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To cite this article: Adriano Mollica, Sveva Pelliccia, Valeria Famiglini, Azzurra Stefanucci, Giorgia Macedonio, Annalisa Chiavaroli, Giustino Orlando, Luigi Brunetti, Claudio Ferrante, Stefano Pieretti, Ettore Novellino, Sandor Benyhe, Ferenc Zador, Anna Erdei, Edina Szucs, Reza Samavati, Szalbolch Dvrorasko, Csaba Tomboly, Rino Ragno, Alexandros Patsilnakos & Romano Silvestri (2017) Exploring the first Rimonabant analog-opioid peptide hybrid compound, as bivalent ligand for CB1 and opioid receptors, *Journal of Enzyme Inhibition and Medicinal Chemistry*, 32:1, 444-451, DOI: [10.1080/14756366.2016.1260565](https://doi.org/10.1080/14756366.2016.1260565)

To link to this article: <http://dx.doi.org/10.1080/14756366.2016.1260565>



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Exploring the first Rimonabant analog-opioid peptide hybrid compound, as bivalent ligand for CB1 and opioid receptors

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ABSTRACT

Cannabinoid (CB) and opioid systems are both involved in analgesia, food intake, mood and behavior. Due to the co-localization of μ -opioid (MOR) and CB1 receptors in various regions of the central nervous system (CNS) and their ability to form heterodimers, bivalent ligands targeting to both these systems may be good candidates to investigate the existence of possible cross-talking or synergistic effects, also at sub-effective doses. In this work, we selected from a small series of new Rimonabant analogs one CB1R reverse agonist to be conjugated to the opioid fragment Tyr-D-Ala-Gly-Phe-NH₂. The bivalent compound (**9**) has been used for *in vitro* binding assays, for *in vivo* antinociception models and *in vitro* hypothalamic perfusion test, to evaluate the neurotransmitters release.

ARTICLE HISTORY

Received 13 August 2016
Revised 31 October 2016
Accepted 8 November 2016

KEYWORDS

Cannabinoid receptor CB1R;
Rimonabant; opioids;
bivalent ligand; pain

Introduction

Cannabinoid and opioid receptors are expressed mostly in the same CNS areas, and both are involved in the control of analgesia, food intake, mood and behavior. In the dorsal horn of the spinal cord, the μ -opioid receptor MOR and CB1R are co-localized at the same neurons, also at the supra-spinal level, such as the periaqueductal gray (PAG), the raphe nuclei and the central-medial thalamic nuclei¹.

According to the several cell line studies where the MOR and CB1R are endogenously co-expressed, they share cAMP signaling pathways, even though they may use different sets of G-proteins².

Studies on opioid and CB1R knock-out mice, demonstrated that their density and activity strongly depend on each other^{3,4}. Also, preclinical and clinical studies stated that the interaction between the opioid and cannabinoid systems can lead to promising therapeutic applications in pain control and in alimentary disorders management^{5–8}. Thus, bivalent ligands binding to CB1R and MOR simultaneously may result in new potent analgesic agents. Recently, Le Naour et al.⁶ proposed a bivalent approach to target both MOR and CB1R, connecting a selective MOR agonist to a CB1R selective inverse agonist, *via* a spacer group of varied length.

One of the synthesized compounds showed an extremely potent activity in *in vivo* antinociceptive tests and was devoid of tolerance. Additive or synergic interactions between opioid and cannabinoid systems in producing analgesia have been previously described and reviewed in detail^{9,10}. Morphine-induced

antinociception is completely reversed by the CB1R antagonist AM251¹¹, and tetrahydrocannabinol (THC)-induced antinociception is blocked by naloxone¹².


The cross-tolerance between THC and morphine and the possibility that these receptors interact pharmacologically were demonstrated by naloxone or CB1R antagonist. Synergism between cannabinoids and opioids at sub-effective doses has also been reported^{13,14}. Additionally, co-administration of morphine with a CB1R antagonist inhibited the development of both acute and chronic tolerance to morphine¹⁵.

Other evidence for interaction was obtained from self-administration studies showing that both receptors are involved in reward processes. In this regard, both CB1R antagonist (SR 141716)¹⁶, and opioid antagonist naloxone decreased self-administration of morphine or Δ^9 -THC¹⁷. In mice lacking μ (MOR) and δ (DOR) opioid receptors, the cannabinoid withdrawal syndrome is reduced¹⁸.

According to our research line based on the design of multitarget compounds^{19,20}, the present study represents a starting point to develop bivalent ligands as pharmacologic tools to investigate the MOR–CB1R mutual interactions. The bivalent ligand designed for this purpose consists of a selective μ -receptor peptide agonist connected to a CB1R selective inverse agonist fused together²¹.

The novel compound was extensively tested for *in vitro* binding, GTP stimulation, neurotransmitters release and antinociceptive *in vivo* activity. The design rationale for targeting MOR and CB1R simultaneously is based on previous studies with bivalent ligands⁶. The MOR agonist pharmacophore derived from the opioid peptide

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 Supplemental data for this article can be accessed [here](#).

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biphalin, was previously employed in the design of several bivalent compounds, due to its property to well tolerate the connection at the C-terminus with another pharmacophore^{22,23,19}.

