

Review

Glutamatergic Dysfunctioning in Alzheimer's Disease and Related Therapeutic Targets

Dénes Zádori^a, Gábor Veres^a, Levente Szalárdy^a, Péter Klivényi^a, József Toldi^{b,c} and László Vécsei^{a,c,*}

^a*Department of Neurology, Faculty of Medicine, Albert Szent-Györgyi Clinical Center, University of Szeged, Szeged, Hungary*

^b*Department of Physiology, Anatomy and Neuroscience, Faculty of Science and Informatics, University of Szeged, Szeged, Hungary*

^c*MTA-SZTE Neuroscience Research Group, Szeged, Hungary*

Accepted 13 February 2014

Abstract. The impairment of glutamatergic neurotransmission plays an important role in the development of Alzheimer's disease (AD). The pathological process, which involves the production of amyloid- β peptides and hyperphosphorylated tau proteins, spreads over well-delineated neuroanatomical circuits. The gradual deterioration of proper synaptic functioning (via GluN2A-containing N-methyl-D-aspartate receptors, NMDARs) and the development of excitotoxicity (via GluN2B-containing NMDARs) in these structures both accompany the disease pathogenesis. Although one of the most important therapeutic targets would be glutamate excitotoxicity, the application of conventional anti-glutamatergic agents could result in further deterioration of synaptic transmission and intolerable side-effects. With regard to NMDAR antagonists with tolerable side-effects, ion channel blockers with low affinity, glycine site agents, and specific antagonists of polyamine site and GluN2B subunit may come into play. However, in the mirror of experimental data, only the application of ion channel blockers with pronounced voltage dependency, low affinity, and rapid unblocking kinetics (e.g., memantine) and specific antagonists of the GluN2B subunit (e.g., ifenprodil and certain kynurenic acid amides) resulted in desirable symptom amelioration. Therefore we propose that these kinds of chemical agents may have therapeutic potential for present and future drug development.

Keywords: Alzheimer's disease, glutamate excitotoxicity, kynurenic acid amides, memantine, neurodegeneration, neuroprotection, therapy

INTRODUCTION

Alzheimer's disease (AD) is a progressive neurodegenerative disorder, the main clinical feature of which is dementia [1, 2]. Indeed, AD is the most common type among dementia syndromes [3] and is responsible for 60–80% of the cases [4], leading to a considerable

socioeconomic burden. Although clinical diagnosis can be determined during the disease course in most cases, currently autopsy is necessary for a definite diagnosis. The main pathological hallmark of AD is the presence of neurofibrillary tangles (NFTs) and senile plaques in specific brain areas [5]. With regard to the involvement of dysfunctional neurotransmission in disease pathogenesis, certain cholinergic and glutamatergic systems are the most affected [6, 7].

The aim of this short review is to highlight aspects of glutamatergic dysfunction in AD and to discuss some possibilities of pharmaceutical interventions by targeting the glutamatergic system.

*Correspondence to: László Vécsei MD, PhD, DSc, Department of Neurology, Faculty of Medicine, Albert Szent-Györgyi Clinical Center, University of Szeged, Semmelweis u. 6, H-6725 Szeged, Hungary. Tel.: +36(62)545351; Fax: +36(62)545597; E-mail: vecsei.laszlo@med.u-szeged.hu.

ALTERATIONS IN GLUTAMATERGIC SIGNALING IN ALZHEIMER'S DISEASE: PATHOLOGICAL BASIS

With regard to the sensitivity and specificity for the diagnosis of AD, the Braak staging system [5] gives the best accuracy (79%) among the neuropathological criteria systems [8]. This system classifies AD into stages mainly by the temporal evolution of NFTs (composed of intracellular aggregates of hyperphosphorylated tau protein), but it also takes into account the loci of extracellular amyloid- β (A β) deposits in the brain. The system distinguishes between the following stages: transentorhinal/entorhinal (stage I, II), limbic (stage III, IV), and neocortical (stage V, VI). This classification shows a good correlation with the severity of dementia [9], though originally the pathological stages were established by Braak irrespective of the clinical stage of the dementia. Certain neuropathological investigations have special significance in the assessment of early stages of AD [10]. The most important ones include the assessment of NFTs in the neurons of the second layer of the entorhinal cortex in the slices of the inferior temporal lobe. The entorhinal cortex receives converging polysynaptic glutamatergic inputs from the multimodal association cortices and limbic areas including the hippocampal formation, while it projects into the hippocampal formation and back to the association cortices [11–13]. One of the main efferent glutamatergic projections of the entorhinal cortex is the perforant pathway, which predominantly originates from the second layer and serves as the main excitatory input of the hippocampal formation. The fourth layer of the entorhinal cortex in turn receives excitatory input from the hippocampal formation. A significant decrease was observed in the neuronal number of the fourth and especially the second layers of the entorhinal cortex in clinically very mild AD [14]. Another study likewise demonstrated a considerable decrease in neuronal number and volume of the entorhinal cortex (especially the second layer) and those of the cornu ammonis (CA)1 region of the hippocampus in preclinical AD cases [15]. It is important to mention that the presence of NFTs can also be observed in these early stages in the CA1-subiculum part of the hippocampal formation and in the perirhinal cortex, inferior temporal gyrus, amygdala, posterior part of the parahippocampal gyrus, the cholinergic basal forebrain and in the dorsal raphe nuclei, but in a lesser extent compared to the second layer of the entorhinal cortex [16]. In the next stages, almost all the limbic structures, notably the hippocampal forma-

tion (consisting of the dentate gyrus, the hippocampus proper, and the subiculum) and the amygdala become considerably damaged [17] in addition to the more expressed involvement of the previously described brain structures. As partially mentioned above, the main glutamatergic input of the hippocampal formation comes from the second (toward the dentate gyrus) and the third (toward the subiculum and CA1 sector of the hippocampus proper) layers of the entorhinal cortex via the perforant and temporo-alvear pathways [18]. Scheff et al. [19] hypothesized that synaptic loss in the outer molecular layer (OML) of the dentate gyrus would be responsible for the transition from mild cognitive impairment to early AD. Total synaptic counts in the OML had a significant negative correlation with NFT density in the entorhinal cortex. Although there was a negative correlation between the individual's Braak score and total synaptic number in the OML, this association was not significant and furthermore, this study did not find significant correlation of Braak staging with the scores of any of the applied psychometric tests. However, a high positive correlation of total synaptic number in the OML with the values of tests of cognitive functions such as the Mini-Mental State Examination and delayed memory recall (one of the most sensitive measures of hippocampal function) was demonstrated, which suggests that synaptic loss would be one of the strongest predictive factors for cognitive decline. As a part of the trisynaptic circuit, the information is transmitted further from the dentate gyrus via intrahippocampal association pathways (via mossy fibers toward the CA3 sector of the hippocampus, and then via Schaffer collaterals toward the CA1 sector) [20]. The synaptic loss can also be observed in the CA1 sector of the hippocampus in mild AD cases [21]. The pyramidal cells of the CA1 sector predominantly innervate the subiculum, which projects to the pre/parasubiculum (parts of the subicular complex which also receives neocortical inputs likewise the entorhinal cortex), the amygdala, the fourth layer of the entorhinal cortex, the anterior and midline thalamic and mammillary nuclei (via the fornix) [22]. Regarding further parts of the Papez circuit, the information processes from the mammillary nuclei to the anterior thalamic nuclei (via the mammillothalamic tract) and further to the cingulate gyrus (via the anterior thalamic radiation) and to the presubiculum (via the cingulum), which projects to the fourth layer of the entorhinal cortex [23]. The pre/parasubiculum also send minor projections to the dentate gyrus [24]. It is important to mention that parts of the hippocampal formation in the two hemispheres

