NMDA) receptors at both the strychnine-insensitive glycine site and on the glutamate recognition site [27, 28]. In addition it is also an antagonist of kainate glutamate receptors [29], while it has a Janus-face effect on α -amino-3-hydroxy-5-methyl-4-isoxazole propionic acid (AMPA) receptors, in nanomolar concentration it has a facilitatory, while in micromolar concentration an inhibitory effect on these receptor [30-32]. KYNA's broad receptorial action indicates that it has a very important role in the regulation of glutamatergic neurotransmission. This regulatory role is further supported by the fact that KYNA is an antagonist of α 7-nicotinic acetylcholine (α 7nACh) receptors, thus is able to modulate presynaptic glutamate release [33]. KYNA also has weak agonistic effects on the G protein coupled receptor 35 (GPCR 35) [34].

QUIN is a weak endogenous agonist of NMDA receptors [35], while cinnabarinic acid acts on type 4 metabotropic glutamate receptors [36].

Besides the direct receptorial actions of tryptophan catabolites, they also participate on oxidative processes, and thus are important factors in mitochondrial energy metabolism [37].

The effects of 5HT are transmitted by 7 receptor subtypes, 5HT1-7 receptors. Throughout them 5HT exerts myriad of functions, modulating mood, memory, appetite and circadian rhythms. With one exception, 5HT3, which is a ligand gated cation channel, 5HT receptors are G protein coupled receptors (GPCRs) [38].

In humans melatonin exerts its effect via two GPCRs, MT1 and MT2 [39-41], regulating circadian rhythms, and participating in the pathomechanism of depression, autism spectrum disorders and neurodegenerative disorders [42-44]. Melatonin is also a potent antioxidant [45].

3. MIGRAINE

Migraine attacks present with an unilateral throbbing headache accompanied by nausea or vomiting, and photo- or phonophobia. Attacks can be divided into four phases (prodrome, aura, headache and postdrome), however not every phase is present in every patient [46].

3.1 Prodrome

The first phase is the prodorme/ premonitory symptom phase occurring hours or even days before the headache attack [47]. Premonitory phase can be present with diverse symptoms including fatigue, mood changes, food cravings, concentration problems, sleep disturbances, gastrointestinal disturbances and yawning [48, 49]. The prevalence of premonitory symptoms shows great variance among different studies ranging from 33 to 80% [46, 50, 51]. Griffin and colleagues have revealed that about two-third of migraine patients are able to predict the occurrence of the headache based on premonitory symptoms [52], suggesting that these symptoms are an integral part of the migraine attack, not just accompaniments. Two hypotheses emerged attempting to explain the mechanisms behind the symptoms, one involving the neurotransmitter dopamine, the other involving the hypothalamus.

The dopamine hypothesis is based on the observation that dopamine agonists produce the same symptoms which are present during the premonitory phase, while antagonists, such as domperidone and metoclopramide may reverse some of these symptoms and even prevent or moderate the headache [53-55].

Hypothalamus is the most important nucleus in the human brain involved in the maintenance of homeostasis; it regulates endocrine functions, circadian rhythms and harmonizes the function of the parasympathetic and sympathetic nervous systems. The diverse functions imply that the impaired operation of the hypothalamus may result in very different and at firs glance not related symptoms, resembling migraine premonitory symptoms. Furthermore, the clear connection between migraine and the sleep-wake cycle supports the involvement of the hypothalamus in the pathomechanism of migraine, but the exact routes of action need to be clarified [56, 57].

3.2 Aura

According to the classification of the International Headache Society the two most common forms of migraine are migraine without aura and migraine with aura [58]. About one-third of the patient suffering from migraine can be classified to the MA group [59], where aura indicates the presence of preceding or accompanying transient focal neurological symptoms related to the headache. The aura symptoms are in the most cases visual, however sensory, language and motor symptoms can also manifest [60]. A typical aura develops within 5 minutes and lasts for 5-60 minutes [58].

Cortical spreading depression (CSD) is considered as the pathophysiological equivalent of migraine aura [61, 62]. During CSD, a wave of transient depolarization spreads along the cortex accompanied by hyper- and hypoperfusion. The spreading velocity of the depolarization wave, 2-5 mm/s, correlates with the spread of the visual aura [61, 63]. Zhang and colleagues showed that after focal induction of CSD in rats a long lasting activation of meningeal nociceptors occurs with a delay, suggesting that CSD is able to directly activate these neurons [64]. However, another study on awake animals did not show pain behaviour after the induction of CSD, challenging the theory of CSD being the primary trigger of migraine headache [65]. Knock in models of familial hemiplegic migraine (FHM), an autosomal dominantly inherited form of migraine, are more susceptible to CSD, suggesting common mechanisms in CSD and FHM development [66]. In the formation of CSD, glutamate neurotransmission and NMDA receptors play a crucial role. Agonist of glutamate receptors can trigger CSD [67], while the non-competitive NMDA antagonist MK 801 is able to suppress the formation of cortical CSD waves [68]. Recent data suggest that different subtypes of NMDA receptors may play diverse roles in the formation of CSD, thus subtype specific antagonist may provide a therapeutic approach against CSD [69].

3.3 Headache phase and postdrome

For the great part of patients the most debilitating part of the migraine attack is the headache phase. Headache can last for 4-72 hours without treatment, with mild or severe strength [58]. The headache is thought to be caused by the activation and sensitization of the trigeminovascular system, consisting of the trigeminal nerve and innervated areas [70]. The trigeminal nerve is the largest among cranial nerves in humans; it is divided into three main divisions, the ophthalmic, maxillary and mandibular branch, of which the ophthalmic gives the sensory innervations of the meninges.

For decades the vascular changes during the migraine attack were thought to be the cause of the headache, later a neuronal basic mechanism was suggested, and more recently the neurovascular origin of the headache was proposed, however, the primary triggering step in migraine generation is still debated [70].

The activation of the primary nociceptive trigeminal afferents leads to peripheral neuropeptide release, calcitonin gene-related peptide (CGRP), neurokinin A and substance P are liberated to the perineural space [71, 72], leading to vasodilatation and plasma protein extravasation and to the activation of mast cells and leukocytes collectively termed as neurogenic inflammation [73]. These phenomena are well characterized in animals, however in humans only little direct evidence supports the actual presence of neurogenic inflammation during the attack [74]. Despite the lack of human data, the importance of neuropeptides, is supported by current drug research data focusing on CGRP receptor antagonist and anti-CGRP antibodies [75, 76].

The continuous activation of the peripheral trigeminal nociceptors leads to peripheral sensitization which presents with the worsening of headache during activities increasing intracranial pressure e.g. coughing and in the throbbing nature of the headache [77].

The next important structure during the process of migraine headache is the caudal part of the spinal trigeminal nucleus (TNC), where most of the primary nociceptive afferents project. Activation at the periphery proceeds to the TNC, mostly by the aid of glutamate, as its level increases in the TNC after stimulation of the first-order trigeminal neurons [78]. Both glutamate [79] and its receptors [80] are present in the trigeminal system providing an important modulatory aspect of trigeminal nociception. Activation of the second-order

neurons in the TNC can lead to central sensitization at this site [81], which presents in migraineurs during the attack as the symptom of allodynia [82].

Activation of the trigeminal system spreads forward to thalamic and cortical sites, resulting in the pain experienced by migraine patients.