Since other authors show that a CB1R inverse agonist is capable of eliminating morphine tolerance and dependence^{24,25}, we selected compound **5** (Scheme 1) with CB1R inverse agonist activity as CB1R pharmacophore (Figure 1), in analogy with the work by Le Naour et al.⁶

Thus, the bivalent ligand **9** was synthesized following the "fused-bivalent approach"²¹, with the expectation that the resulting product would be capable to interact at CB1R, MOR and CB1R-MOR heterodimer form. The bivalent compound was prepared by coupling each Boc-protected amino acid of the opioid peptide sequence to the secondary amine of 4-aminopiperidine group present in the structure of Rimonabant analog **5**.

The opioid fragment Tyr-D-Ala-Gly-Phe-NH₂ (**10**) was used as reference compound for opioid activity on MOR and DOR¹⁹.

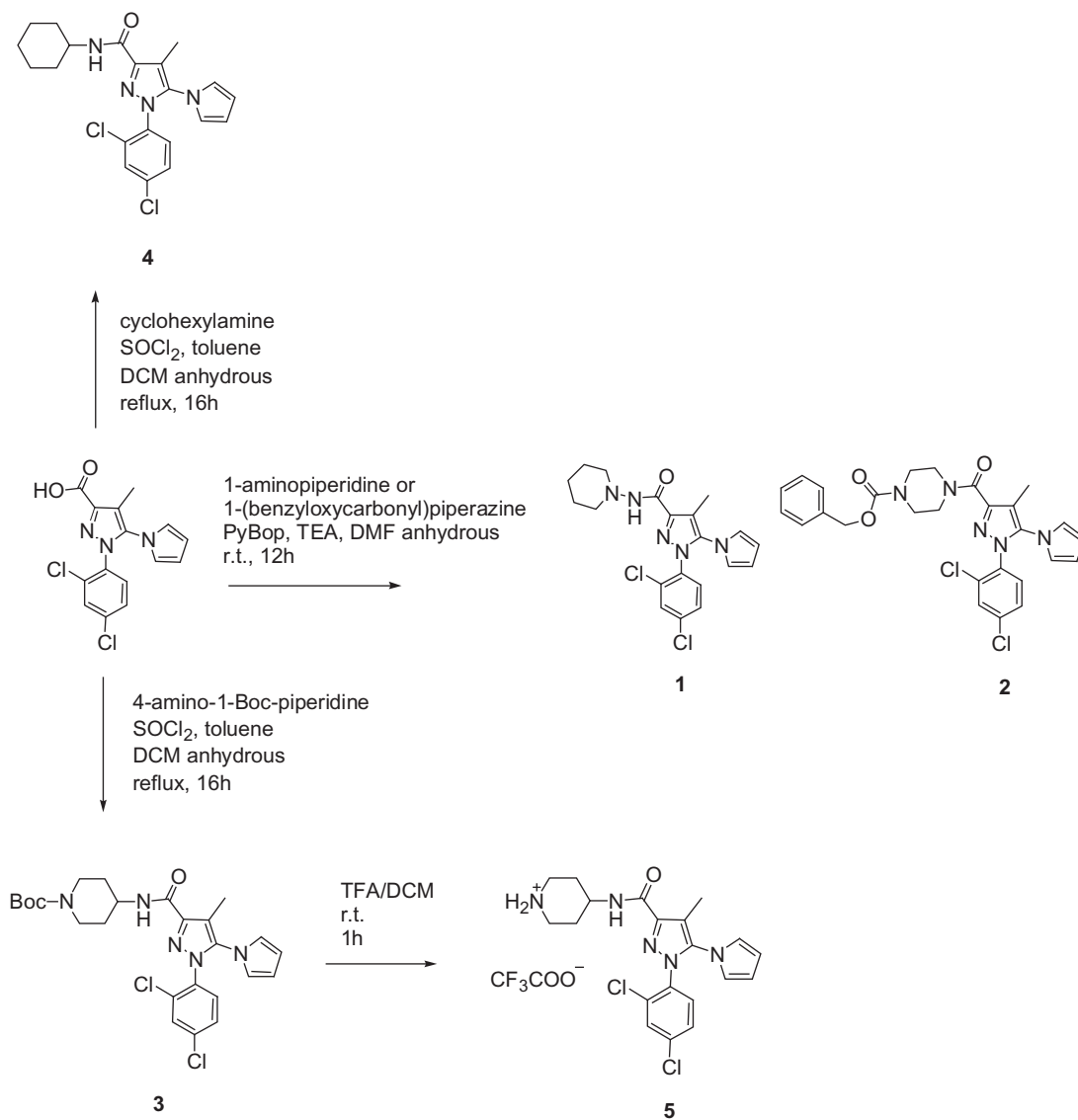
Results and discussion

A series of novel 1-aryl-5-(1H-pyrrol-1-yl)-1H-pyrazole-3-carboxamides Rimonabant analogs were tested for the stimulation of G

protein to evaluate their inverse agonist activity at CB1R to develop a bivalent compound (Scheme 1).

Compounds **1** and **2** have been previously synthesized and characterized by Silvestri et al.²⁶ for the development of potent CB1R inverse agonists. The molecules were designed by pursuing a bioisosteric approach on Rimonabant, from which **5** resulted to be one of the most interesting candidate with the advantage to be easily derivatisable by coupling with an opioid peptide on the piperidine secondary nitrogen in place of the *tert*-butoxycarbonyl (Boc) group. Indeed *N*-Boc derivative **3** resulted to be active, showing that the substitution of a bulky moiety at the secondary nitrogen of the 4-aminopiperidine terminus was possible without any loss of activity. Firstly, we investigated the cannabinoid receptor (CBR) binding affinity of the Rimonabant analogs compared with Rimonabant in competition binding assays using the non-selective cannabinoid receptor radioligand [³H]WIN55212 (CB1R < CB2R) (Figure 2). In this test, the Rimonabant analogs **1-4** exhibited nanomolar K_i values: compounds **1** (K_i = 125.9 nM) and **3** (K_i = 192.9 nM) were the most potent derivatives as compared with Rimonabant (K_i = 25 nM) (Table S1, see SI)^{27,28}.