are strongly interconnected via commissural fibers. The amygdaloid complex, which consists of distinct nuclei, receives inputs from multiple brain regions via several kinds of transmitter systems, including glutamatergic pathways [25]. The major sources of sensory and polymodal information to the amygdala are certain parts of the cerebral cortex, including the association and prefrontal cortices [26]. The amygdala also forms reciprocal and strong connections with areas related to long-term declarative memory system, including the perirhinal and entorhinal cortices and the hippocampal formation [27]. Furthermore, the amygdaloid complex has widespread projections to certain cortical, subcortical, and brainstem structures [25]. The key feature of advanced stages of AD (stage V-VI) is the occurrence of severe destruction of neocortical association areas [28, 29]. Although NFT pathology only becomes expressed in advanced stages of AD in neocortical areas, the alteration in the level of some molecular markers of synaptic dysfunctioning can be observed even in early stages of AD. Accordingly, vesicular glutamate transporter (VGLUT)1 expression is found to be decreased in the prefrontal, parietal and occipital and inferior temporal cortices, while it was unaltered in the lateral temporal cortex [30–32]. With regard to the murine models of AD, a significant reduction of VGLUT1 was observed in both the frontal cortex and the hippocampus [33, 34]. The expression of VGLUT2 and synaptophysin was altered only in the prefrontal cortex in human AD cases [30]. Loss of VGLUT1 and VGLUT2 in the prefrontal cortex correlated with cognitive status even at early phases of cognitive decline [30]. Although the typical spreading of neuropathological alterations over the above-mentioned glutamatergic structures with strong connections (Fig. 1) can be well observed in most cases, limbic-predominant and hippocampal-sparing subtypes of AD cases were also reported [35].

ALTERATIONS IN GLUTAMATERGIC SIGNALING IN ALZHEIMER'S DISEASE: MOLECULAR BASIS

The main culprits responsible for the disconnection of the previously delineated glutamatergic networks would be the A β peptide and the tau protein [36]. A β ₁₋₄₂ aggregates are capable of inducing tau hyperphosphorylation [36] and promote *in vitro* tau aggregation in a dose-dependent manner [37]. In addition to NFTs, soluble tau also would have neurotoxic properties [38]. A β can influence gluta-

matergic neurotransmission in several ways. Although under physiological concentrations, endogenous A β is necessary for proper neurotransmitter release [39], in excess it weakens synaptic transmission affecting the synaptic vesicle pools [40]. Accordingly, A β is co-localized in glutamatergic boutons immunoreactive for VGLUT1 and VGLUT2 in postmortem AD brains [41]. Furthermore, soluble A β oligomers induce the disruption of dendritic spines, resulting in severe neuropil damage [42]. The degeneration of synapses and dendritic spines is one of the earliest feature of AD [43]. Glutamatergic synapses contain α -amino-3-hydroxy-5-methyl-4-isoxazolepropionic acid receptors (AMPA) and N-methyl-D-aspartate receptors (NMDARs) localized on dendritic spines. The basal synaptic transmission is mainly mediated by AMPARs. However, in view of receptor dysfunction in AD, the NMDAR would be the major site of A β action, and in turn, NMDAR activation enhances A β production [44]. A conventional NMDAR is composed of two glycine or D-serine-binding GluN1 and 2 glutamate-binding GluN2 (A-D) subunits, forming a heterotetramer. The GluN1 subunits form the ion channel, while the GluN2 subunits have more of a regulatory and refining role. It has been shown that the GluN2B subunit-containing NMDARs predominate at the extrasynaptic site [45], which preferential localization becomes more predominant by the phosphorylation at Tyr1336 [46]. Oligomeric A β promotes Fyn kinase activation via binding to the post-synaptic prion protein (PrP^C), resulting in the increased phosphorylation of the GluN2B subunits at Tyr1472 [47]. This activation induces altered NMDAR localization with destabilization of dendritic spines and the loss of surface NMDARs. It is important to mention that several other receptors are regulated by PrP^C, including metabotropic glutamate receptor (mGluR) 1 and 5 [48]. The available data suggest that the activation of NMDARs at the synaptic site promotes neuronal survival, while activation at the extrasynaptic site mediates neurotoxic effects [49]. However, some recent findings suggest that the simultaneous activation of synaptic NMDARs are also necessary for the initiation of cell death program [50]. So in brief, the inactivation of glutamatergic synaptic transmission and the activation of that at the extrasynaptic sites would both accompany the pathomechanism of AD. Oligomeric A β impairs long-term potentiation (LTP; a form of synaptic strengthening following brief, high frequency stimulation [51]) and enhances long-term depression (LTD; a form of synaptic weakening following low frequency stimulation

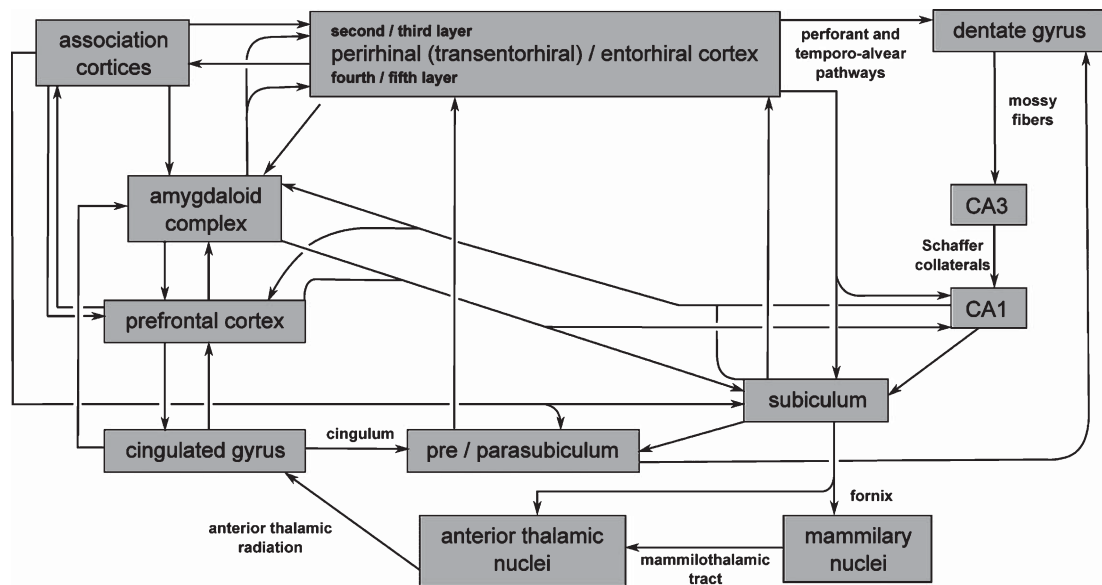


Fig. 1. The schematic depiction of the predominant connections between the affected glutamatergic brain areas in Alzheimer's disease. (CA, cornu ammonis).