Imaging studies revealed, that specific brain regions (periaqueductal grey matter, nucleus raphe magnus, dorsal raphe nucleus, locus coeruleus) are active during the headache phase, raising the possibility of the existence of a migraine generator region [83, 84]. These nuclei are the part of the descending pain modulatory system, their activation suggest that the processing and transmission of nociceptive information can be dysfunctional in migraine, however these theory needs to be confirmed.

After the cessation of the migraine pain patients often experience tiredness, difficulties in concentration, weakness, dizziness, which can be regarded as the postdrome phase, the least well-defined phase of the migraine attack [85, 86].

3.4 Cortical hyperexcitability in migraine

Cortical hyperexcitability is thought to contribute to the formation of CSD in migraine. The first evidence for this hypothesis is that the mutations of the three FHM genes can lead to hyperexcitability. FHM 1 is caused by the mutation in the *CACNA1A* gene located on the chromosome 19p13 coding the α subunit of the voltage-gated P/Q type Cav2.1 calcium channel [87]. In knock in animals this mutation results in increased glutamate release in the brain [88]. The second type of FHM, FHM II is caused by the mutation in the *ATP1A2* gene [89] encoding the α subunit of Na/K pumps. Mutations of this gene can cause decreased uptake of glutamate and K+ from the synaptic cleft. Mutations in the *SCN1A* gene are responsible for the third type of FHM, FHM III. This gene encodes the α 1 subunit of the neuronal Nav1.1 voltage-gated sodium channels [90], and leads to accelerated recovery of the channel from fast inactivation, leading to a higher firing rate [91]. These data indicate that FHM mutations can result in cortical hyperexcitability, suggesting that this phenomenon may also be involved in the formation of simple migraine.

Migraine patients show increased response to various sensory stimuli, further implicating the presence of a hyperexcitable state [92]. The lack of habituation to repeated sensory stimuli also confirms the presence of a hyper-responsible state even between migraine attacks [93-95]. With positron emission tomography (PET) Boulloche and colleagues found that an altered activation of the visual cortex is present in the interictal phase in migraineurs compared to controls, confirming the possibility of hyperexcitability with functional imaging [96].

Increased glutamate levels can be the basis of the hyperexcitable state; however, measurements of glutamate content in migraineurs provided conflicting results. Platelet and plasma measurements of glutamate indicated lower or similar levels in migraineurs and controls [97, 98], while in other studies increased basal glutamate levels were found [99]. During the attack increased levels of glutamate were found in the cerebrospinal fluid of migraine patients [97]. Proton magnetic resonance spectroscopy revealed that a higher glutamate/glutamine ratio is present in migrainous women during the interictal phase compared to controls in the occipital cortex [100], supporting the presence of altered GLU metabolism in migraine.

4. INVOLVEMENT OF TRYPTOPHAN CATABOLITES IN MIGRAINE

4.1 Serotonin in migraine

The importance of 5HT in migraine pathomechanism was first proposed by Sicuteri and colleagues in 1961 after detecting increased urinary 5HIAA levels during migraine attacks [101]. Based on these findings numerous studies examined peripheral 5HT and 5HIAA levels in plasma and platelets, and their results suggest that no alteration is present during the migraine attack in 5HT levels compared to controls [102]. These findings are not surprising if we consider that the main source of 5HT and 5HIAA is the gut, where enterochromaffin cells synthesize 5HT to fulfil its role in the modulation of peristalsis [103].

Serotonin is a vasoconstrictor [104], and it was revealed that the peripheral veins of migraineurs show higher reactivity to 5HT than controls, furthermore veins of women suffering from migraine react even more to 5HT than veins of migrainous men [102] supporting the involvement of 5HT in migraine.

The measurements of brain 5HT levels and synthetic rates also yielded conflicting results. PET scans with the aid of α -[11C] methyl-L tryptophan, a 5HT precursor, revealed an increased 5HT synthesis capacity and also a higher brain uptake rate in women with migraine [105]. Using the same method, another study found no differences in 5HT synthetic rates between migraineurs and controls during the attack, but interictally a lower synthetic rate was found in migraineurs [106]. The same group examined interictal 5HT synthesis in non-menopausal women, and found no alteration in synthetic rates, however after administration of a 5HT receptor agonist, eletriptan a decreased rate was found in migraineurs but not in controls, suggesting an altered regulation of 5HT synthesis between attacks [107].

Genetic research also attempted to link 5HT to migraine. Studies examining the genetic association between migraine and the serotonin synthesizing and metabolizing enzymes TPH, AADA, MAO detected no connection [108-111]. Genetic research on 5HT receptors revealed no obvious relation with migraine in the case of 5HT1A [112], 5HT2A [113-115] and 5HT2C [116, 117] receptors; however in a small number of affected families association was found with 5HT1D receptors [118]. A more promising subject is the 5HT transporter, as its gene polymorphisms were found to be associated with migraine in independent meta-analyses [119, 120].

In spite of the conflicting result regarding the levels of 5HT in migraineurs, the effectiveness of its agonist, triptans in migraine therapy confirms the importance of this compound in disease pathogenesis. Triptans activate 5HT1B/1D and 5HT1F receptors but many questions are unanswered regarding their mechanism and site of action [121, 122].

4.2 Melatonin in migraine

Several studies have found decreased nocturnal melatonin levels in migraineurs, especially in women [123, 124]. In an other study, no difference was found between pain free periods of migraine patients and controls, while a decrease of melatonin level was detected in migraineurs during headache attacks [125]. Based on these and on other results connecting migraine seasonal variations with melatonin [126], assumption raised, that melatonin may play an important role in migraine pathogenesis. Subsequently trials were conducted to examine the efficacy of melatonin in migraine therapy, but controversial results were obtained. In an open-label study of Peres an colleagues melatonin was effective in migraine prevention [127], while in a randomized, double-blind, placebo-controlled crossover study of Alstadhaug and co-workers no beneficiary effect of melatonin was detected [56]. Considering the importance of melatonin in the regulation of circadian rhythms and its close relation with hypothalamic functions, participation in migraine pathomechanism is highly possible, however the exact molecular mechanism and thus possible therapeutic efficacy need to be further confirmed.

4.3 Kynurenines in migraine

Since Sicuteri's observations about the involvement of 5HT in migraine pathomechanism [101] research focused on the 5HT pathway of TRP metabolism, although in the past years the KP's involvement was also confirmed [4, 128].

TRP was used as an analgesic agent [129], and later on it was revealed that its KP catabolites L-KYN, KYNA, AA, XA, PIC and QUIN also possess analgesic properties in the tail-flick and hot-plate tests when administered intraperitoneally to rats [130] further supporting their involvement in modulation of nociceptive processing and thus in migraine pathomechanism.

KP catabolites are present both peripherally and centrally in the trigeminal system [21], providing the opportunity to modulate trigeminal nociception at various sites. Knyihár-Csillik and colleagues found that after electrical stimulation of the trigeminal ganglion, a well characterized model of trigeminal activation and dural neurogenic inflammation, KAT immunoreactivity of the cerebral dura mater decreased in rats [131]. These results suggest that endogenous KYNA synthesis in the dura may contribute to the modulation of NMDA receptor function.