In the following step, we investigated the effect of Rimonabant and its analogs on G-protein activity. In these experiments, we



Scheme 1. Synthesized Rimonabant analogs.

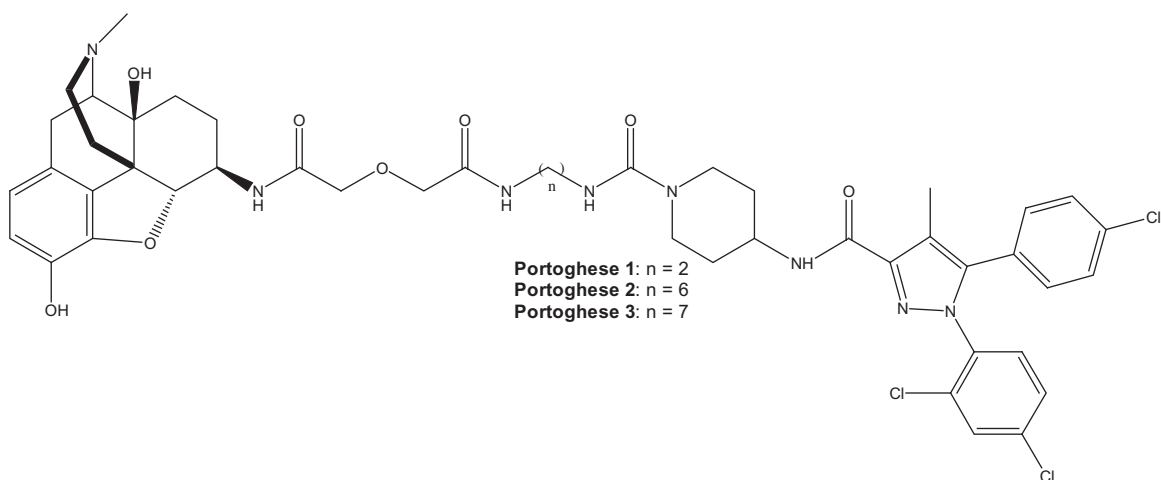


Figure 1. Structure of bivalent compounds previously designed by Le Naour et al.⁶