247 or synaptic inactivity [52]) and the depotentiation
 248 of LTP, thereby causing synaptic dysfunctioning
 249 [53, 54]. Oligomeric A β -induced internalization of
 250 synaptic AMPARs and NMDARs [55, 56] and non-
 251 apoptotic caspase activation [57] both accompany LTD
 252 enhancement. Although several forms of synaptic plas-
 253 ticity depend on NMDAR-driven calcium flux [58],
 254 some recent data indicate that A β -mediated synap-
 255 tic AMPAR depression requires NMDAR activation
 256 in a metabotropic manner, i.e., without ion flow via
 257 the NMDAR [59]. NMDARs also have an important
 258 role in spontaneous glutamate release-induced depres-
 259 sion of evoked neurotransmission, thereby influencing
 260 synaptic efficacy as well [60]. In addition to the demon-
 261 strated alteration of glutamatergic neurotransmission
 262 via postsynaptic and extrasynaptic NMDARs in AD,
 263 recent experimental data provide increasing evidence
 264 of the involvement of presynaptic NMDARs in the
 265 enhancement of timing-dependent LTD, resulting in
 266 impaired memory functions, which phenomenon may
 267 have implications in the development of cognitive
 268 decrement in AD [61–63]. With regard to caspase-
 269 3 activation, the increased activity of the pyramidal
 270 neurons of the entorhinal cortex, the subiculum, and
 271 the CA1-3 sector of the hippocampus was found in
 272 early stages of AD [64]. The second layer of the
 273 entorhinal cortex showed the highest activity. A β accu-
 274 mulation activates NMDARs at early stages of AD
 275 [65], and *in vitro* studies suggest that this activation
 276 might be mediated by GluN2B-containing NMDARs

[66]. It has been also demonstrated that NMDARs
 are connected to neuronal nitric oxide synthase by a
 scaffolding protein PSD-95 (postsynaptic density pro-
 tein of molecular weight 95 kDa), which binds to the
 GluN2B subunit of the NMDAR [67]. Thus, PSD-
 95 would have an important role in the evocation of
 downstream excitotoxic events mediated by GluN2B
 subunit-containing NMDARs via the production of
 nitric oxide in an excessive amount [68]. Recent data
 indicate that the activation of NMDARs by A β ₁₋₄₂ may
 be secondary to its binding to postsynaptic anchoring
 proteins such as PSD-95 [42]. Extrasynaptic NMDAR
 activation triggers the increased production of A β due
 to the shift of amyloid β -protein precursor (A β PP)
 production from A β PP695 to Kunitz protease inhibitory
 domain-containing isoforms with higher amyloido-
 genic potential [69]. This kind of positive feedback
 leads to the formation of a vicious circle [70]. GluN2B-
 mediated neurotransmission also seems to be involved
 in tau-induced neurotoxicity [71]. Tau phosphorylation
 causes tau mislocalization and subsequent synaptic
 impairment as phosphorylated tau can accumulate in
 dendritic spines, where it may affect the synaptic traf-
 ficking and/or anchoring of glutamate receptors [72].
 The interaction of tau with fyn targets fyn to dendritic
 spines, where it can exert the above-mentioned phos-
 phosphorylation of GluN2B subunit of NMDAR, thereby
 enhancing the excitotoxic process [73]. In addition to
 its neuronal effects, A β also downregulates glutamate
 uptake capacity of astrocytes and thereby induces a

277
 278
 279
 280
 281
 282
 283
 284
 285
 286
 287
 288
 289
 290
 291
 292
 293
 294
 295
 296
 297
 298
 299
 300
 301
 302
 303
 304
 305
 306

dysfunctional extracellular glutamate clearance [74]. Besides the elevated levels of glutamate in the extracellular space, the presence of an energy impairment, as a consequence of mitochondrial dysfunction and oxidative stress, would be another causative factor in glutamate excitotoxicity, which leads to a partial membrane depolarization resulting in relief of the Mg^{2+} blockade of the NMDAR channel and calcium overload [75].

THERAPEUTIC APPROACHES TARGETING THE GLUTAMATERGIC NEUROTRANSMISSION SYSTEM WITH A SPECIAL VIEW OF NMDA RECEPTORS IN ALZHEIMER'S DISEASE: PITFALLS AND POSSIBILITIES

The application of agents that completely block NMDAR activity has limited usefulness due to severe clinical side-effects such as hallucinations, agitation, memory impairment, catatonia, nausea, vomiting, a peripheral sensory disturbance, and sympathomimetic effects such as increased blood pressure [76, 77]. In order to achieve neuroprotection by targeting the NMDARs in AD, the best therapeutic strategy could be the normalization of synaptic GluN1/GluN2A activity and the abolishment of excitotoxicity mediated by extrasynaptic GluN1/GluN2B subunits. In view of NMDAR antagonists with tolerable side-effects, ion channel blockers with lower affinity, glycine site agents as well as specific antagonists of the polyamine site or the GluN2B subunit may come into play (Fig. 2) [78]. Memantine (3,5-dimethyladamantan-1-amine) is a low affinity open channel blocker, which preferentially antagonizes excessively activated NMDARs without affecting physiological NMDAR activity [79]. Accordingly, this substance has recently been demonstrated to selectively target mainly GluN2B-containing extrasynaptic NMDARs [80], i.e., it is three times more potent in the inhibition of calcium influx via GluN1/GluN2B than via GluN1/GluN2A subunit-containing NMDARs [81]. Furthermore, memantine concentration-dependently inhibited the expression of Kunitz protease inhibitory domain-containing A β PP isoforms as well as neuronal production and release of A β [69, 82]. Accordingly, memantine is a widely applied medicament in the treatment of moderate-advanced stages of AD with beneficial effects as regards language, memory, praxis, and communication dysfunction as well as the activities of daily living [83]. Although memantine has some potential side-effects

such as somnolence, weight gain, confusion, hypertension, nervous system disorders, and falling [84], to date this is the only commercially available NMDAR antagonist in the treatment of AD. In summary, the good effect/side-effect profile would be explained by its pronounced voltage dependency, low affinity, and rapid unblocking kinetics, properties which make the restoration of the desired signal-to-noise ratio in glutamatergic neurotransmission available [85].

Kynurenic acid (KYNA; produced by kynurenine aminotransferases, KATs), a side-product of the main pathway of the tryptophan metabolism, can influence glutamatergic neurotransmission at several levels [86], and exerted neuroprotective effects in several paradigms [86–90]. On the one hand, KYNA can exert wide-spectrum endogenous antagonism of ionotropic excitatory amino acid receptors [91], mainly targeting the strychnine-insensitive glycine-binding site on the GluN1 subunit of the NMDA receptor [92]. This action requires relatively high (~ 10 – $20 \mu M$) concentrations of KYNA under physiological conditions [93]; the basal extracellular concentration of KYNA in rats (15–23 nM) [94, 95] is far below the required level to directly interfere with glutamate receptor functions. Accordingly, only excessive elevation of the KYNA level could be accompanied by adverse effects in rats, such as reduced exploratory activity, ataxia, stereotypy, sleeping, and respiratory depression, while there was only a slight effect on the learning ability [96]. However, human postmortem analyses revealed elevated levels of KYNA in the striatum and hippocampus of AD patients [97], alteration of which is suggested to accompany to the cognitive dysfunction in AD rather than to exert a compensatory protective role. Accordingly, the achievement of lowering brain KYNA levels by knocking out one of its producing enzyme (KAT II) resulted in the improvement of cognitive functions in mice [98]. With regard to the mechanisms of influencing glutamatergic transmission, on the other hand, KYNA non-competitively blocks the $\alpha 7$ -nicotinic acetylcholine receptors [99], thereby inhibiting glutamate release at the presynaptic site [100]. This blockade can be effective at high nanomolar concentrations ($IC_{50} = \sim 7 \mu M$), and can also influence hippocampus-dependent cognitive functions [101]. In addition to the multiplex receptor antagonism, recent studies showed that KYNA is capable of facilitating AMPA receptor responses in nanomolar concentrations [102, 103]. The significance of this phenomenon is not really known yet.