Several strategies exist to enhance the production of KYNA in the brain and to decrease the amounts of QUIN and 3HK, which are thought to be neurotoxic. These involve the application of KYNA precursors, inhibitors of KMO, to decrease the synthesis of 3HK and the development of new KYNA-like compounds [132]. These approaches were applied also in animal models of trigeminal activation for the evaluation of the involvement of KP catabolites in migraine. KYNA passes the blood-brain barrier (BBB) poorly [133], therefore its precursor L-KYN was applied in animal studies. In the previously mentioned electrical stimulation model the application of L-KYN in combination with probenecid was able to decrease the activation of the second order neurons in the TNC[134]. Probenecid is an inhibitor of organic acid transport [135], and in joint application with L-KYN it can prevent the depletion of KYNA from the central nervous system, raising its levels significantly [136].

The same combination was applied in the nitroglycerine induced model (NTG model), where it was also able to decrease the activation of the trigeminal system [137]. These results suggest that elevated KYNA levels may be protective against trigeminal hyperactivity.

In the NTG model sensitization processes also occur, [138, 139], presenting with enhanced neuronal nitric oxide synthase (nNOS) [140], calmodulin-dependent protein kinase II α (CamKII α) [141] and decreased CGRP content in the TNC [142]. The combination treatment was able to attenuate nNOS [136], while L-KYN by itself attenuated the changes of CamKII α and CGRP levels caused by NTG [143].

Two synthetic KYNA amides, 2 (2 N,N dimethylaminoethylamine-1-carbonyl)-1H-quinolin-4-one hydrochloride (KYNA-A1) and N-(2-N-pyrrolidinylethyl)-4-oxo-1H-quinoline-2-carboxamide hydrochloride (KYNA-A2) were also tested in the NTG model. KYNA-A1 was able to mitigate nNOS, CamKIIα and CGRP immunoreactivity changes after NTG administration, and KYNA-A2 had the same, but dose dependent effect [136, 143, 144]. These results clearly demonstrate the important role of L-KYN and KYNA in trigeminal nociceptive processing, and support their important role in migraine pathomechanism. However, the exact routes throughout nociceptive modulation occurs are elusive. The general consideration is that both KYNA and its derivatives modulate NMDA receptor function, but if their effect takes place peripherally or centrally needs to be clarified [136, 143, 144].

Studies on experimental CSD in rats conducted by Chauvel and colleagues support the interaction between KYNA and NMDA receptors in CSD formation. In their experiments both combined and standalone application of L-KYN and probenecid were able to decrease CSD frequency, the combined application having the greatest effect compared to control, via increasing the KYNA content in the brain [145]. Their results also show that there is a sex-dependence of the effects of L-KYN-probenecid treatment, female rats being more sensitive to it [145]. Other results have shown that KYNA administered peripherally can also modulate CSD frequency and BBB permeability [146]. Based on these result we can assume, that KYNA has an important role in mitigating CSD formation, and therefore it is involved in generation of migraine aura.

In studies conducted so far with KYNA derivatives the site of application was peripheral. Although the purpose of both the combinatory treatment and the application of derivatives are to increase KYNA content in the brain, the possible peripheral involvement of KYNA in migraine processes can not be excluded. On the periphery both KYNA and other KP catabolites can participate in the formation and modulation of neurogenic inflammation (Figure 2). At the level of the secondary trigeminal neurons KYNA and QUIN could modulate glutamatergic neurotransmission of nociceptive signals, while at higher levels, at the thalamus and in the cortex they may also influence trigeminal pain pathways. In a recent work, Kageneck and colleagues showed that KYNA was able to inhibit capsaicin induced CGRP release in the TNC, but had no effect on dural CGRP release, supporting the idea, that KYNA may exert altering effects at different sites of the trigeminal system [147]. Further knowledge about the KP's involvement in trigeminal nociceptive processing could lead to a better understanding of migraine pathomechanism.

5. CONCLUSIONS

The aim of the present work was to draw some attention on the role of different tryptophan catabolites, especially kynurenine catabolites, in migraine pathomechanism. 5HT have been shown to be involved in the nociceptive processing of trigeminal signals, and various aspects of migraine generation, and the other important transmitter of the same metabolic pathway, melatonin was also proposed to play a part in these processes. The close relationship of the 5HT pathway and the KP of tryptophan degradation suggest, that alterations in one arm of the pathway, may influence the other, thus refer to a highly probable involvement of the KP in migraine. Research conducted so far in relation with the KP and migraine were focused on KYNA, and results confirm the involvement of this compound in migraine related processes. However, further studies are needed to detail the possible sites and receptors of effect of KYNA and also the influence of other KP catabolites.

CONFLICT OF INTEREST

The Authors declare that they do not have any conflict of interest.