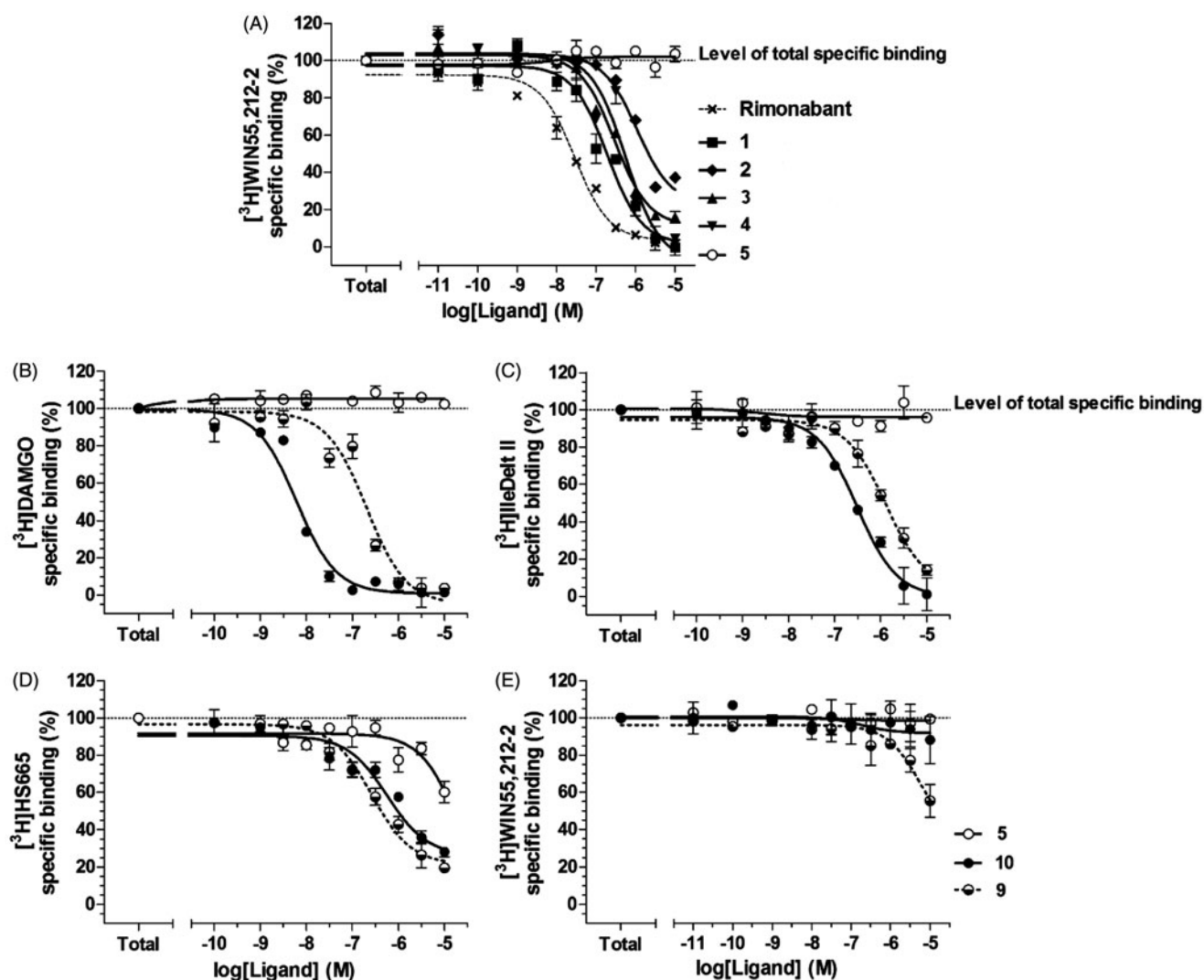


Figure 2. The binding affinity of Rimonabant and its analogs on CBR (A) and the MOR (B), DOR (C), KOR (D) and CBR binding affinity of 5, 9 and 10 (Tyr-D-Ala-Gly-Phe-NH₂) (E) in competition binding experiments. Figures represent the specific binding of [³H]WIN55 212-2, [³H]DAMGO, [³H]IleDelt II and [³H]HS665 in percentage in the presence of increasing concentrations (10⁻¹¹–10⁻⁵ M) of the indicated unlabeled ligands performed in rat (A, B and E) or in guinea pig (D) whole brain membrane homogenates. “Total” on the x-axis indicates the total specific binding of radioligand, which is measured in the absence of the unlabeled compounds. The level of total specific binding was defined as 100% and is presented with a dotted line. Points represent means ± SEM for at least three experiments performed in duplicates.

determined their agonist, antagonist or inverse agonist nature. The experiments were performed by [³⁵S]GTP γ S binding assays to control the GDP to GTP exchange of the α -subunit of G_i-protein. The analogs **1**, **2**, **3** and **4** decreased [³⁵S]GTP γ S specific binding

Table 1. The maximal G-protein efficacy (E_{\max}) and ligand potency ($\log EC_{50}$) of the Rimonabant and its analogs **1**, **2**, **3**, **4**, **5**, bivalent compound **9** and opioid peptide **10** in [³⁵S]GTP γ S binding assays on rat brain membrane homogenates. The values were calculated according to dose-response curves in Figure S1 (see SI) as described in the "Data analysis" section.

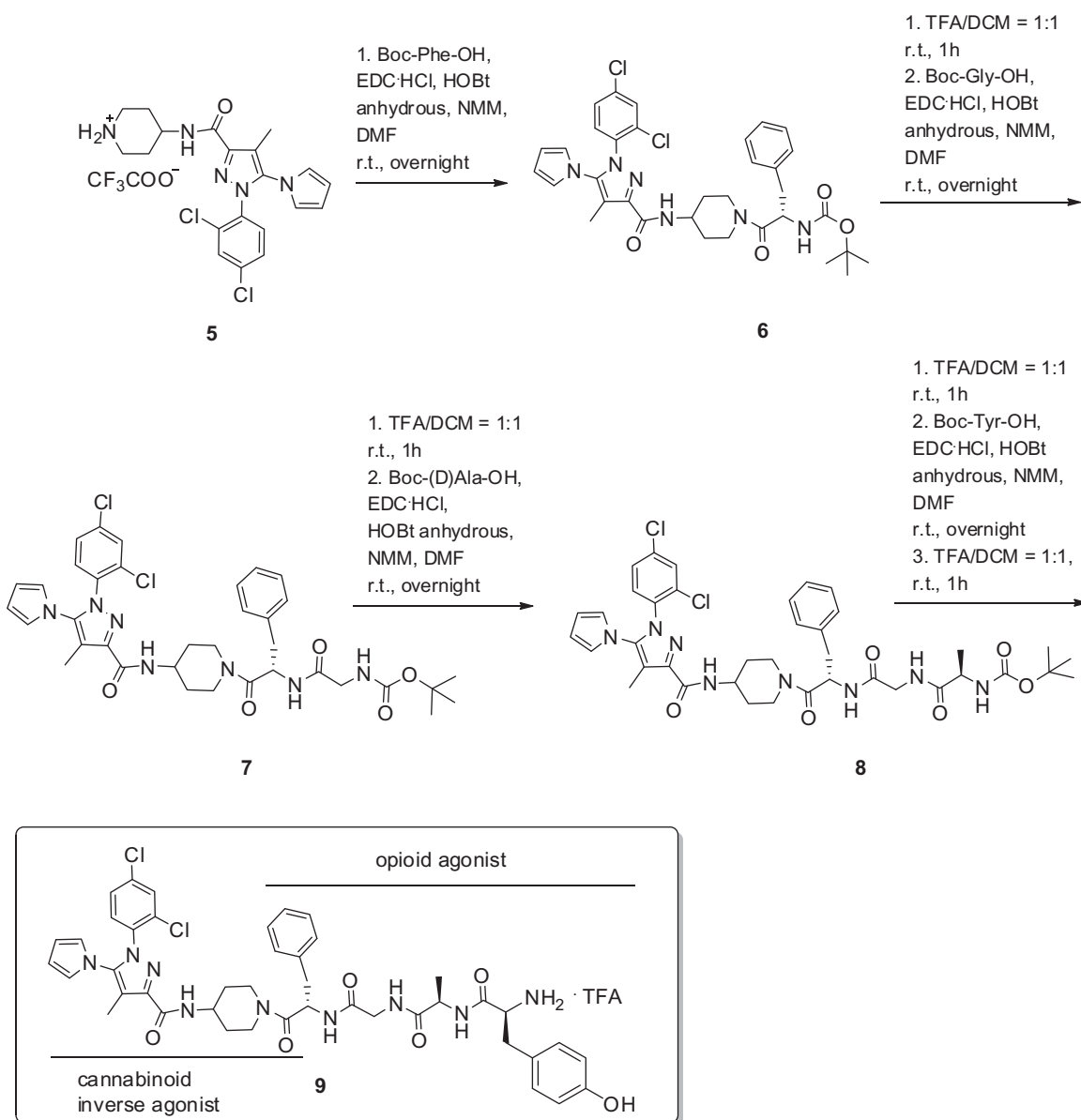
	$E_{\max} \pm$ S.E.M. (%)	$\log EC_{50} \pm$ S.E.M. (EC_{50})
Rimonabant	17.5 \pm 5.72	-5.65 \pm 0.12 (2.2 μ M)
1	26 \pm 5.27	-5.78 \pm 0.11 (1.62 μ M)
2	22.95 \pm 4.3	-5.81 \pm 0.09 (1.54 μ M)
3	12.78 \pm 8.48	-5.57 \pm 0.15 (2.69 μ M)
4	55.95 \pm 8.44	-5.72 \pm 0.3 (1.87 μ M)
5	99.2 \pm 46.1	ambiguous*
Bivalent compound (9)	36.62 \pm 10.35	-5.44 \pm 0.21 (3.6 μ M)
Tyr-D-Ala-Gly-Phe-NH ₂ (10)	161.5 \pm 4.08	-7.09 \pm 0.22 (81.3 nM)