The selective inhibition of GluN2B subunit-containing NMDARs could be another successful

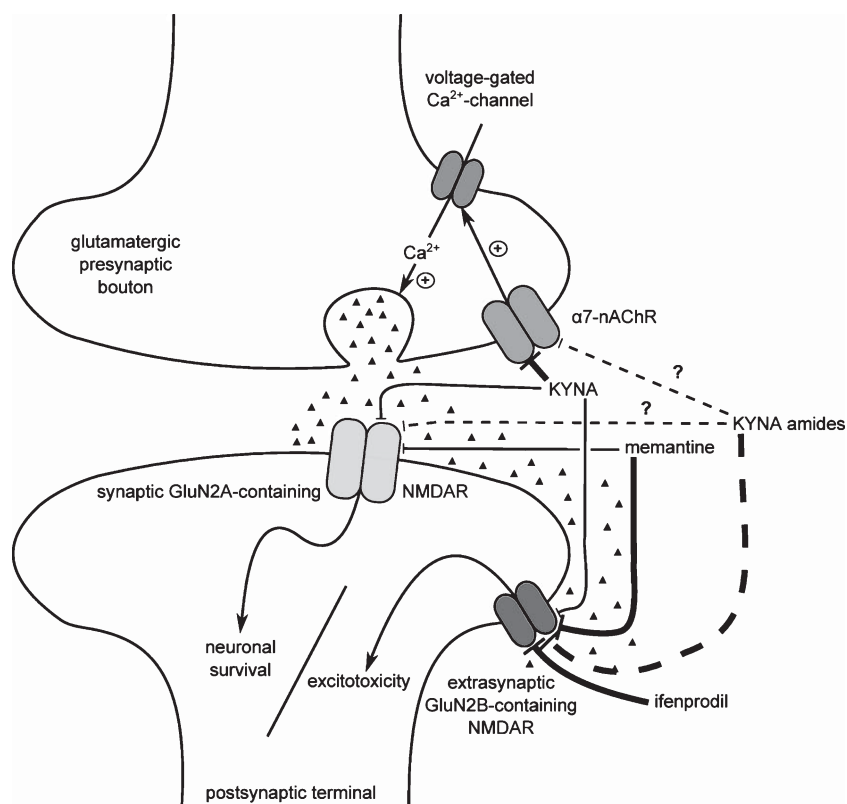


Fig. 2. Some possibilities of influencing glutamatergic dysfunctioning in Alzheimer's disease. ($\alpha 7$ -nAChR, alpha7-nicotinic acetylcholine receptors; KYNA, kynurenic acid; NMDAR, N-methyl-D-aspartate receptor; ▲, glutamate, the thickness of the lines represents the extent of inhibition, while dashed lines refers to possible mechanism of action).

408 strategy in the amelioration of neurodegenerative
 409 processes [104]. Ifenprodil (α -(4-hydroxyphenyl)- β -
 410 methyl-4-benzyl-1-piperidineethanol) is a synthetic
 411 negative allosteric modulator of such of receptors,
 412 with relatively high affinity ($IC_{50} = \sim 150$ nM) [105].
 413 Ifenprodil binding seems to interact with polyamine
 414 binding in a negative allosteric manner, i.e., it can
 415 inhibit the potentiation of NMDAR currents evoked
 416 by certain polyamines [106, 107]. It has a consid-
 417 erably good side-effect profile: only mouth dryness,
 418 nausea, headache, and palpitations were observed.
 419 Accordingly, several derivatives, including Ro 25-
 420 6981 ([R-(R*,S*)]- α -(4-hydroxyphenyl)- β -methyl-4-
 421 benzyl-1-piperidinepropanol), have been synthesized
 422 with the aim of presenting lead compounds in pharma-
 423 ceutical development in the field of neurodegenerative
 424 disorders [104]. With regard to AD, $A\beta$ -induced endo-
 425 plasmic reticulum and oxidative stress was prevented
 426 by ifenprodil [108]. Furthermore, this substance and
 427 Ro 25-6981 also prevented the $A\beta$ -mediated inhibi-
 428 tion of LTP in rodent hippocampal slices [109–112].
 429 Indeed, Ro 25-6981 abolished LTD enhancement and

430 learning impairment in rats as well [113]. Evotec's
 431 EVT 101, another GluN2B antagonist which has been
 432 shown to penetrate into the human brain, was well
 433 tolerated in a double-blind, 4-week phase Ib study
 434 (<http://www.evotec.com>).

435 A possible pharmaceutical modification of KYNA
 436 is amidation at the carboxyl moiety [114, 115].
 437 The resulting KYNA amides may be of special
 438 interest since they have been shown to preferen-
 439 tially act on GluN2B subunit-containing extrasynaptic
 440 NMDARs [116]. This feature may also offer the
 441 opportunity to establish an extracellular concen-
 442 tration that is capable of inhibiting the tonic
 443 extrasynaptic NMDAR currents without impairing
 444 synaptic glutamatergic neurotransmission. Accord-
 445 ingly, one of the KYNA amide compounds synthesized
 446 by our group, N-(2-N,N-dimethylaminoethyl)-4-oxo-
 447 1H-quinoline-2-carboxamide hydrochloride exerted
 448 protective effects both in the four-vessel occlu-
 449 sion model of cerebral ischemia (rats; [117]) and
 450 in the N171-82Q transgenic mouse model of HD
 451 [118].

Finally, in addition to directly influencing NMDARs, it is important to mention that there are some indirect regulators of NMDAR functioning, targeting of which can be used as alternative therapeutic approaches in the amelioration of glutamatergic dysfunction in AD. These targets include some metabotropic glutamatergic receptors [119] and certain adenosine receptors [120, 121].

CONCLUSION

Although more and more details are being revealed regarding the pathomechanism of AD, the recent therapeutic strategies are restricted only to few pharmaceutical agents. The glutamatergic system is presumed to be the major altered neurotransmitter system in AD; therefore, there is a great need for the development of pharmacokons targeting this system with acceptable side-effect profile. From this respect, ion channel blockers with lower affinity as well as GluN2B subunit specific antagonists might be the most promising candidates for future AD therapy. Although the present short review focused on the possibilities of therapeutic amelioration via targeting the glutamatergic neurotransmission system with special attention to NMDARs, it should be noted that achieving neuroprotection in AD—especially in terms of ‘synaptoprotection’—is a complex issue, with pharmacological targets and approaches we could not detail here, but have already been comprehensively discussed by others [122, 123].

ACKNOWLEDGMENTS

This work was supported by the projects OTKA (K 75628), KTIA_NAP_13 – Hungarian National Brain Research Program and TÁMOP-4.2.2.A-11/1/KONV-2012-0052. Furthermore, this research was realized in the frames of TÁMOP 4.2.4. A/1-11-1-2012-0001 “National Excellence Program – Elaborating and operating an inland student and researcher personal support system”. The project was subsidized by the European Union and co-financed by the European Social Fund.

Authors’ disclosures available online (<http://www.j-alz.com/disclosures/view.php?id=2158>).