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REFERENCES

- [1] Stovner LJ, Andree C. Prevalence of headache in Europe: a review for the Eurolight project. J Headache Pain, 2010; 11: 289-99.
- [2] Steiner TJ, Birbeck GL, Jensen RH, Katsarava Z, Stovner LJ, Martelletti P. Headache disorders are third cause of disability worldwide. J Headache Pain, 2015; 16: 58.
- [3] Russell MB, Olesen J. Increased familial risk and evidence of genetic factor in migraine. BMJ, 1995; 311: 541-4.
- [4] Párdutz A, Fejes A, Bohár Z, Tar L, Toldi J, Vécsei L. Kynurenines and headache. J Neural Transm, 2012; 119: 285-96.
- [5] Curto M, Lionetto L, Fazio F, Mitsikostas DD, Martelletti P. Fathoming the kynurenine pathway in migraine: why understanding the enzymatic cascades is still critically important. Intern Emerg Med, 2015; 10: 413-21.
- [6] Leklem JE. Quantitative aspects of tryptophan metabolism in humans and other species: a review. Am J Clin Nutr, 1971; 24: 659-72.
- [7] Ferry G, Ubeaud C, Lambert PH, Bertin S, Cogé F, Chomarat P, Delagrange P, Serkiz B, Bouchet JP, Truscott RJ, Boutin JA. Molecular evidence that melatonin is enzymatically oxidized in a different manner than tryptophan: investigations with both indoleamine 2,3-dioxygenase and myeloperoxidase. Biochem J, 2005; 388: 205-15.
- [8] Shimizu T, Nomiyama S, Hirata F, Hayaishi O. Indoleamine 2,3-dioxygenase. Purification and some properties. J Biol Chem, 1978; 253: 4700-6.
- [9] Ball HJ, Sanchez-Perez A, Weiser S, Austin CJ, Astelbauer F, Miu J, McQuillan JA, Stocker R, Jermiin LS, Hunt NH. Characterization of an indoleamine 2,3-dioxygenase-like protein found in humans and mice. Gene, 2007; 396: 203-13.
- [10] Prendergast GC, Metz R, Muller AJ, Merlo LM, Mandik-Nayak L. IDO2 in Immunomodulation and Autoimmune Disease. Front Immunol, 2014; 5: 585.
- [11] Haber R, Bessette D, Hulihan-Giblin B, Durcan MJ, Goldman D. Identification of tryptophan 2,3dioxygenase RNA in rodent brain. J Neurochem, 1993; 60: 1159-62.
- [12] Minatogawa Y, Suzuki S, Ando Y, Tone S, Takikawa O. Tryptophan pyrrole ring cleavage enzymes in placenta. Adv Exp Med Biol, 2003; 527: 425-34.
- [13] Batabyal D, Yeh SR. Human tryptophan dioxygenase: a comparison to indoleamine 2,3-dioxygenase. J Am Chem Soc, 2007; 129: 15690-701.
- [14] Malherbe P, Alberati-Giani D, Köhler C, Cesura AM. Identification of a mitochondrial form of kynurenine aminotransferase/glutamine transaminase K from rat brain. FEBS Lett, 1995; 367: 141-4.
- [15] Alberati-Giani D, Malherbe P, Köhler C, Lang G, Kiefer V, Lahm HW, Cesura AM. Cloning and characterization of a soluble kynurenine aminotransferase from rat brain: identity with kidney cysteine conjugate beta-lyase. J Neurochem, 1995; 64: 1448-55.
- [16] Guidetti P, Okuno E, Schwarcz R. Characterization of rat brain kynurenine aminotransferases I and II. J Neurosci Res, 1997; 50: 457-65.
- [17] Han Q, Cai T, Tagle DA, Robinson H, Li J. Substrate specificity and structure of human aminoadipate aminotransferase/kynurenine aminotransferase II. Biosci Rep, 2008; 28: 205-15.
- [18] Han Q, Robinson H, Cai T, Tagle DA, Li J. Biochemical and structural properties of mouse kynurenine aminotransferase III. Mol Cell Biol, 2009; 29: 784-93.
- [19] Guidetti P, Amori L, Sapko MT, Okuno E, Schwarcz R. Mitochondrial aspartate aminotransferase: a third kynurenate-producing enzyme in the mammalian brain. J Neurochem, 2007; 102: 103-11.
- [20] Han Q, Robinson H, Cai T, Tagle DA, Li J. Biochemical and structural characterization of mouse mitochondrial aspartate aminotransferase, a newly identified kynurenine aminotransferase-IV. Biosci Rep, 2011; 31: 323-32.
- [21] Han Q, Cai T, Tagle DA, Li J. Structure, expression, and function of kynurenine aminotransferases in human and rodent brains. Cell Mol Life Sci, 2010; 67: 353-68.
- [22] Lima S, Khristoforov R, Momany C, Phillips RS. Crystal structure of Homo sapiens kynureninase. Biochemistry, 2007; 46: 2735-44.
- [23] Opitz CA, Litzenburger UM, Sahm F, Ott M, Tritschler I, Trump S, Schumacher T, Jestaedt L, Schrenk D, Weller M, Jugold M, Guillemin GJ, Miller CL, Lutz C, Radlwimmer B, Lehmann I, von Deimling A, Wick W, Platten M. An endogenous tumour-promoting ligand of the human aryl hydrocarbon receptor. Nature, 2011; 478: 197-203.
- [24] DiNatale BC, Murray IA, Schroeder JC, Flaveny CA, Lahoti TS, Laurenzana EM, Omiecinski CJ, Perdew GH. Kynurenic acid is a potent endogenous aryl hydrocarbon receptor ligand that synergistically induces interleukin-6 in the presence of inflammatory signaling. Toxicol Sci, 2010; 115: 89-97.

- [25] Lowe MM, Mold JE, Kanwar B, Huang Y, Louie A, Pollastri MP, Wang C, Patel G, Franks DG, Schlezinger J, Sherr DH, Silverstone AE, Hahn ME, McCune JM. Identification of cinnabarinic acid as a novel endogenous aryl hydrocarbon receptor ligand that drives IL-22 production. PLoS One, 2014; 9: e87877.
- [26] Nguyen NT, Nakahama T, Le DH, Van Son L, Chu HH, Kishimoto T. Aryl hydrocarbon receptor and kynurenine: recent advances in autoimmune disease research. Frontiers in Immunology, 2014; 5: 551.
- [27] Kessler M, Terramani T, Lynch G, Baudry M. A glycine site associated with N-methyl-D-aspartic acid receptors: characterization and identification of a new class of antagonists. J Neurochem, 1989; 52: 1319-28.
- [28] Danysz W, Fadda E, Wroblewski JT, Costa E. Kynurenate and 2-amino-5-phosphonovalerate interact with multiple binding sites of the N-methyl-D-aspartate-sensitive glutamate receptor domain. Neurosci Lett, 1989; 96: 340-4.
- [29] Perkins MN, Stone TW. Actions of kynurenic acid and quinolinic acid in the rat hippocampus in vivo. Exp Neurol, 1985; 88: 570-9.
- [30] Patel DR, Young AM, Croucher MJ. Presynaptic alpha-amino-3-hydroxy-5-methyl-4-isoxazole propionate receptor-mediated stimulation of glutamate and GABA release in the rat striatum in vivo: a dual-label microdialysis study. Neuroscience, 2001; 102: 101-11.
- [31] Prescott C, Weeks AM, Staley KJ, Partin KM. Kynurenic acid has a dual action on AMPA receptor responses. Neurosci Lett, 2006; 402: 108-12.
- [32] Rozsa E, Robotka H, Vecsei L, Toldi J. The Janus-face kynurenic acid. J Neural Transm, 2008; 115: 1087-91.
- [33] Hilmas C, Pereira EF, Alkondon M, Rassoulpour A, Schwarcz R, Albuquerque EX. The brain metabolite kynurenic acid inhibits alpha7 nicotinic receptor activity and increases non-alpha7 nicotinic receptor expression: physiopathological implications. J Neurosci, 2001; 21: 7463-73.
- [34] Wang J, Simonavicius N, Wu X, Swaminath G, Reagan J, Tian H, Ling L. Kynurenic acid as a ligand for orphan G protein-coupled receptor GPR35. J Biol Chem, 2006; 281: 22021-8.
- [35] Stone TW, Perkins MN. Quinolinic acid: a potent endogenous excitant at amino acid receptors in CNS. Eur J Pharmacol, 1981; 72: 411-2.
- [36] Fazio F, Lionetto L, Molinaro G, Bertrand HO, Acher F, Ngomba RT, Notartomaso S, Curini M, Rosati O, Scarselli P, Di Marco R, Battaglia G, Bruno V, Simmaco M, Pin JP, Nicoletti F, Goudet C. Cinnabarinic acid, an endogenous metabolite of the kynurenine pathway, activates type 4 metabotropic glutamate receptors. Mol Pharmacol, 2012; 81: 643-56.
- [37] Reyes Ocampo J, Lugo Huitron R, Gonzalez-Esquivel D, Ugalde-Muniz P, Jimenez-Anguiano A, Pineda B, Pedraza-Chaverri J, Rios C, Perez de la Cruz V. Kynurenines with neuroactive and redox properties: relevance to aging and brain diseases. Oxidative Medicine and Cellular Longevity, 2014; 2014: 646909.
- [38] Nichols DE, Nichols CD. Serotonin receptors. Chem Rev, 2008; 108: 1614-41.
- [39] Reppert SM, Weaver DR, Ebisawa T. Cloning and characterization of a mammalian melatonin receptor that mediates reproductive and circadian responses. Neuron, 1994; 13: 1177-85.
- [40] Reppert SM, Godson C, Mahle CD, Weaver DR, Slaugenhaupt SA, Gusella JF. Molecular characterization of a second melatonin receptor expressed in human retina and brain: the Mel1b melatonin receptor. Proc Natl Acad Sci U S A, 1995; 92: 8734-8.
- [41] Dubocovich ML, Delagrange P, Krause DN, Sugden D, Cardinali DP, Olcese J. International Union of Basic and Clinical Pharmacology. LXXV. Nomenclature, classification, and pharmacology of G protein-coupled melatonin receptors. Pharmacol Rev, 2010; 62: 343-80.
- [42] Hardeland R. Melatonin: signaling mechanisms of a pleiotropic agent. Biofactors, 2009; 35: 183-92.
- [43] Wu YH, Ursinus J, Zhou JN, Scheer FA, Ai-Min B, Jockers R, van Heerikhuize J, Swaab DF. Alterations of melatonin receptors MT1 and MT2 in the hypothalamic suprachiasmatic nucleus during depression. J Affect Disord, 2013; 148: 357-67.
- [44] Melke J, Goubran Botros H, Chaste P, Betancur C, Nygren G, Anckarsäter H, Rastam M, Ståhlberg O, Gillberg IC, Delorme R, Chabane N, Mouren-Simeoni MC, Fauchereau F, Durand CM, Chevalier F, Drouot X, Collet C, Launay JM, Leboyer M, Gillberg C, Bourgeron T. Abnormal melatonin synthesis in autism spectrum disorders. Mol Psychiatry, 2008; 13: 90-8.
- [45] Reiter RJ, Tan DX, Manchester LC, Qi W. Biochemical reactivity of melatonin with reactive oxygen and nitrogen species: a review of the evidence. Cell Biochem Biophys, 2001; 34: 237-56.
- [46] Quintela E, Castillo J, Muñoz P, Pascual J. Premonitory and resolution symptoms in migraine: a prospective study in 100 unselected patients. Cephalalgia, 2006; 26: 1051-60.
- [47] Rossi P, Ambrosini A, Buzzi MG. Prodromes and predictors of migraine attack. Funct Neurol, 2005; 20: 185-91.