*Since the compound did not alter significantly the total specific binding of the radioligand, thus $\log EC_{50}$ values cannot be interpreted.

(thus G-protein activity) compared with basal activity by more than 80% (E_{\max} = 17.5). Compounds **1-4** showed strong reduction of G-protein activity comparable to Rimonabant²⁹⁻³¹ and therefore displayed inverse agonist effects (Table 1). We examined the opioid receptor binding affinity of **5** in competition binding experiments, using opioid receptor selective radioligands. As expected, **5** did not bind to MOR and DOR and the affinity for the KOR was negligible (Table 1S, Figure 2(B-D))³¹⁻³⁷.

Opioid fragment **10** displayed a high affinity for MOR and DOR (MOR > DOR), while the compound had modest affinity for KOR, even at high nanomolar concentration (Table S1). As expected, the opioid peptide fragment did not show any affinity for CBR1 (Table 1S, Figure 2(E)); it increased G-protein activity with a maximum efficacy (E_{\max}) of nearly 70% above basal activity with EC_{50} of 81.3 nM, thus we can assert that this compound behaved as an agonist.

In the second part of our work, the designed bivalent compound **9** was synthesized (Scheme 2) and fully characterized (see SI). Peptide **10** has been prepared following the standard synthetic procedure for coupling reactions¹⁹.



Scheme 2. Synthesis of bivalent compound **9**.

Compound **9** behaved as inverse agonist as it decreased [35S]GTP γ S specific binding, thus reducing G-protein activity compared with the basal level (Table 1). Attaching the opioid fragment to Rimonabant analog **5** to give the bivalent compound **9** resulted in a rather selective DOR ligand with an unexpected improvement of affinity for KOR, and a loss in affinity for MOR and DOR, as compared with the opioid fragment alone (Table S1, Figure 2(B-D)). According to the affinities displayed by the opioid peptide **10** and **5**, the bivalent compound **9** showed modest affinity towards CB1R (Table S1, Figure 2(E)).

The results obtained in the hot plate and tail flick tests after i.c.v. injection in mice, are reported in Figure 3. In these experiments, compound **9** was administered i.c.v. at doses of 1, 5 and 10 μ g/mouse. Neither in the hot plate nor in the tail flick test treatments modified the behavioral response to thermal nociceptive stimuli. In the hot plate test, two-way ANOVA revealed no difference in treatments [$F_{3,120}=2.65$, $p=0.522$] and in time [$F_{4,120}=2.30$, $p=0.0629$] paradigms. Similar absence of effect was observed in the tail flick test, since treatment [$F_{3,120}=2.6$, $p=0.0551$] did not affect the latency of the nociceptive response all over the time [$F_{4,120}=1.32$, $p=0.2649$].

To further explore the pharmacologic profile of the bivalent ligand, hypothalamic perfusion test was performed with the aim to compare the release of hypothalamic neurotransmitters, after administration of the two separate pharmacophores and compound **9** in the preparation of synaptosomes. In this experiment, the three combinations produced different effects, showing a possible influence of the contemporary stimulation of CBRs and opioid systems on neurotransmitters release. As regards to **10**, we

found a significant stimulatory effect on norepinephrine (NE) and an inhibitory effect on serotonin (5-HT) and dopamine (DA) release, from hypothalamic synaptosomes (Figures 4 and 5).

