Elevated levels of PPAR-gamma in the cerebrospinal fluid of patients with multiple sclerosis

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Abstract

Peroxisome proliferator-activated receptor gamma (PPAR\(\gamma\)), a ligand-activated transcriptional factor involved in the regulation of glucose and lipid metabolism, has gained interest as a potential therapeutic target in multiple sclerosis (MS) due to its potent immunoregulatory properties and the therapeutic efficacy of its ligands in experimental autoimmune encephalitis (EAE). Elevated expression of PPAR\(\gamma\) has been observed in the spinal cord of EAE mice and in an in vitro model of antigen-induced demyelination; however, no reports have yet been available on the PPAR\(\gamma\) status in the central nervous system of human individuals with MS. Aiming to identify a possible alteration, the present study assessed the levels of PPAR\(\gamma\) protein in the cerebrospinal fluid (CSF) of MS patients via ELISA technique. We report a pronounced elevation in the CSF levels of PPAR\(\gamma\) in MS patients (\(n=35\)) compared to non-inflammatory controls (\(n=22\)). This elevation was independent of blood-CSF barrier integrity, but correlated with CSF white blood cell count and IgG index, associating the observed elevation with neuroinflammation. Controlling for potential confounders, the CSF levels of PPAR\(\gamma\) further displayed a moderate but significant association with clinical severity. Corroborating with prior experimental findings, these results may contribute to our understanding about the role of PPAR\(\gamma\) in MS, and may implicate this protein as a potential CSF biomarker of the disease.

Key words: peroxisome proliferator-activated receptor gamma; multiple sclerosis; cerebrospinal fluid; ELISA; biomarker.
1. Introduction

Peroxisome proliferator-activated receptors (PPARs) are members of a nuclear hormone receptor superfamily of ligand-activated transcriptional factors that also includes thyroid, steroid and retinoid receptors. Out of the three subtypes known (PPARα, PPARβ/δ and PPARγ), PPARγ has been the most extensively investigated. PPARγ is known to be critically involved in the regulation of glucose and lipid metabolism, but more recent results also revealed an important role of PPARγ in the regulation immune response [6], gathering an increasing research interest in the field of disorders involving autoimmune processes in the pathomechanism, such as multiple sclerosis (MS) [11, 27].

MS is a chronic debilitating inflammatory central nervous system (CNS) disorder, characterized by demyelination, axonal and neuronal degeneration. The disease is supposed to be of autoimmune origin, in which encephalitogenic T-helper 1 (Th1) and interleukin 17 (IL-17)-producing Th17 lymphocytes are proposed to be the main mediators [1]. In the past years, natural and synthetic agonists of PPARγ were shown to effectively ameliorate the course of experimental autoimmune encephalitis (EAE), a widely used experimental model of MS, by inhibiting the expansion of encephalitogenic T cells [8, 9, 13, 21, 23, 29, 35]. More recently, the activation of PPARγ in both human and murine CD4+ T cells have been shown to potently inhibit the differentiation towards the Th17 lineage [16], and the eicozapentaenoic acid-induced amelioration of EAE has been reported to be accompanied by increased PPARγ in association with decreased IL-17 expression in CNS-infiltrating CD4+ T cells [35]. Conversely, pharmacological inhibition of [28, 29] or a heterozygous deficiency in PPARγ [22, 28, 29] leads to an exacerbated course of EAE in mice in association with an augmented T cell response, suggesting altogether that the activation of the PPARγ pathway may be of
therapeutic relevance in MS. Furthermore, elevated expression of PPARγ has been reported within the spinal cord of EAE mice [8, 9] and in an \textit{in vitro} model of antigen-induced demyelination [12], supporting the notion that PPARγ may have a physiological role in the regulation of CNS inflammation in rodents. However, no reports have yet been available on the PPARγ status in the CNS of human individuals with MS. This study aimed to identify a possible alteration of PPARγ expression in MS, by assessing the levels of PPARγ protein in the CSF of MS patients.

2. Patients, materials and methods

The study included CSF samples of 35 MS patients meeting the 2009 McDonald criteria for definite relapsing-remitting (RR) MS. Non-inflammatory control samples were collected from 22 age- and sex-matched patients whose differential diagnostic process necessitated a lumbar puncture (LP) but the results of which along with that of the imaging modalities revealed no abnormalities in the CNS. Written informed consent was obtained from all subjects and the study was approved by the local Ethical Committee at the University of Szeged. The demographic, clinical and laboratory parameters of the cohorts are summarized in Table 1. LPs were performed between 2010 and 2013. The patients were free of immunomodulatory therapy, and for patients experiencing a relapse (n = 27) LP was performed before the initiation of corticosteroid therapy.