REFERENCES

- [1] Blennow K, de Leon MJ, Zetterberg H (2006) Alzheimer’s disease. *Lancet* **368**, 387-403.
- [2] Zádori D, Datki Z, Penke B (2007) [The role of chronic brain hypoperfusion in the pathogenesis of Alzheimer’s

- disease—facts and hypotheses]. *Ideggyogy Sz* **60**, 428-437.
- [3] Scott KR, Barrett AM (2007) Dementia syndromes: Evaluation and treatment. *Expert Rev Neurother* **7**, 407-422.
- [4] Hebert LE, Scherr PA, Bienias JL, Bennett DA, Evans DA (2003) Alzheimer disease in the US population: Prevalence estimates using the 2000 census. *Arch Neurol* **60**, 1119-1122.
- [5] Braak H, Braak E (1991) Neuropathological stageing of Alzheimer-related changes. *Acta Neuropathol* **82**, 239-259.
- [6] Vogels OJ, Broere CA, ter Laak HJ, ten Donkelaar HJ, Nieuwenhuys R, Schulte BP (1990) Cell loss and shrinkage in the nucleus basalis Meynert complex in Alzheimer’s disease. *Neurobiol Aging* **11**, 3-13.
- [7] Neill D (1995) Alzheimer’s disease: Maladaptive synaptoplasticity hypothesis. *Neurodegeneration* **4**, 217-232.
- [8] Geddes JW, Tekirian TL, Soultanian NS, Ashford JW, Davis DG, Markesbery WR (1997) Comparison of neuropathologic criteria for the diagnosis of Alzheimer’s disease. *Neurobiol Aging* **18**, S99-105.
- [9] Mortimer JA, Borenstein AR, Gosche KM, Snowdon DA (2005) Very early detection of Alzheimer neuropathology and the role of brain reserve in modifying its clinical expression. *J Geriatr Psychiatry Neurol* **18**, 218-223.
- [10] Perl DP, Purohit DP (1997) Proposal to revise the morphologic criteria for the diagnosis of Alzheimer’s disease. *Neurobiol Aging* **18**, S81-S84.
- [11] Van Hoesen G, Pandya DN (1975) Some connections of the entorhinal (area 28) and perirhinal (area 35) cortices of the rhesus monkey. I. Temporal lobe afferents. *Brain Res* **95**, 1-24.
- [12] Van Hoesen G, Pandya DN, Butters N (1975) Some connections of the entorhinal (area 28) and perirhinal (area 35) cortices of the rhesus monkey. II. Frontal lobe afferents. *Brain Res* **95**, 25-38.
- [13] Van Hoesen GW, Pandya DN (1975) Some connections of the entorhinal (area 28) and perirhinal (area 35) cortices of the rhesus monkey. III. Efferent connections. *Brain Res* **95**, 39-59.
- [14] Gomez-Isla T, Price JL, McKeel DW Jr, Morris JC, Growdon JH, Hyman BT (1996) Profound loss of layer II entorhinal cortex neurons occurs in very mild Alzheimer’s disease. *J Neurosci* **16**, 4491-4500.
- [15] Price JL, Ko AI, Wade MJ, Tsou SK, McKeel DW, Morris JC (2001) Neuron number in the entorhinal cortex and CA1 in preclinical Alzheimer disease. *Arch Neurol* **58**, 1395-1402.
- [16] Arriagada PV, Growdon JH, Hedley-Whyte ET, Hyman BT (1992) Neurofibrillary tangles but not senile plaques parallel duration and severity of Alzheimer’s disease. *Neurology* **42**, 631-639.
- [17] Hooper WM, Vogel FS (1976) The limbic system in Alzheimer’s disease. *Am J Pathol* **85**, 1-19.
- [18] Augustinack JC, Helmer K, Huber KE, Kakunoori S, Zollei L, Fischl B (2010) Direct visualization of the perforant pathway in the human brain with *ex vivo* diffusion tensor imaging. *Front Hum Neurosci* **4**, 42.
- [19] Scheff SW, Price DA, Schmitt FA, Mufson EJ (2006) Hippocampal synaptic loss in early Alzheimer’s disease and mild cognitive impairment. *Neurobiol Aging* **27**, 1372-1384.
- [20] Witter MP, Wouterlood FG, Naber PA, Van Haefen T (2000) Anatomical organization of the parahippocampal-hippocampal network. *Ann N Y Acad Sci* **911**, 1-24.
- [21] Scheff SW, Price DA, Schmitt FA, DeKosky ST, Mufson EJ (2007) Synaptic alterations in CA1 in mild Alzheimer

- disease and mild cognitive impairment. *Neurology* **68**, 1501-1508.
- [22] Braak H, Braak E, Yilmazer D, Bohl J (1996) Functional anatomy of human hippocampal formation and related structures. *J Child Neurol* **11**, 265-275.
- [23] Shah A, Jhavar SS, Goel A (2012) Analysis of the anatomy of the Papez circuit and adjoining limbic system by fiber dissection techniques. *J Clin Neurosci* **19**, 289-298.
- [24] Kohler C (1985) Intrinsic projections of the retrohippocampal region in the rat brain. I. The subicular complex. *J Comp Neurol* **236**, 504-522.
- [25] Sah P, Faber ES, Lopez De Armentia M, Power J (2003) The amygdaloid complex: Anatomy and physiology. *Physiol Rev* **83**, 803-834.
- [26] McDonald AJ (1998) Cortical pathways to the mammalian amygdala. *Prog Neurobiol* **55**, 257-332.
- [27] Milner B, Squire LR, Kandel ER (1998) Cognitive neuroscience and the study of memory. *Neuron* **20**, 445-468.
- [28] Khachaturian ZS (1985) Diagnosis of Alzheimer's disease. *Arch Neurol* **42**, 1097-1105.
- [29] Mirra SS, Heyman A, McKeel D, Sumi SM, Crain BJ, Brownlee LM, Vogel FS, Hughes JP, van Belle G, Berg L (1991) The Consortium to Establish a Registry for Alzheimer's Disease (CERAD). Part II. Standardization of the neuropathologic assessment of Alzheimer's disease. *Neurology* **41**, 479-486.
- [30] Kirvell SL, Esiri M, Francis PT (2006) Down-regulation of vesicular glutamate transporters precedes cell loss and pathology in Alzheimer's disease. *J Neurochem* **98**, 939-950.
- [31] Kashani A, Lepicard E, Poirel O, Videau C, David JP, Fallet-Bianco C, Simon A, Delacourte A, Giros B, Epelbaum J, Betancur C, El Mestikawy S (2008) Loss of VGLUT1 and VGLUT2 in the prefrontal cortex is correlated with cognitive decline in Alzheimer disease. *Neurobiol Aging* **29**, 1619-1630.
- [32] Mitew S, Kirkcaldie MT, Dickson TC, Vickers JC (2013) Altered synapses and gliotransmission in Alzheimer's disease and AD model mice. *Neurobiol Aging* **34**, 2341-2351.
- [33] Cassano T, Serviddio G, Gaetani S, Romano A, Dipasquale P, Cianci S, Bellanti F, Laconca L, Romano AD, Padalino I, LaFerla FM, Nicoletti F, Cuomo V, Vendemiale G (2012) Glutamatergic alterations and mitochondrial impairment in a murine model of Alzheimer disease. *Neurobiol Aging* **33**, 1121 e1121-1112.
- [34] Canas PM, Simoes AP, Rodrigues RJ, Cunha RA (2014) Predominant loss of glutamatergic terminal markers in a beta-amyloid peptide model of Alzheimer's disease. *Neuropharmacology* **76**(Pt A), 51-56.
- [35] Jellinger KA (2012) Neuropathological subtypes of Alzheimer's disease. *Acta Neuropathol* **123**, 153-154.
- [36] Mota SI, Ferreira IL, Rego AC (2014) Dysfunctional synapse in Alzheimer's disease – A focus on NMDA receptors. *Neuropharmacology* **76**(Pt A), 16-26.
- [37] Rank KB, Pauley AM, Bhattacharya K, Wang Z, Evans DB, Fleck TJ, Johnston JA, Sharma SK (2002) Direct interaction of soluble human recombinant tau protein with Abeta 1-42 results in tau aggregation and hyperphosphorylation by tau protein kinase II. *FEBS Lett* **514**, 263-268.
- [38] Crimins JL, Pooler A, Polydoro M, Luebke JI, Spiess-Jones TL (2013) The intersection of amyloid beta and tau in glutamatergic synaptic dysfunction and collapse in Alzheimer's disease. *Ageing Res Rev* **12**, 757-763.
- [39] Puzzo D, Privitera L, Fa M, Staniszewski A, Hashimoto G, Aziz F, Sakurai M, Ribe EM, Troy CM, Mercken M, Jung SS, Palmeri A, Arancio O (2011) Endogenous amyloid-beta is necessary for hippocampal synaptic plasticity and memory. *Ann Neurol* **69**, 819-830.
- [40] Abramov E, Dolev I, Fogel H, Ciccotosto GD, Ruff E, Slutsky I (2009) Amyloid-beta as a positive endogenous regulator of release probability at hippocampal synapses. *Nat Neurosci* **12**, 1567-1576.
- [41] Sokolow S, Luu SH, Nandy K, Miller CA, Vinters HV, Poon WW, Gylys KH (2012) Preferential accumulation of amyloid-beta in presynaptic glutamatergic terminals (VGLUT1 and VGLUT2) in Alzheimer's disease cortex. *Neurobiol Dis* **45**, 381-387.
- [42] Lacor PN, Buniel MC, Furlow PW, Clemente AS, Velasco PT, Wood M, Viola KL, Klein WL (2007) Abeta oligomer-induced aberrations in synapse composition, shape, and density provide a molecular basis for loss of connectivity in Alzheimer's disease. *J Neurosci* **27**, 796-807.
- [43] Yu W, Lu B (2012) Synapses and dendritic spines as pathogenic targets in Alzheimer's disease. *Neural Plast* **2012**, 247150.
- [44] Lesne S, Ali C, Gabriel C, Croci N, MacKenzie ET, Glabe CG, Plotkine M, Marchand-Verrecchia C, Vivien D, Buisson A (2005) NMDA receptor activation inhibits alpha-secretase and promotes neuronal amyloid-beta production. *J Neurosci* **25**, 9367-9377.
- [45] Tovar KR, Westbrook GL (1999) The incorporation of NMDA receptors with a distinct subunit composition at nascent hippocampal synapses *in vitro*. *J Neurosci* **19**, 4180-4188.
- [46] Goebel-Goody SM, Davies KD, Alvestad Linger RM, Freund RK, Browning MD (2009) Phospho-regulation of synaptic and extrasynaptic N-methyl-D-aspartate receptors in adult hippocampal slices. *Neuroscience* **158**, 1446-1459.
- [47] Um JW, Nygaard HB, Heiss JK, Kostylev MA, Stagi M, Vortmeyer A, Wisniewski T, Gunther EC, Strittmatter SM (2012) Alzheimer amyloid-beta oligomer bound to postsynaptic prion protein activates Fyn to impair neurons. *Nat Neurosci* **15**, 1227-1235.
- [48] Beraldo FH, Arantes CP, Santos TG, Machado CF, Roffe M, Hajj GN, Lee KS, Magalhaes AC, Caetano FA, Mancini GL, Lopes MH, Americo TA, Magdesian MH, Ferguson SS, Linden R, Prado MA, Martins VR (2011) Metabotropic glutamate receptors transduce signals for neurite outgrowth after binding of the prion protein to laminin gamma1 chain. *FASEB J* **25**, 265-279.
- [49] Hardingham GE, Fukunaga Y, Bading H (2002) Extrasynaptic NMDARs oppose synaptic NMDARs by triggering CREB shut-off and cell death pathways. *Nat Neurosci* **5**, 405-414.
- [50] Zhou Q, Sheng M (2013) NMDA receptors in nervous system diseases. *Neuropharmacology* **74**, 69-75.
- [51] Baudry M, Lynch G (2001) Remembrance of arguments past: How well is the glutamate receptor hypothesis of LTP holding up after 20 years? *Neurobiol Learn Mem* **76**, 284-297.
- [52] Escobar ML, Derrick B (2007) Long-term potentiation and depression as putative mechanisms for memory formation. In *Neural Plasticity and Memory: From Genes to Brain Imaging*, Bermúdez-Rattoni F, ed. CRC Press, Boca Raton (FL), pp. 15-45.
- [53] Wang HW, Pasternak JF, Kuo H, Ristic H, Lambert MP, Chromy B, Viola KL, Klein WL, Stine WB, Krafft GA, Trommer BL (2002) Soluble oligomers of beta amyloid (1-42) inhibit long-term potentiation but not long-term depression in rat dentate gyrus. *Brain Res* **924**, 133-140.