- [48] Becker WJ. The premonitory phase of migraine and migraine management. Cephalalgia, 2013; 33: 1117-21.
- [49] Houtveen JH, Sorbi MJ. Prodromal functioning of migraine patients relative to their interictal state--an ecological momentary assessment study. PLoS One, 2013; 8: e72827.
- [50] Kelman L. The premonitory symptoms (prodrome): a tertiary care study of 893 migraineurs. Headache, 2004; 44: 865-72.
- [51] Schoonman GG, Evers DJ, Terwindt GM, van Dijk JG, Ferrari MD. The prevalence of premonitory symptoms in migraine: a questionnaire study in 461 patients. Cephalalgia, 2006; 26: 1209-13.
- [52] Giffin NJ, Ruggiero L, Lipton RB, Silberstein SD, Tvedskov JF, Olesen J, Altman J, Goadsby PJ, Macrae A. Premonitory symptoms in migraine: an electronic diary study. Neurology, 2003; 60: 935-40.
- [53] Blin O, Azulay JP, Masson G, Aubrespy G, Serratrice G. Apomorphine-induced yawning in migraine patients: enhanced responsiveness. Clin Neuropharmacol, 1991; 14: 91-5.
- [54] Waelkens J. Dopamine blockade with domperidone: bridge between prophylactic and abortive treatment of migraine? A dose-finding study. Cephalalgia, 1984; 4: 85-90.
- [55] Friedman BW, Corbo J, Lipton RB, Bijur PE, Esses D, Solorzano C, Gallagher EJ. A trial of metoclopramide vs sumatriptan for the emergency department treatment of migraines. Neurology, 2005; 64: 463-8.
- [56] Alstadhaug K, Salvesen R, Bekkelund S. 24-hour distribution of migraine attacks. Headache, 2008; 48: 95-100.
- [57] Kelman L, Rains JC. Headache and sleep: examination of sleep patterns and complaints in a large clinical sample of migraineurs. Headache, 2005; 45: 904-10.
- [58] Bes A, Kunkel R, Lance J, Nappi G, Pfaffenrath V, Rose F, Schoenberg B, Soyka D, Tfelt-Hansen P, Welch K, Wilkinson M, Olesen J, Bousser M, Diener H, Dodick D, First M, Goadsby P, Gobel H, Lainez M, Lipton R, Sakai F, Schoenen J, Silberstein S, Steiner T, Bendtsen L, Ducros A, Evers S, Hershey A, Katsarava Z, Levin M, Pascual J, Russell M, Schwedt T, Tassorelli C, Terwindt G, Vincent M, Wang S, Charles A, Lipton R, Bolay H, Lanteri-Minet M, MacGregor E, Takeshima T, Schytz H, Ashina S, Goicochea M, Hirata K, Holroyd K, Lampl C, Mitsikostas D, Goadsby P, Boes C, Bordini C, Cittadini E, Cohen A, Leone M, May A, Newman L, Pareja J, Park J, Rozen T, Waldenlind E, Fuh J, Ozge A, Pareja J, Peres M, Young W, Yu S, Abu-Arafeh I, Gladstone J, Huang S, Jensen R, Lainez J, Obelieniene D, Sandor P, Scher A, IHS. The International Classification of Headache Disorders, 3rd edition (beta version). Cephalalgia, 2013; 33: 629-808.
- [59] Manzoni GC, Stovner LJ. Epidemiology of headache. Handb Clin Neurol, 2010; 97: 3-22.
- [60] DeLange JM, Cutrer FM. Our evolving understanding of migraine with aura. Curr Pain Headache Rep, 2014; 18: 453.
- [61] Milner PM. Note on a possible correspondence between the scotomas of migraine and spreading depression of Leão. Electroencephalogr Clin Neurophysiol, 1958; 10: 705.
- [62] Hadjikhani N, Sanchez Del Rio M, Wu O, Schwartz D, Bakker D, Fischl B, Kwong KK, Cutrer FM, Rosen BR, Tootell RB, Sorensen AG, Moskowitz MA. Mechanisms of migraine aura revealed by functional MRI in human visual cortex. Proc Natl Acad Sci U S A, 2001; 98: 4687-92.
- [63] Leao A. Spreading depression of activity in the cerebral cortex. J Neurophysiol, 1944; 7: 359-390.
- [64] Zhang X, Levy D, Noseda R, Kainz V, Jakubowski M, Burstein R. Activation of meningeal nociceptors by cortical spreading depression: implications for migraine with aura. J Neurosci, 2010; 30: 8807-14.
- [65] Akcali D, Sayin A, Sara Y, Bolay H. Does single cortical spreading depression elicit pain behaviour in freely moving rats? Cephalalgia, 2010; 30: 1195-206.
- [66] van den Maagdenberg AM, Pietrobon D, Pizzorusso T, Kaja S, Broos LA, Cesetti T, van de Ven RC, Tottene A, van der Kaa J, Plomp JJ, Frants RR, Ferrari MD. A Cacna1a knockin migraine mouse model with increased susceptibility to cortical spreading depression. Neuron, 2004; 41: 701-10.
- [67] Lauritzen M, Rice ME, Okada Y, Nicholson C. Quisqualate, kainate and NMDA can initiate spreading depression in the turtle cerebellum. Brain Res, 1988; 475: 317-27.
- [68] Willette RN, Lysko PG, Sauermelch CF. A comparison of (+)SK&F 10047 and MK-801 on cortical spreading depression. Brain Res, 1994; 648: 347-51.
- [69] Shatillo A, Salo RA, Giniatullin R, Gröhn OH. Involvement of NMDA receptor subtypes in cortical spreading depression in rats assessed by fMRI. Neuropharmacology, 2015; 93: 164-70.
- [70] Olesen J, Burstein R, Ashina M, Tfelt-Hansen P. Origin of pain in migraine: evidence for peripheral sensitisation. Lancet Neurol, 2009; 8: 679-90.
- [71] Ebersberger A, Averbeck B, Messlinger K, Reeh PW. Release of substance P, calcitonin gene-related peptide and prostaglandin E2 from rat dura mater encephali following electrical and chemical stimulation in vitro. Neuroscience, 1999; 89: 901-7.
- [72] Samsam M, Coveñas R, Csillik B, Ahangari R, Yajeya J, Riquelme R, Narváez JA, Tramu G. Depletion of substance P, neurokinin A and calcitonin gene-related peptide from the contralateral and ipsilateral