Following the LP, CSF samples were centrifuged in 8,000 rpm for 10 minutes. The supernatants were stored in sterile polypropylene tubes in \textdegree{80}C until use. Commercially available sandwich enzyme-linked immunosorbent assay (ELISA) kits were applied for the quantitative detection of PPARγ, following the manufacturers’ instruction (Uscn Life Science Inc., Wuhan, China). Samples and standards were run in duplicates. The optical density
values were detected at 450/560 nm with a plate reader (Awareness Technology Inc, Palm City, FL, USA) and the respective concentrations were read from the standard curves fitted by Sigmaplot 10.0 (Systat Software Inc., Richmond, CA, USA). The intraassay and interassay coefficients of variation were 9% and 15%, respectively. The analytical sensitivity of the assay is 51 pg/ml; values below the detection limit (12/22 in the Control vs 4/35 in the MS group) were interpolated from the standard curve to calculate median and provide measures for statistical computations. The CSF total protein concentration, the white blood cell count, the albumin quotient (CSF albumin / serum albumin) and the IgG index ((CSF IgG / serum IgG) / albumin quotient) were determined in our diagnostic laboratory and were available in the clinical records. The Expanded Disability Status Scale (EDSS) score was evaluated for each patient. To provide standardised clinical measures, EDSS scores were comprehensively revised by a trained MS investigator (K.B.) who was blind to the experimental findings. The statistical analysis was performed by SPSS Statistics 17.0 (SPSS Inc., Chicago, IL, USA). The normality of data was assessed by Shapiro-Wilk test. Due to the non-Gaussian distribution of the obtained data in the MS cohort, non-parametric tests (Mann-Whitney U, Spearman’s correlation) were used. Data in the text and in Table 1 are presented as median [interquartile range]. A \( p < 0.05 \) value was regarded as significant; \( p \) values of correlation analyses were corrected for multiple testing by the Bonferroni-Dunn method. Controlling for potential confounders, a binary logistic regression model and an ordinal regression model was established to assess the association of PPAR\(\gamma\) with relapse and EDSS, respectively.

3. Results

In our study cohort, CSF levels of PPAR\(\gamma\) were highly and significantly elevated in the CSF of patients with MS compared to Control (167.5 pg/ml [75.6–372.1 pg/ml] vs 40.4 pg/ml [16.5–73.1 pg/ml], \( p < 0.0005 \), Fig. 1). The elevation of PPAR\(\gamma\) appeared to be independent of
blood-CSF barrier integrity as suggested by the lack of correlations with CSF total protein levels (Spearman’s Rho = 0.110) and albumin quotient (Spearman’s Rho = −0.076) within the MS group. However, the CSF levels of PPARγ significantly and strongly correlated with the CSF cell count ($p < 0.0005$, Spearman’s Rho = 0.722, Fig. 2) and moderately correlated with the IgG index ($p = 0.007$, Spearman’s Rho = 0.528, Figure 2). Though the highest levels of PPARγ were measured in MS patients experiencing a relapse at sampling and the median level of PPARγ was considerably higher among patients in relapse compared to those in remission (197.39 pg/ml [87.58–397.50 pg/ml] vs 112.98 pg/ml [67.76–245.07 pg/ml]), CSF PPARγ level was not a significant predictor of relapse in a binary logistic regression model controlling for CSF cell count, total protein level, IgG index and albumin quotient as potential confounding covariates ($p = 0.596$, Exp(B) = 1.001, 95% confidence interval = 0.997–1.005, Wald Chi$^2$ = 0.281, df = 1). Controlling for the same covariates in an ordinal regression model, CSF level of PPARγ displayed a moderate but significant ($p = 0.011$) positive association with EDSS with an estimate of 0.004 (95% confidence interval = 0.001–0.007; df = 1), which calls for an odds ratio of 1.49 to have an increased EDSS by every 100 pg/ml elevation in the CSF concentration of PPARγ.