Administration of compound **5** decreased only the DA release, whereas the treatment with **9** resulted in the reduction of the NE release's stimulation and DA inhibition (in less extent), with no significant effects on the 5-HT release. This is consistent with the inverse agonist activity on the G-protein system of compound **9**. The significant stimulatory effect on NE release could indicate a possible involvement in the regulation of energy balance. By contrast, the reduced inhibitory effect on hypothalamic DA release,

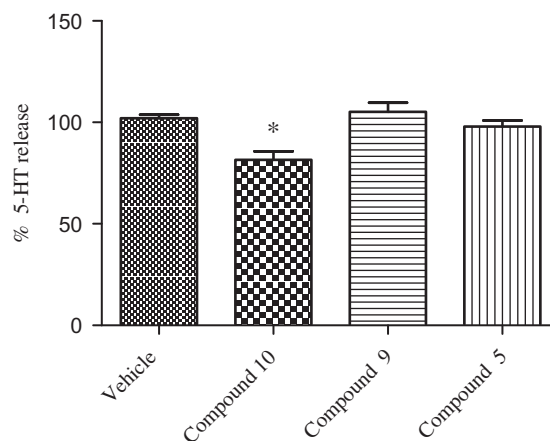


Figure 5. Effect of opioid peptide **10**, bivalent compound **9** and **5** on 5-HT release from hypothalamic synaptosomes, *in vitro*. ANOVA $p < 0.05$, * $p < 0.05$ vs. vehicle.

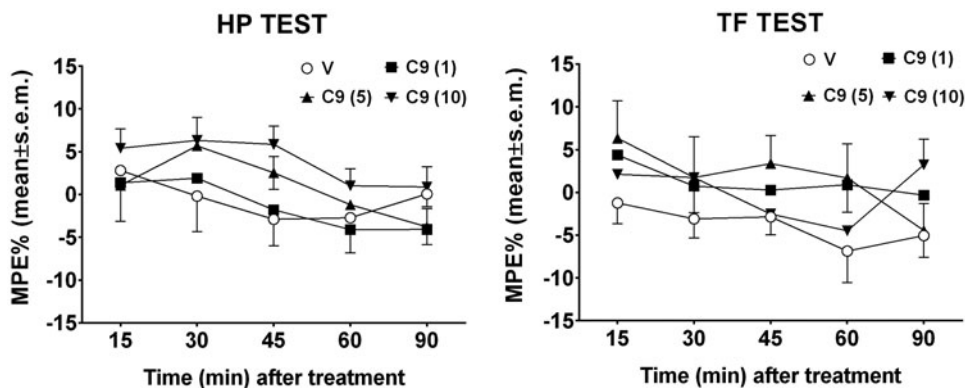


Figure 3. Hot-plate and Tail flick test. In these experiments, compound **9** (C9) was administered i.c.v. at doses of 1, 5 and 10 μ g/mouse. V is for vehicle. $N = 10$.

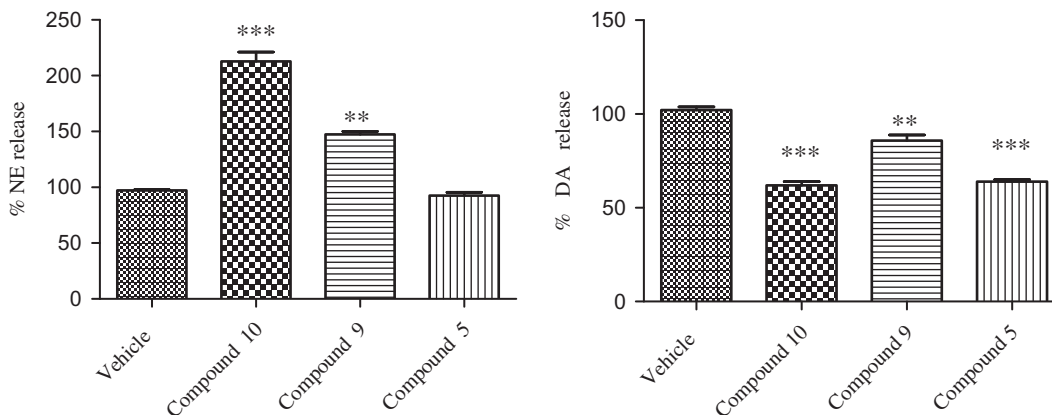


Figure 4. Effect of opioid peptide **10**, bivalent compound **9** and **5** on NE and DA release from hypothalamic synaptosomes, *in vitro*. ANOVA $p < 0.0001$, *** $p < 0.001$, ** $p < 0.01$ vs. vehicle.

Table 2. AdmetSAR regression derived data for diverse chemicals associated with known ADMET profiles for compounds **5**, **9** and **10**.

	Model	Unit	5	9	10
Absorption	Aqueous solubility	LogS	-3.0087	-3.1063	-3.3561
	Caco-2 permeability	LogP _{app} , cm/s	0.4939	0.1530	-0.2667
Toxicity	Rat acute toxicity	LD ₅₀ , mol/kg	2.6224	2.6993	2.2387
	Fish toxicity	pLC ₅₀ , mg/L	1.6053	1.3864	1.6515
	Tetrahymena pyriformis toxicity	pIGC ₅₀ , µg/L	0.5156	0.4875	-0.1386

compared with both parent molecules, is coherent with a minor potential of behavioral adverse effects *in vivo*^{38,39}.