caudal trigeminal nucleus following unilateral electrical stimulation of the trigeminal ganglion; a possible neurophysiological and neuroanatomical link to generalized head pain. J Chem Neuroanat, 2001; 21: 161-9.

- [73] Williamson DJ, Hargreaves RJ. Neurogenic inflammation in the context of migraine. Microsc Res Tech, 2001; 53: 167-78.
- [74] Knotkova H, Pappagallo M. Imaging intracranial plasma extravasation in a migraine patient: a case report. Pain Med, 2007; 8: 383-7.
- [75] Bell IM. Calcitonin gene-related peptide receptor antagonists: new therapeutic agents for migraine. J Med Chem, 2014; 57: 7838-58.
- [76] Bigal ME, Walter S. Monoclonal antibodies for migraine: preventing calcitonin gene-related peptide activity. CNS Drugs, 2014; 28: 389-99.
- [77] Blau JN, Dexter SL. The site of pain origin during migraine attacks. Cephalalgia, 1981; 1: 143-7.
- [78] Oshinsky ML, Luo J. Neurochemistry of trigeminal activation in an animal model of migraine. Headache, 2006; 46 Suppl 1: S39-44.
- [79] Kai-Kai MA, Howe R. Glutamate-immunoreactivity in the trigeminal and dorsal root ganglia, and intraspinal neurons and fibres in the dorsal horn of the rat. Histochem J, 1991; 23: 171-9.
- [80] Tallaksen-Greene SJ, Young AB, Penney JB, Beitz AJ. Excitatory amino acid binding sites in the trigeminal principal sensory and spinal trigeminal nuclei of the rat. Neurosci Lett, 1992; 141: 79-83.
- [81] Burstein R, Yamamura H, Malick A, Strassman AM. Chemical stimulation of the intracranial dura induces enhanced responses to facial stimulation in brain stem trigeminal neurons. J Neurophysiol, 1998; 79: 964-82.
- [82] Burstein R, Cutrer MF, Yarnitsky D. The development of cutaneous allodynia during a migraine attack clinical evidence for the sequential recruitment of spinal and supraspinal nociceptive neurons in migraine. Brain, 2000; 123 (Pt 8): 1703-9.
- [83] Weiller C, May A, Limmroth V, Jüptner M, Kaube H, Schayck RV, Coenen HH, Diener HC. Brain stem activation in spontaneous human migraine attacks. Nat Med, 1995; 1: 658-60.
- [84] Diener HC. Positron emission tomography studies in headache. Headache, 1997; 37: 622-5.
- [85] Kelman L. The postdrome of the acute migraine attack. Cephalalgia, 2006; 26: 214-20.
- [86] Ng-Mak DS, Fitzgerald KA, Norquist JM, Banderas BF, Nelsen LM, Evans CJ, Healy CG, Ho TW, Bigal M. Key concepts of migraine postdrome: a qualitative study to develop a post-migraine questionnaire. Headache, 2011; 51: 105-17.
- [87] Ophoff RA, Terwindt GM, Vergouwe MN, van Eijk R, Oefner PJ, Hoffman SM, Lamerdin JE, Mohrenweiser HW, Bulman DE, Ferrari M, Haan J, Lindhout D, van Ommen GJ, Hofker MH, Ferrari MD, Frants RR. Familial hemiplegic migraine and episodic ataxia type-2 are caused by mutations in the Ca2+ channel gene CACNL1A4. Cell, 1996; 87: 543-52.
- [88] van den Maagdenberg AM, Haan J, Terwindt GM, Ferrari MD. Migraine: gene mutations and functional consequences. Curr Opin Neurol, 2007; 20: 299-305.
- [89] De Fusco M, Marconi R, Silvestri L, Atorino L, Rampoldi L, Morgante L, Ballabio A, Aridon P, Casari G. Haploinsufficiency of ATP1A2 encoding the Na+/K+ pump alpha2 subunit associated with familial hemiplegic migraine type 2. Nat Genet, 2003; 33: 192-6.
- [90] Dichgans M, Freilinger T, Eckstein G, Babini E, Lorenz-Depiereux B, Biskup S, Ferrari MD, Herzog J, van den Maagdenberg AM, Pusch M, Strom TM. Mutation in the neuronal voltage-gated sodium channel SCN1A in familial hemiplegic migraine. Lancet, 2005; 366: 371-7.
- [91] Silberstein SD, Dodick DW. Migraine genetics: Part II. Headache, 2013; 53: 1218-29.
- [92] Aurora SK, Wilkinson F. The brain is hyperexcitable in migraine. Cephalalgia, 2007; 27: 1442-53.
- [93] Schoenen J, Wang W, Albert A, Delwaide PJ. Potentiation instead of habituation characterizes visual evoked potentials in migraine patients between attacks. Eur J Neurol, 1995; 2: 115-22.
- [94] Ambrosini A, Rossi P, De Pasqua V, Pierelli F, Schoenen J. Lack of habituation causes high intensity dependence of auditory evoked cortical potentials in migraine. Brain, 2003; 126: 2009-15.
- [95] Valeriani M, de Tommaso M, Restuccia D, Le Pera D, Guido M, Iannetti GD, Libro G, Truini A, Di Trapani G, Puca F, Tonali P, Cruccu G. Reduced habituation to experimental pain in migraine patients: a CO(2) laser evoked potential study. Pain, 2003; 105: 57-64.
- [96] Boulloche N, Denuelle M, Payoux P, Fabre N, Trotter Y, Géraud G. Photophobia in migraine: an interictal PET study of cortical hyperexcitability and its modulation by pain. J Neurol Neurosurg Psychiatry, 2010; 81: 978-84.
- [97] Martínez F, Castillo J, Rodríguez JR, Leira R, Noya M. Neuroexcitatory amino acid levels in plasma and cerebrospinal fluid during migraine attacks. Cephalalgia, 1993; 13: 89-93.
- [98] Cananzi AR, D'Andrea G, Perini F, Zamberlan F, Welch KM. Platelet and plasma levels of glutamate and glutamine in migraine with and without aura. Cephalalgia, 1995; 15: 132-5.