4. Discussion

Our results demonstrate for the first time a pronounced elevation of PPARγ levels in the CSF of patients with MS, in association with increased CNS inflammatory parameters. These results are in line with the prior reports on elevated expression of PPARγ in the CNS of EAE mice [8, 9] and may reflect a possible physiological compensatory role of PPARγ in the regulation of CNS autoimmunity in humans. This experimentally demonstrated regulatory function is supposed to be primarily attributable to the attenuation of inflammatory cytokine expression within microglia/macrophages and astrocytes [30, 33, 37] and the (partly
concomitant) suppression of the expansion of encephalitogenic Th1 [21], Th17 cells [16] and B lymphocytes [24], accompanied by a switch towards a Th2 response [10, 11]. In addition, activation of PPAR\(\gamma\) expression in brain endothelial cells has been shown to be a negative regulator of lymphocytic transmigration [17]. Besides, PPAR\(\gamma\) activation have also been demonstrated to have protective effects on neurons and oligodendrocytes [2, 10-12, 20, 26, 34, 36]. Furthermore, the activation of PPAR\(\gamma\) has been associated with the promotion of oligodendrocyte differentiation and myelin production [3, 7, 15, 25, 31, 34]. These properties may all contribute to the positive effects of PPAR\(\gamma\) activation in experimental CNS inflammation, and may be exploited in MS therapy. Based on the experimental findings, two small clinical studies have been performed with the synthetic agonist pioglitazone, the results of both supporting that the use of PPAR\(\gamma\) agonists may be beneficial in MS [14, 32]. Moreover, in a population-based study, the Pro12Ala polymorphism in the PPARG gene strongly associated with a delayed clinical onset of MS, which draws a further link between MS and PPAR\(\gamma\) function [19].

The source of the elevated levels of PPAR\(\gamma\) observed in this study, however, is not entirely clear, and its determination will require further investigations. It is known that PPAR\(\gamma\) is expressed in neurons, oligodendrocytes, astrocytes, microglia/macrophages [3], T and B lymphocytes, dendritic cells [38] and brain endothelial cells [17]. In a murine EAE model of MS an increased expression of PPAR\(\gamma\) has been reported in spinal cord homogenates [9] and in microglia/macrophages and astrocytes within the spinal cord lesions [8]. Furthermore, a strong upregulation of PPAR\(\gamma\) has been observed in three-dimensional rat brain cultures following demyelination induced by antibodies directed against myelin oligodendrocyte glycoprotein [12]. Considering the neuropathological alterations in MS, the above listed experimental findings and our observation that PPAR\(\gamma\) levels correlated with CSF inflammatory parameters, it is possible that PPAR\(\gamma\) protein leaking out from
injured/degenerating oligodendrocytes, neurons and astrocytes contribute to the observed elevation of its levels in the CSF. Considering that PPARγ upregulation has been demonstrated in activated macrophages [30] and activated T lymphocytes [4, 5], and that PPARγ levels correlated with the CSF cell count in our study, the contribution of such cellular origin to the observed phenomenon may not be fully excluded either; however, the fact that our CSF samples were centrifuged almost immediately after LP suggest that the extent of a putative cell disintegration within the obtained sample should most probably be insufficient to cause such a significant alteration in the concentration of PPARγ in the supernatant. Though future investigations are necessary to conclude on the exact source of the observed elevation, the possible sources discussed – in light with the findings of prior experimental studies – suggest that the level of PPARγ in the CSF may reflect the severity of CNS involvement, which could account for the observed positive association with EDSS in our study.

The literature regarding alterations in PPARγ status in MS is rather sparse. Somewhat contradicting to our findings, another small clinical study examining peripheral white blood cells of MS patients reported decreased expression of PPARγ compared to control [18]. The findings of the two studies, however, are hard to compare, given that the studies focus on different compartments; furthermore, the patients enrolled in that study were either under immunomodulatory therapy or already in a secondary progressive phase, most probably representing different conditions compared to our untreated RR patients.

5. Conclusions

CSF levels of PPARγ were pronouncedly elevated in MS patients. This elevation was independent of blood-CSF barrier integrity, strongly correlated with white blood cell count, moderately correlated with IgG index, and displayed a moderate association with clinical
severity. These results may contribute to our understanding about the role of PPARγ in the pathogenesis MS, and implicate this protein as a potential CSF biomarker of the disease. Further research is necessary to determine the exact background of the observed alteration, and to assess the potential clinical relevance of this novel observation.

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7. Conflicts of interest

The authors report no conflicts of interest.
8. References


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Fig. 1. PPARγ concentrations are elevated ($p < 0.0005$) in the CSF of patients with MS (n = 35) compared to control (n = 22). The box-plots present median and quartile values. Abbrevations: CSF, cerebrospinal fluid; MS, multiple sclerosis; PPARγ, peroxisome proliferator-activated receptor gamma.
Fig. 2. PPARγ levels correlate with white blood cell count and IgG index in the CSF of patients with MS. Abbreviations: CSF, cerebrospinal fluid; MS, multiple sclerosis; PPARγ, peroxisome proliferator-activated receptor gamma.
Table 1. Demographic, laboratory and clinical parameters of the subject cohorts. The measured data are presented as median [interquartile range]. Abbrevations: EDSS, expanded disability status scale; MS, multiple sclerosis.