- 693 [54] Kim JH, Anwyl R, Suh YH, Djamgoz MB, Rowan MJ
694 (2001) Use-dependent effects of amyloidogenic fragments
695 of (beta)-amyloid precursor protein on synaptic plasticity in
696 rat hippocampus *in vivo*. *J Neurosci* **21**, 1327-1333.
- 697 [55] Hsieh H, Boehm J, Sato C, Iwatsubo T, Tomita T, Sisodia
698 S, Malinow R (2006) AMPAR removal underlies
699 Abeta-induced synaptic depression and dendritic spine loss.
700 *Neuron* **52**, 831-843.
- 701 [56] Snyder EM, Nong Y, Almeida CG, Paul S, Moran T, Choi
702 EY, Nairn AC, Salter MW, Lombroso PJ, Gouras GK, Green-
703 gard P (2005) Regulation of NMDA receptor trafficking by
704 amyloid-beta. *Nat Neurosci* **8**, 1051-1058.
- 705 [57] D'Amelio M, Cavallucci V, Middei S, Marchetti C, Pacioni
706 S, Ferri A, Diamantini A, De Zio D, Carrara P, Battistini L,
707 Moreno S, Bacci A, Ammassari-Teule M, Marie H, Ceconi
708 F (2011) Caspase-3 triggers early synaptic dysfunction in
709 a mouse model of Alzheimer's disease. *Nat Neurosci* **14**,
710 69-76.
- 711 [58] Collingridge GL, Isaac JT, Wang YT (2004) Receptor traf-
712 ficking and synaptic plasticity. *Nat Rev Neurosci* **5**, 952-962.
- 713 [59] Kessels HW, Nabavi S, Malinow R (2013) Metabotropic
714 NMDA receptor function is required for beta-amyloid-
715 induced synaptic depression. *Proc Natl Acad Sci U S A* **110**,
716 4033-4038.
- 717 [60] Nosyreva E, Szabla K, Autry AE, Ryazanov AG, Monteggia
718 LM, Kavalali ET (2013) Acute suppression of spontaneous
719 neurotransmission drives synaptic potentiation. *J Neurosci*
720 **33**, 6990-7002.
- 721 [61] Saura CA, Choi SY, Beglopoulos V, Malkani S, Zhang D,
722 Shankaranarayana Rao BS, Chattarji S, Kelleher RJ, 3rd,
723 Kandel ER, Duff K, Kirkwood A, Shen J (2004) Loss of pre-
724 senilin function causes impairments of memory and synaptic
725 plasticity followed by age-dependent neurodegeneration.
726 *Neuron* **42**, 23-36.
- 727 [62] Corlew R, Wang Y, Ghermazien H, Erisir A, Philpot BD
728 (2007) Developmental switch in the contribution of presyn-
729 aptic and postsynaptic NMDA receptors to long-term
730 depression. *J Neurosci* **27**, 9835-9845.
- 731 [63] Aoki C, Lee J, Nedeleescu H, Ahmed T, Ho A, Shen J (2009)
732 Increased levels of NMDA receptor NR2A subunits at pre-
733 and postsynaptic sites of the hippocampal CA1: An early
734 response to conditional double knockout of presenilin 1 and
735 2. *J Comp Neurol* **517**, 512-523.
- 736 [64] Gastard MC, Troncoso JC, Koliatsos VE (2003) Cas-
737 pase activation in the limbic cortex of subjects with early
738 Alzheimer's disease. *Ann Neurol* **54**, 393-398.
- 739 [65] Parameshwaran K, Dhanasekaran M, Suppiramaniam V
740 (2008) Amyloid beta peptides and glutamatergic synaptic
741 dysregulation. *Exp Neurol* **210**, 7-13.
- 742 [66] Ferreira IL, Bajouco LM, Mota SI, Auberson YP, Oliveira
743 CR, Rego AC (2012) Amyloid beta peptide 1-42 disturbs
744 intracellular calcium homeostasis through activation of
745 GluN2B-containing N-methyl-d-aspartate receptors in
746 cortical cultures. *Cell Calcium* **51**, 95-106.
- 747 [67] Sattler R, Xiong Z, Lu WY, Hafner M, MacDonald JF,
748 Tymianski M (1999) Specific coupling of NMDA receptor
749 activation to nitric oxide neurotoxicity by PSD-95 protein.
750 *Science* **284**, 1845-1848.
- 751 [68] Liu Y, Wong TP, Aarts M, Rooyackers A, Liu L, Lai TW,
752 Wu DC, Lu J, Tymianski M, Craig AM, Wang YT (2007)
753 NMDA receptor subunits have differential roles in medi-
754 ating excitotoxic neuronal death both *in vitro* and *in vivo*. *J*
755 *Neurosci* **27**, 2846-2857.
- 756 [69] Bordji K, Becerril-Ortega J, Nicole O, Buisson A (2010)
757 Activation of extrasynaptic, but not synaptic, NMDA recep-
tors modifies amyloid precursor protein expression pattern
and increases amyloid-ss production. *J Neurosci* **30**, 15927-
15942.
- [70] Danysz W, Parsons CG (2012) Alzheimer's disease,
beta-amyloid, glutamate, NMDA receptors and memantine-
searching for the connections. *Br J Pharmacol* **167**, 324-352.
- [71] Amadoro G, Ciotti MT, Costanzi M, Cestari V, Calissano
P, Canu N (2006) NMDA receptor mediates tau-induced
neurotoxicity by calpain and ERK/MAPK activation. *Proc*
Natl Acad Sci U S A **103**, 2892-2897.
- [72] Hoover BR, Reed MN, Su J, Penrod RD, Kotilinek LA,
Grant MK, Pitstick R, Carlson GA, Lanier LM, Yuan LL,
Ashe KH, Liao D (2010) Tau mislocalization to dendritic
spines mediates synaptic dysfunction independently of neuro-
degeneration. *Neuron* **68**, 1067-1081.
- [73] Ittner LM, Ke YD, Delerue F, Bi M, Gladbach A, van Eersel
J, Wolfing H, Chieng BC, Christie MJ, Napier IA, Eckert A,
Staufenbiel M, Hardeman E, Gotz J (2010) Dendritic func-
tion of tau mediates amyloid-beta toxicity in Alzheimer's
disease mouse models. *Cell* **142**, 387-397.
- [74] Matos M, Augusto E, Oliveira CR, Agostinho P (2008)
Amyloid-beta peptide decreases glutamate uptake in cul-
tured astrocytes: Involvement of oxidative stress and
mitogen-activated protein kinase cascades. *Neuroscience*
156, 898-910.
- [75] Novelli A, Reilly JA, Lysko PG, Henneberry RC (1988) Glu-
tamate becomes neurotoxic via the N-methyl-D-aspartate
receptor when intracellular energy levels are reduced. *Brain*
Res **451**, 205-212.
- [76] Huang YJ, Lin CH, Lane HY, Tsai GE (2012) NMDA neuro-
transmission dysfunction in behavioral and psychological
symptoms of Alzheimer's disease. *Curr Neuroparmacol*
10, 272-285.
- [77] Muir KW, Lees KR (1995) Clinical experience with excita-
tory amino acid antagonist drugs. *Stroke* **26**, 503-513.
- [78] Muir KW (2006) Glutamate-based therapeutic approaches:
Clinical trials with NMDA antagonists. *Curr Opin Pharma-
col* **6**, 53-60.
- [79] Lipton SA (2004) Paradigm shift in NMDA receptor
antagonist drug development: Molecular mechanism of
uncompetitive inhibition by memantine in the treatment
of Alzheimer's disease and other neurologic disorders. *J*
Alzheimers Dis **6**, S61-S74.
- [80] Xia P, Chen HS, Zhang D, Lipton SA (2010) Memantine
preferentially blocks extrasynaptic over synaptic NMDA
receptor currents in hippocampal autapses. *J Neurosci* **30**,
11246-11250.
- [81] Grimwood S, Gilbert E, Ragan CI, Hutson PH (1996)
Modulation of 45Ca²⁺ influx into cells stably expressing
recombinant human NMDA receptors by ligands acting at
distinct recognition sites. *J Neurochem* **66**, 2589-2595.
- [82] Ray B, Banerjee PK, Greig NH, Lahiri DK (2010) Meman-
tine treatment decreases levels of secreted Alzheimer's
amyloid precursor protein (APP) and amyloid beta (A beta)
peptide in the human neuroblastoma cells. *Neurosci Lett*
470, 1-5.
- [83] Wilkinson D (2012) A review of the effects of memantine
on clinical progression in Alzheimer's disease. *Int J Geriatr*
Psychiatry **27**, 769-776.
- [84] Yang Z, Zhou X, Zhang Q (2013) Effectiveness and safety of
memantine treatment for Alzheimer's disease. *J Alzheimers*
Dis **36**, 445-458.
- [85] Parsons CG, Stoffler A, Danysz W (2007) Memantine:
A NMDA receptor antagonist that improves memory by
restoration of homeostasis in the glutamatergic system—too