The modulatory effects on hypothalamic biogenic amines suggest a possible involvement of both **10** and **5** in the interconnected neuronal pathways for the energy balance control. 5-HT release plays an anorectic role in the hypothalamus⁴⁰; DA injection in the perifornical hypothalamus inhibited food intake⁴¹, while DA administration in the lateral hypothalamus stimulated feeding⁴². NE could also exert both anorexigenic and orexigenic effects, related to the activation of α 1- and α 2-adrenoceptors, respectively⁴³. In addition, NE binding to hypothalamic α 2-adrenoceptors could increase energy expenditure, through the stimulated sympathetic activity⁴⁴. Previously, we observed that endomorphin-2 (EM-2), a selective MOR agonist, was able to stimulate food intake⁴⁵. The orexigenic effect was, albeit partially, related to the stimulated hypothalamic DA and NE activity which is also consistent with the increased oxygen consumption induced by EM-2 administration to mice⁴⁶.

The Absorption, Distribution, Metabolism, Excretion and Toxicity (ADMET) properties of compounds **5**, **9** and **10** were assessed by means of web servers and specialized programs. Several tools are available to profile compounds ADMET properties using *in silico* calculations. In Tables 2S–4S are reported the admetSAR⁴⁷, Molinspiration and cbligand.org ADMET calculated properties. In general, compounds **5**, **9** and **10** are not substrate for cytochromes and present low toxicity profiles (Table 2).

Regarding adsorption, derivative **9** violates three Lipinski's rules (high MW, number of HB donators and acceptors) that could prevent its oral bioavailability. AdmetSAR BBB and CACO predicted permeability indicate compound **5** as likely permeable, while compounds **9** and **10** are predicted at low probability to permeate BBB and CACO cells. These values are somehow in agreement with those reported in Table 4S where eight QSAR models indicate **5** as fully able to penetrate BBB, while **9** and **10** are predicted on the edge among positive and negative BBB penetrating molecules, as evinced from plots and threshold values reported in Table 5S.

To further inspect on possible BBB permeability, new models were herein developed by means of the python programming language, open-source cheminformatics library rdkit (www.rdkit.org), mathematical and scientific libraries numpy and scipy, machine-learning library scikit-learn⁴⁸. Application of the two new models confirmed that compound **5** is able to penetrate BBB, while compounds **9** and **10**, although predicted not able to penetrate BBB, show some probability to cross the BBB. The graphical analyses of similarity maps (Figure 2S) indicate the molecular portions likely responsible for the positive/negative BBB penetration⁴⁹.

According to our experiments, we can conclude that the receptor binding profile and biological activity of the bivalent compound **9** significantly changed compared with the individual components. The bivalent compound **9** showed higher selectivity for MOR than the enkephalin-like opioid peptide fragment **10**. More interestingly, it showed an improved KOR affinity compared with **5** and **10**. The stimulation of the G-protein system could be

explained by the inverse agonist effect of the bivalent compound. The lack of antinociceptive effect deals with the biological profile of an inverse agonist triggering the G-protein cascade. Also the ADMET properties of **9** are critical and establish benchmarks for further development of this class of bivalent compounds. Further studies on an alternative design of novel bivalent compounds based on an opioid peptide and a Rimobant analog are currently undergoing in our laboratory.

Disclosure statement

The authors report no conflicts of interest. The authors alone are responsible for the content and writing of this article.

Funding

The binding experiments were supported by National Research Development and Innovation Office (NKFIH) [grant number OTKA 108518] and by the ÚNKP-ÚNKP-16-3 New National Excellence Program of the Ministry of human capacities (Hungary).

References

- Zador F, Wollemann M. Receptome: interactions between three pain related receptors or the Triumvirate of cannabinoid, opioid and TRPV1 receptors. *Pharmacol Res* 2015;102:254–63.
- Shapira M, Vogel Z, Sarne Y. Opioid and cannabinoid receptors share a common pool of GTP-binding proteins in cotransfected cells, but not in cells which endogenously coexpress the receptors. *Cell Mol Neurobiol* 2000;20:291–304.
- Berrendero F, Mendizabal V, Murtra P, et al. Cannabinoid receptor and WIN 55 212-2-stimulated [35S]-GTPgammaS binding in the brain of mu-, delta- and kappa-opioid receptor knockout mice. *Eur J Neurosci* 2003;18:2197–202.
- Uriguen L, Berrendero F, Ledent C, et al. Kappa- and delta-opioid receptor functional activities are increased in the caudate putamen of cannabinoid CB1 receptor knockout mice. *Eur J Neurosci* 2005;22:2106–10.
- Jagerovic N, Fernandez-Fernandez C, Erdozain AM, et al. Combining rimonabant and fentanyl in a single entity: preparation and pharmacological results. *Drug Des Dev Ther* 2014;8:263–77.
- Le Naour M, Akgun E, Yekkirala A, et al. Bivalent ligands that target μ -opioid (MOP) and cannabinoid1 (CB1) receptors are potent analgesics devoid of tolerance. *J Med Chem* 2013;56:5505–13.
- Poras H, Bonnard E, Dange E, et al. New orally active dual enkephalinase inhibitors (DENKIs) for central and peripheral pain treatment. *J Med Chem* 2014;57:5748–63.
- Cota D, Tschop MH, Horvath TL, Levine AS. Cannabinoids, opioids and eating behavior: the molecular face of hedonism?. *Brain Res Rev* 2006;51:85–107.
- Desroches J, Beaulieu P. Opioids and cannabinoids interactions: involvement in pain management. *Curr Drug Targets* 2010;11:462–73.
- Bushlin I, Rozenfeld R, Devi LA. Cannabinoid-opioid interactions during neuropathic pain and analgesia. *Curr Opin Pharmacol* 2010;10:80–6.
- da Fonseca Pacheco D, Klein A, de Castro Perez A, et al. The μ -opioid receptor agonist morphine, but not agonists at δ -