- [99] Ferrari MD, Odink J, Bos KD, Malessy MJ, Bruyn GW. Neuroexcitatory plasma amino acids are elevated in migraine. Neurology, 1990; 40: 1582-6.
- [100] González de la Aleja J, Ramos A, Mato-Abad V, Martínez-Salio A, Hernández-Tamames JA, Molina JA, Hernández-Gallego J, Alvarez-Linera J. Higher glutamate to glutamine ratios in occipital regions in women with migraine during the interictal state. Headache, 2013; 53: 365-75.
- [101] Sicuteri F, A. T, F. A. Biochemical Investigations in Headache: Increase in the Hydroxyindoleacetic Acid Excretion During Migraine Attacks. Int Arch Allergy, 1961; 19: 55-58.
- [102] Panconesi A. Serotonin and migraine: a reconsideration of the central theory. J Headache Pain, 2008; 9: 267-76.
- [103] Barter R, Pearse AG. Mammalian enterochromaffin cells as the source of serotonin (5hydroxytryptamine). J Pathol Bacteriol, 1955; 69: 25-31.
- [104] Rapport MM, Green AA, Page IH. Purification of the substance which is responsible for the vasoconstrictor activity of serum. Fed Proc, 1947; 6: 184.
- [105] Chugani DC, Niimura K, Chaturvedi S, Muzik O, Fakhouri M, Lee ML, Chugani HT. Increased brain serotonin synthesis in migraine. Neurology, 1999; 53: 1473-9.
- [106] Sakai Y, Dobson C, Diksic M, Aubé M, Hamel E. Sumatriptan normalizes the migraine attack-related increase in brain serotonin synthesis. Neurology, 2008; 70: 431-9.
- [107] Sakai Y, Nishikawa M, Diksic M, Aubé M. α-[11C] methyl-L tryptophan-PET as a surrogate for interictal cerebral serotonin synthesis in migraine without aura. Cephalalgia, 2014; 34: 165-73.
- [108] Johnson MP, Griffiths LR. A genetic analysis of serotonergic biosynthetic and metabolic enzymes in migraine using a DNA pooling approach. J Hum Genet, 2005; 50: 607-10.
- [109] Erdal N, Herken H, Yilmaz M, Erdal E, Bayazit YA. The A218C polymorphism of tryptophan hydroxylase gene and migraine. J Clin Neurosci, 2007; 14: 249-51.
- [110] Marziniak M, Mössner R, Benninghoff J, Syagailo YV, Lesch KP, Sommer C. Association analysis of the functional monoamine oxidase A gene promotor polymorphism in migraine. J Neural Transm, 2004; 111: 603-9.
- [111] Filic V, Vladic A, Stefulj J, Cicin-Sain L, Balija M, Sucic Z, Jernej B. Monoamine oxidases A and B gene polymorphisms in migraine patients. J Neurol Sci, 2005; 228: 149-53.
- [112] Yang XS, Xu XP, Yang QD. No association of C-1019G promoter polymorphism of 5-HT1A receptor gene with migraine. Eur J Neurol, 2006; 13: 533-5.
- [113] Nyholt DR, Curtain RP, Gaffney PT, Brimage P, Goadsby PJ, Griffiths LR. Migraine association and linkage analyses of the human 5-hydroxytryptamine (5HT2A) receptor gene. Cephalalgia, 1996; 16: 463-7.
- [114] Erdal ME, Herken H, Yilmaz M, Bayazit YA. Association of the T102C polymorphism of 5-HT2A receptor gene with aura in migraine. J Neurol Sci, 2001; 188: 99-101.
- [115] Juhasz G, Zsombok T, Laszik A, Gonda X, Sotonyi P, Faludi G, Bagdy G. Association analysis of 5-HTTLPR variants, 5-HT2a receptor gene 102T/C polymorphism and migraine. J Neurogenet, 2003; 17: 231-40.
- [116] Burnet PW, Harrison PJ, Goodwin GM, Battersby S, Ogilvie AD, Olesen J, Russell MB. Allelic variation in the serotonin 5-HT2C receptor gene and migraine. Neuroreport, 1997; 8: 2651-3.
- [117] Johnson MP, Lea RA, Curtain RP, MacMillan JC, Griffiths LR. An investigation of the 5-HT2C receptor gene as a migraine candidate gene. Am J Med Genet B Neuropsychiatr Genet, 2003; 117B: 86-9.
- [118] Thompson MD, Noble-Topham S, Percy ME, Andrade DM, Ebers GC. Chromosome 1p36 in migraine with aura: association study of the 5HT(1D) locus. Neuroreport, 2012; 23: 45-8.
- [119] Schürks M, Rist PM, Kurth T. STin2 VNTR polymorphism in the serotonin transporter gene and migraine: pooled and meta-analyses. J Headache Pain, 2010; 11: 317-26.
- [120] Liu H, Liu M, Wang Y, Wang XM, Qiu Y, Long JF, Zhang SP. Association of 5-HTT gene polymorphisms with migraine: a systematic review and meta-analysis. J Neurol Sci, 2011; 305: 57-66.
- [121] Ahn AH, Basbaum AI. Where do triptans act in the treatment of migraine? Pain, 2005; 115: 1-4.
- [122] Dussor G. Serotonin, 5HT1 agonists, and migraine: new data, but old questions still not answered. Curr Opin Support Palliat Care, 2014; 8: 137-42.
- [123] Claustrat B, Loisy C, Brun J, Beorchia S, Arnaud JL, Chazot G. Nocturnal plasma melatonin levels in migraine: a preliminary report. Headache, 1989; 29: 242-5.
- [124] Brun J, Claustrat B, Saddier P, Chazot G. Nocturnal melatonin excretion is decreased in patients with migraine without aura attacks associated with menses. Cephalalgia, 1995; 15: 136-9; discussion 79.
- [125] Masruha MR, de Souza Vieira DS, Minett TS, Cipolla-Neto J, Zukerman E, Vilanova LC, Peres MF. Low urinary 6-sulphatoxymelatonin concentrations in acute migraine. J Headache Pain, 2008; 9: 221-4.