- little activation is bad, too much is even worse. *Neuropharmacology* **53**, 699-723.
- [86] Zadori D, Klivenyi P, Plangar I, Toldi J, Vecsei L (2011) Endogenous neuroprotection in chronic neurodegenerative disorders: With particular regard to the kynurenines. *J Cell Mol Med* **15**, 701-717.
- [87] Zadori D, Klivenyi P, Szalardy L, Fulop F, Toldi J, Vecsei L (2012) Mitochondrial disturbances, excitotoxicity, neuroinflammation and kynurenines: Novel therapeutic strategies for neurodegenerative disorders. *J Neurol Sci* **322**, 187-191.
- [88] Schwarcz R, Bruno JP, Muchowski PJ, Wu HQ (2012) Kynurenines in the mammalian brain: When physiology meets pathology. *Nat Rev Neurosci* **13**, 465-477.
- [89] Vecsei L, Szalardy L, Fulop F, Toldi J (2013) Kynurenines in the CNS: Recent advances and new questions. *Nat Rev Drug Discov* **12**, 64-82.
- [90] Plangar I, Zadori D, Klivenyi P, Toldi J, Vecsei L (2011) Targeting the kynurenine pathway-related alterations in Alzheimer's disease: A future therapeutic strategy. *J Alzheimers Dis* **24**(Suppl 2), 199-209.
- [91] Perkins MN, Stone TW (1982) An iontophoretic investigation of the actions of convulsant kynurenines and their interaction with the endogenous excitant quinolinic acid. *Brain Res* **247**, 184-187.
- [92] Kessler M, Terramani T, Lynch G, Baudry M (1989) A glycine site associated with N-methyl-D-aspartic acid receptors: Characterization and identification of a new class of antagonists. *J Neurochem* **52**, 1319-1328.
- [93] Stone TW (1993) Neuropharmacology of quinolinic and kynurenic acids. *Pharmacol Rev* **45**, 309-379.
- [94] Swartz KJ, During MJ, Freese A, Beal MF (1990) Cerebral synthesis and release of kynurenic acid: An endogenous antagonist of excitatory amino acid receptors. *J Neurosci* **10**, 2965-2973.
- [95] Linderholm K, Powell S, Olsson E, Holtze M, Snodgrass R, Erhardt S (2010) Role of the NMDA-receptor in prepulse inhibition in the rat. *Int J Tryptophan Res* **3**, 1-12.
- [96] Vecsei L, Beal MF (1991) Comparative behavioral and pharmacological studies with centrally administered kynurenine and kynurenic acid in rats. *Eur J Pharmacol* **196**, 239-246.
- [97] Baran H, Jellinger K, Deecke L (1999) Kynurenine metabolism in Alzheimer's disease. *J Neural Transm* **106**, 165-181.
- [98] Potter MC, Elmer GI, Bergeron R, Albuquerque EX, Guidetti P, Wu HQ, Schwarcz R (2010) Reduction of endogenous kynurenic acid formation enhances extracellular glutamate, hippocampal plasticity, and cognitive behavior. *Neuropsychopharmacology* **35**, 1734-1742.
- [99] Hilmas C, Pereira EF, Alkondon M, Rassoulpour A, Schwarcz R, Albuquerque EX (2001) The brain metabolite kynurenic acid inhibits alpha7 nicotinic receptor activity and increases non-alpha7 nicotinic receptor expression: Physiopathological implications. *J Neurosci* **21**, 7463-7473.
- [100] Marchi M, Risso F, Viola C, Cavazzani P, Raiteri M (2002) Direct evidence that release-stimulating alpha7* nicotinic cholinergic receptors are localized on human and rat brain glutamatergic axon terminals. *J Neurochem* **80**, 1071-1078.
- [101] Pocivavsek A, Wu HQ, Potter MC, Elmer GI, Pellicciari R, Schwarcz R (2011) Fluctuations in endogenous kynurenic acid control hippocampal glutamate and memory. *Neuropsychopharmacology* **36**, 2357-2367.
- [102] Prescott C, Weeks AM, Staley KJ, Partin KM (2006) Kynurenic acid has a dual action on AMPA receptor responses. *Neurosci Lett* **402**, 108-112.
- [103] Rozsa E, Robotka H, Vecsei L, Toldi J (2008) The Janus-face kynurenic acid. *J Neural Transm* **115**, 1087-1091.
- [104] Mony L, Kew JN, Gunthorpe MJ, Paoletti P (2009) Allosteric modulators of NR2B-containing NMDA receptors: Molecular mechanisms and therapeutic potential. *Br J Pharmacol* **157**, 1301-1317.
- [105] Williams K (1993) Ifenprodil discriminates subtypes of the N-methyl-D-aspartate receptor: Selectivity and mechanisms at recombinant heteromeric receptors. *Mol Pharmacol* **44**, 851-859.
- [106] Kew JN, Kemp JA (1998) An allosteric interaction between the NMDA receptor polyamine and ifenprodil sites in rat cultured cortical neurones. *J Physiol* **512**(Pt 1), 17-28.
- [107] Han X, Tomitori H, Mizuno S, Higashi K, Full C, Fukiwake T, Terui Y, Leewanih P, Nishimura K, Toida T, Williams K, Kashiwagi K, Igarashi K (2008) Binding of spermine and ifenprodil to a purified, soluble regulatory domain of the N-methyl-D-aspartate receptor. *J Neurochem* **107**, 1566-1577.
- [108] Costa RO, Lacor PN, Ferreira IL, Resende R, Auberson YP, Klein WL, Oliveira CR, Rego AC, Pereira CM (2012) Endoplasmic reticulum stress occurs downstream of GluN2B subunit of N-methyl-d-aspartate receptor in mature hippocampal cultures treated with amyloid-beta oligomers. *Aging Cell* **11**, 823-833.
- [109] Hu NW, Klyubin I, Anwyl R, Rowan MJ (2009) GluN2B subunit-containing NMDA receptor antagonists prevent Abeta-mediated synaptic plasticity disruption *in vivo*. *Proc Natl Acad Sci U S A* **106**, 20504-20509.
- [110] Ronicke R, Mikhaylova M, Ronicke S, Meinhardt J, Schroder UH, Fandrich M, Reiser G, Kreutz MR, Reymann KG (2011) Early neuronal dysfunction by amyloid beta oligomers depends on activation of NR2B-containing NMDA receptors. *Neurobiol Aging* **32**, 2219-2228.
- [111] Li S, Jin M, Koeglsperger T, Shepardson NE, Shankar GM, Selkoe DJ (2011) Soluble Abeta oligomers inhibit long-term potentiation through a mechanism involving excessive activation of extrasynaptic NR2B-containing NMDA receptors. *J Neurosci* **31**, 6627-6638.
- [112] Rammes G, Hasenjager A, Sroka-Saidi K, Deussing JM, Parsons CG (2011) Therapeutic significance of NR2B-containing NMDA receptors and mGluR5 metabotropic glutamate receptors in mediating the synaptotoxic effects of beta-amyloid oligomers on long-term potentiation (LTP) in murine hippocampal slices. *Neuropharmacology* **60**, 982-990.
- [113] Wong TP, Howland JG, Robillard JM, Ge Y, Yu W, Titterness AK, Brebner K, Liu L, Weinberg J, Christie BR, Phillips AG, Wang YT (2007) Hippocampal long-term depression mediates acute stress-induced spatial memory retrieval impairment. *Proc Natl Acad Sci U S A* **104**, 11471-11476.
- [114] Fulop F, Szatmari I, Vamos E, Zadori D, Toldi J, Vecsei L (2009) Syntheses, transformations and pharmaceutical applications of kynurenic acid derivatives. *Curr Med Chem* **16**, 4828-4842.
- [115] Nagy K, Plangar I, Tuka B, Gellert L, Varga D, Demeter I, Farkas T, Kis Z, Marosi M, Zadori D, Klivenyi P, Fulop F, Szatmari I, Vecsei L, Toldi J (2011) Synthesis and biological effects of some kynurenic acid analogs. *Bioorg Med Chem* **19**, 7590-7596.
- [116] Borza I, Kolok S, Galgoczy K, Gere A, Horvath C, Farkas S, Greiner I, Domany G (2007) Kynurenic acid amides as novel NR2B selective NMDA receptor antagonists. *Bioorg Med Chem Lett* **17**, 406-409.

