PANCREATIC EPITHELIAL FLUID AND BICARBONATE SECRETION IS SIGNIFICANTLY ELEVATED IN THE ABSENCE OF PHERIPHERAL SEROTONIN

József Maléth ¹^{\$}, Tamara Madácsy^{1,\$}, Petra Pallagi ¹, Anita Balázs¹, Viktória Venglovecz², Zoltán Rakonczay Jr.¹, Péter Hegyi^{1,3}

¹First Department of Medicine, ²Department of Pharmacology and Pharmacotherapy, University of Szeged, Szeged, Hungary; ³MTA-SZTE Momentum Translational Gastroenterology Research Group, University of Szeged, Szeged, Hungary; [§] These authors contributed equally to the work.

Corresponding author:

Péter Hegyi, M.D., Ph.D., D.Sc.
University of Szeged
Faculty of Medicine
First Department of Medicine
P.O. Box 427, H-6701, Szeged, Hungary
Phone: (36)(62)545-200
Fax: (36)(62)545-185
Email: hegyi.peter@med.u-szeged.hu

Dear Editor,

We read the manuscript by Sonda et al.¹ recently published in Gut with great interest. The authors elegantly demonstrated that lack of peripheral serotonin (5-HT) in tryptophan hydroxylase 1 knockout (TPH1^{-/-}) mice remarkably limited pancreatic damage and leukocyte infiltration during the early phase of cerulein-induced acute pancreatitis (AP) and identified 5-HT as an important regulator of zymogen secretion in acinar cells. Although the study was very comprehensive, the ductal function of TPH1^{-/-} mice was not investigated, which might be another key player in this protection. Notably, 5-HT was shown to inhibit fluid and HCO₃⁻ secretion of pancreatic ductal epithelial cells (PDEC)² that play pivotal role in pancreatic physiology and can influence the severity of AP³. Thus we investigated the possible alterations of pancreatic ductal secretion in TPH1^{-/-} mice.

To achieve our aims intra/interlobular pancreatic ducts were isolated from the pancreas of wild type (TPH1^{+/+}) and TPH1^{-/-} mice. HCO₃⁻ secretion was measured by three different, but complementary methods using microfluorometry⁴, whereas fluid secretion was measured by the swelling technique using videomicroscopy³. *In vivo* basal pancreatic fluid secretion was determined in anesthetized mice³.

The alkali-load technique (basolateral administration of 20mM NH₄Cl) revealed that the apical bicarbonate secretion was markedly increased in TPH1^{-/-} mice compared to TPH1^{+/+} (Fig.1.A,D). Notably, *in vitro* administration of 0.1 μ M 5-HT impaired the recovery in both cases (not shown). The inhibitory-stop method (basolateral administration of 0.2mM dihydro-4,4'-diisothiocyanostilbene-2,2'-disulfonic acid (H₂DIDS) and 0.2mM amiloride) confirmed the marked increase of bicarbonate efflux in TPH1^{-/-} mice (Fig.1.B,E).

After that we provided direct evidence that the Cl^{-}/HCO_{3}^{-} exchanger is crucial in the elevated ductal secretion, since the rate of pH_i recovery was significantly elevated in TPH1^{-/-} mice after Cl⁻ withdrawal from the lumen of microperfused pancreatic ducts (Fig.1.C,F).

These three independent, but complementary methods clearly demonstrated an increased pancreatic bicarbonate secretion in TPH1^{-/-} mice.

In the next step we compared the *in vitro* and *in vivo* pancreatic fluid secretion in TPH1^{+/+} and TPH1^{-/-} mice. Using sealed pancreatic ducts we showed that administration of HCO_3^- resulted in an immediate increase of the relative luminal volume, which was further increased by the administration of 5µM forskolin and 100µM 3-isobutyl-1-methylxanthine (IBMX). These experiments showed that both bicarbonate and secretagogue-induced *in vitro* fluid secretion is significantly elevated in TPH1^{-/-} mice (Fig.2.A). *In vivo* pancreatic ductal fluid secretion was

analyzed in anesthetized mice, which was 0.33±0.05ml/hour/bwkg in TPH1^{-/-} mice, almost twice as much as the TPH1^{+/+} (0.18±0.017ml/hour/bwkg) (Fig.2.B).

These findings indicate that the fluid and HCO₃⁻ secretion is markedly increased in the absence of peripheral 5-HT, which may also contribute to the decreased severity of AP in TPH1^{-/-} mice, besides the altered cytoskeleton dynamics of pancreatic acinar cells.

Figure 1. Pancreatic ductal bicarbonate secretion is markedly increased in TPH1^{-/-} mice. (A-C) pH_i traces of PDEC in standard $HCO_3^{-/}CO_2$ solution. (A) Recovery from alkalosis during NH₄Cl administration, (B) the rate of acidosis caused by H₂DIDS and amiloride, (C) the rate of recovery after luminal Cl⁻ removal. Bar charts show summary data for the (D) base fluxes [-J(B⁻/min)] (calculated from the dpH/dt as described earlier⁴) afterNH₄Cl, and (E) transporter inhibitor administration, or (F) luminal Cl⁻ removal. Data are shown as means \pm SEM from 35-50 ROIs in 8-10 ducts. a: p<0.01 vs. TPH1^{+/+}.

Figure 2. Pancreatic fluid secretion is significantly increased in TPH1^{-/-} mice. (A) Changes in the relative luminal volume of pancreatic ducts from TPH1^{+/+} (red line, n = 8 from three animals) and TPH1^{-/-} (blue line, n = 8 from three animals) mice. (B) The volume of pancreatic juice collected *in vivo* from anesthetized TPH1^{+/+} and TPH1^{-/-} mice. Data are shown as means ± SEM, n:5/group; a: p<0.01 vs. TPH1^{+/+}.

Acknowledgements. The authors are grateful to Prof. Dr. Rolf Graf (Department of Visceral and Transplantation Surgery, University Hospital, Zurich, Switzerlandand) and to Prof. Dr. Michael Bader (Max-Delbrück-Center for Molecular Medicine, Berlin, Germany) for providing us the TPH1^{-/-} mice. Our work was supported by the MTA-SZTE Momentum Grant (LP2014-10/2014) and the National Scientific Research Found grant K109756.

References

- 1. Sonda S, Silva AB, Grabliauskaite K, Saponara E, Weber A, Jang JH, et al. Serotonin regulates amylase secretion and acinar cell damage during murine pancreatitis. *Gut* 2013;62(6):890-8.
- 2. Suzuki A, Naruse S, Kitagawa M, Ishiguro H, Yoshikawa T, Ko SB, et al. 5hydroxytryptamine strongly inhibits fluid secretion in guinea pig pancreatic duct cells. *The Journal of clinical investigation* 2001;108(5):749-56.
- 3. Pallagi P, Balla Z, Singh AK, Dosa S, Ivanyi B, Kukor Z, et al. The role of pancreatic ductal secretion in protection against acute pancreatitis in mice*. *Critical care medicine* 2014;42(3):e177-88.
- 4. Maleth J, Balazs A, Pallagi P, Balla Z, Kui B, Katona M, et al. Alcohol disrupts levels and function of the cystic fibrosis transmembrane conductance regulator to promote development of pancreatitis. *Gastroenterology* 2015;148(2):427-39 e16.