- [126] Salvesen R, Bekkelund SI. Migraine, as compared to other headaches, is worse during midnight-sun summer than during polar night. A questionnaire study in an Arctic population. Headache, 2000; 40: 824-9.
- [127] Peres MF, Zukerman E, da Cunha Tanuri F, Moreira FR, Cipolla-Neto J. Melatonin, 3 mg, is effective for migraine prevention. Neurology, 2004; 63: 757.
- [128] Fejes A, Párdutz A, Toldi J, Vécsei L. Kynurenine metabolites and migraine: experimental studies and therapeutic perspectives. Curr Neuropharmacol, 2011; 9: 376-87.
- [129] King RB. Pain and tryptophan. J Neurosurg, 1980; 53: 731.
- [130] Heyliger SO, Goodman CB, Ngong JM, Soliman KF. The analgesic effects of tryptophan and its metabolites in the rat. Pharmacol Res, 1998; 38: 243-50.
- [131] Knyihár-Csillik E, Chadaide Z, Okuno E, Krisztin-Péva B, Toldi J, Varga C, Molnár A, Csillik B, Vécsei L. Kynurenine aminotransferase in the supratentorial dura mater of the rat: effect of stimulation of the trigeminal ganglion. Exp Neurol, 2004; 186: 242-7.
- [132] Vécsei L, Szalárdy L, Fülöp F, Toldi J. Kynurenines in the CNS: recent advances and new questions. Nat Rev Drug Discov, 2013; 12: 64-82.
- [133] Fukui S, Schwarcz R, Rapoport SI, Takada Y, Smith QR. Blood-brain barrier transport of kynurenines: implications for brain synthesis and metabolism. J Neurochem, 1991; 56: 2007-17.
- [134] Knyihár-Csillik E, Toldi J, Krisztin-Péva B, Chadaide Z, Németh H, Fenyo R, Vécsei L. Prevention of electrical stimulation-induced increase of c-fos immunoreaction in the caudal trigeminal nucleus by kynurenine combined with probenecid. Neurosci Lett, 2007; 418: 122-6.
- [135] Cunningham RF, Israili ZH, Dayton PG. Clinical pharmacokinetics of probenecid. Clin Pharmacokinet, 1981; 6: 135-51.
- [136] Vámos E, Párdutz A, Varga H, Bohár Z, Tajti J, Fülöp F, Toldi J, Vécsei L. 1-kynurenine combined with probenecid and the novel synthetic kynurenic acid derivative attenuate nitroglycerin-induced nNOS in the rat caudal trigeminal nucleus. Neuropharmacology, 2009; 57: 425-9.
- [137] Knyihár-Csillik E, Toldi J, Mihály A, Krisztin-Péva B, Chadaide Z, Németh H, Fenyo R, Vécsei L. Kynurenine in combination with probenecid mitigates the stimulation-induced increase of c-fos immunoreactivity of the rat caudal trigeminal nucleus in an experimental migraine model. J Neural Transm, 2007; 114: 417-21.
- [138] de Tommaso M, Libro G, Guido M, Difruscolo O, Losito L, Sardaro M, Cerbo R. Nitroglycerin induces migraine headache and central sensitization phenomena in patients with migraine without aura: a study of laser evoked potentials. Neurosci Lett, 2004; 363: 272-5.
- [139] Tassorelli C, Greco R, Morocutti A, Costa A, Nappi G. Nitric oxide-induced neuronal activation in the central nervous system as an animal model of migraine: mechanisms and mediators. Funct Neurol, 2001; 16: 69-76.
- [140] Pardutz A, Krizbai I, Multon S, Vecsei L, Schoenen J. Systemic nitroglycerin increases nNOS levels in rat trigeminal nucleus caudalis. Neuroreport, 2000; 11: 3071-5.
- [141] Pardutz A, Hoyk Z, Varga H, Vecsei L, Schoenen J. Oestrogen-modulated increase of calmodulindependent protein kinase II (CamKII) in rat spinal trigeminal nucleus after systemic nitroglycerin. Cephalalgia, 2007; 27: 46-53.
- [142] Pardutz A, Multon S, Malgrange B, Parducz A, Vecsei L, Schoenen J. Effect of systemic nitroglycerin on CGRP and 5-HT afferents to rat caudal spinal trigeminal nucleus and its modulation by estrogen. Eur J Neurosci, 2002; 15: 1803-9.
- [143] Vámos E, Fejes A, Koch J, Tajti J, Fülöp F, Toldi J, Párdutz A, Vécsei L. Kynurenate derivative attenuates the nitroglycerin-induced CamKIIα and CGRP expression changes. Headache, 2010; 50: 834-43.
- [144] Fejes-Szabó A, Bohár Z, Vámos E, Nagy-Grócz G, Tar L, Veres G, Zádori D, Szentirmai M, Tajti J, Szatmári I, Fülöp F, Toldi J, Párdutz Á, Vécsei L. Pre-treatment with new kynurenic acid amide dose-dependently prevents the nitroglycerine-induced neuronal activation and sensitization in cervical part of trigemino-cervical complex. J Neural Transm, 2014; 121: 725-38.
- [145] Chauvel V, Vamos E, Pardutz A, Vecsei L, Schoenen J, Multon S. Effect of systemic kynurenine on cortical spreading depression and its modulation by sex hormones in rat. Exp Neurol, 2012; 236: 207-14.
- [146] Oláh G, Herédi J, Menyhárt A, Czinege Z, Nagy D, Fuzik J, Kocsis K, Knapp L, Krucsó E, Gellért L, Kis Z, Farkas T, Fülöp F, Párdutz A, Tajti J, Vécsei L, Toldi J. Unexpected effects of peripherally administered kynurenic acid on cortical spreading depression and related blood-brain barrier permeability. Drug Des Devel Ther, 2013; 7: 981-7.
- [147] Kageneck C, Nixdorf-Bergweiler BE, Messlinger K, Fischer MJ. Release of CGRP from mouse brainstem slices indicates central inhibitory effect of triptans and kynurenate. J Headache Pain, 2014; 15: 7.

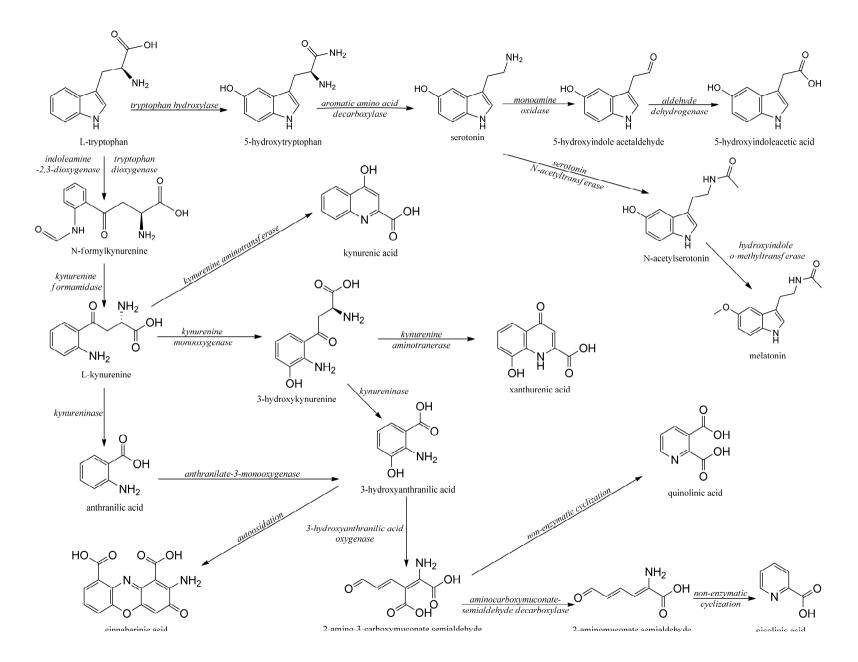


Figure 1. Metabolic routes of tryptophan

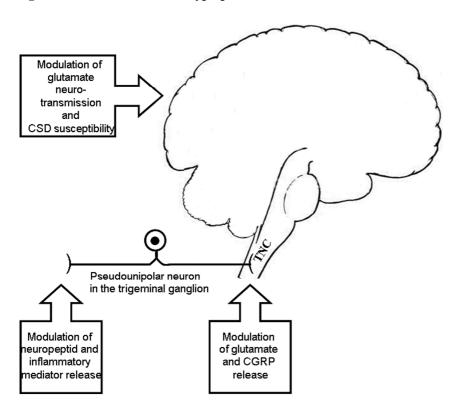


Figure 2. Possible routes of action of kynurenic acid and its derivatives