- 953 [117] Gellert L, Fuzik J, Goblos A, Sarkozi K, Marosi M, Kis
954 Z, Farkas T, Szatmari I, Fulop F, Vecsei L, Toldi J (2011)
955 Neuroprotection with a new kynurenic acid analog in the
956 four-vessel occlusion model of ischemia. *Eur J Pharmacol*
957 **667**, 182-187.
- 958 [118] Zadori D, Nyiri G, Szonyi A, Szatmari I, Fulop F, Toldi
959 J, Freund TF, Vecsei L, Klivenyi P (2011) Neuroprotective
960 effects of a novel kynurenic acid analogue in a transgenic
961 mouse model of Huntington's disease. *J Neural Transm* **118**,
962 865-875.
- 963 [119] Caraci F, Battaglia G, Sortino MA, Spampinato S, Molinaro
964 G, Copani A, Nicoletti F, Bruno V (2012) Metabotropic
965 glutamate receptors in neurodegeneration/neuroprotection:
966 Still a hot topic? *Neurochem Int* **61**, 559-565.
- [120] Takahashi RN, Pamplona FA, Prediger RD (2008) Adeno-
sine receptor antagonists for cognitive dysfunction: A
review of animal studies. *Front Biosci* **13**, 2614-2632.
- [121] Rahman A (2009) The role of adenosine in Alzheimer's
disease. *Curr Neuropharmacol* **7**, 207-216.
- [122] Selkoe DJ (2002) Alzheimer's disease is a synaptic failure.
Science **298**, 789-791.
- [123] Coleman P, Federoff H, Kurlan R (2004) A focus on the
synapse for neuroprotection in Alzheimer disease and other
dementias. *Neurology* **63**, 1155-1162.

Uncorrected Author Proof