- or k-opioid receptors, induces peripheral antinociception mediated by cannabinoid receptors. *Br J Pharmacol* 2008;154:1143–9.
12. Manzanares J, Corchero J, Romero J, et al. Pharmacological and biochemical interactions between opioids and cannabinoids. *Trends Pharmacol Sci* 1999;20:287–94.
 13. Wollemann M, Toth F, Benyhe S. Protein kinase C inhibitor BIM suspended TRPV1 effect on mu-opioid receptor. *Brain Res Bull* 2013;90:114–17.
 14. Parolaro D, Rubino T, Vigano D, et al. Cellular mechanisms underlying the interaction between cannabinoid and opioid system. *Curr Drug Targets* 2010;11:393–405.
 15. Manzanares J, Ortiz S, Oliva JM, et al. Interactions between cannabinoid and opioid receptor systems in the mediation of ethanol effects. *Alcohol Alcohol* 2005;40:25–34.
 16. Scavone JL, Sterling RC, Van Bockstaele EJ. Cannabinoid and opioid interactions: implications for opiate dependence and withdrawal. *Neuroscience* 2013;248:637–54.
 17. Befort K. Interactions of the opioid and cannabinoid systems in reward: insights from knockout studies. *Front Pharmacol* 2015;56:6.
 18. Moreira FA, Aguiar DC, Campos AC, et al. Antiaversive effects of cannabinoids: is the periaqueductal gray involved? *Neural Plast* 2009;2009:625469.
 19. Mollica A, Costante R, Novellino E, et al. Design, synthesis and biological evaluation of two opioid agonist and Cav 2.2 blocker multitarget ligands. *Chem Biol Drug Des* 2015;86:156–62.
 20. Monti L, Stefanucci A, Pieretti S, et al. Evaluation of the analgesic effect of 4-anilidopiperidine scaffold containing ureas and carbamates. *J Enzyme Inhib Med Chem* 2016;31:1638–47.
 21. Dvoracko S, Stefanucci A, Novellino E, Mollica A. The design of multitarget ligands for chronic and neuropathic pain. *Future Med Chem* 2015;7:2469–83.
 22. Podolsky AT, Sandweiss A, Hu J, et al. Novel fentanyl-based dual μ/δ -opioid agonists for the treatment of acute and chronic pain. *Life Sci* 2013;93:1010–16.
 23. Deekonda S, Wugalter L, Rankin D, et al. Design and synthesis of novel bivalent ligands (MOR and DOR) by conjugation of enkephalin analogues with 4-anilidopiperidine derivatives. *Bioorg Med Chem Lett* 2015;25:4683–8.
 24. Caille S, Parsons LH. Cannabinoid modulation of opiate reinforcement through the ventral striatopallidal pathway. *Neuropsychopharmacology* 2006;31:804–13.
 25. Azizi P, Haghparast A, Hassanpour-Ezatti M. Effects of CB1 receptor antagonist within the nucleus accumbens on the acquisition and expression of morphine-induced conditioned place preference in morphine-sensitized rats. *Behav Brain Res* 2009;197:119–24.
 26. Silvestri R, Cascio MG, La Regina G, et al. Synthesis, cannabinoid receptor affinity, and molecular modeling studies of substituted 1-aryl-5-(1H-pyrrol-1-yl)-1H-pyrazole-3-carboxamides. *J Med Chem* 2008;51:1560–76.
 27. Rinaldi-Carmona M, Barth F, Héaulme M. Biochemical and pharmacological characterisation of SR141716A, the first potent and selective brain cannabinoid receptor antagonist. *Life Sci* 1995;56:1941–7.
 28. Rinaldi-Carmona M, Barth F, Héaulme M, et al. SR141716A, a potent and selective antagonist of the brain cannabinoid receptor. *FEBS Lett* 1994;350:240–4.
 29. MacLennan SJ, Reynen PH, Kwan J, Bonhaus DW. Evidence for inverse agonism of SR141716A at human recombinant cannabinoid CB1 and CB2 receptors. *Br J Pharmacol* 1998;124:619–22.
 30. Sim-Selley LJ, Brunk LK, Selley DE. Inhibitory effects of SR141716A on G-protein activation in rat brain. *Eur J Pharmacol* 2001;414:135–43.
 31. Cinar R, Szücs M. CB1 receptor-independent actions of SR141716 on G-protein signaling: coapplication with the μ -opioid agonist Tyr-D-Ala-Gly-(NMe)Phe-Gly-ol unmasks novel, pertussis toxin-insensitive opioid signaling in μ -opioid receptor-Chinese hamster ovary cells. *J Pharmacol Exp Ther* 2009;330:567–74.
 32. Fong TM, Shearman LP, Stribling DS, et al. Pharmacological efficacy and safety profile of taranabant in preclinical species. *Drug Dev Res* 2009;70:349–62.
 33. Kathmann M, Flau K, Redmer A, et al. Cannabidiol is an allosteric modulator at mu- and delta-opioid receptors. *Naunyn Schmiedebergs Arch Pharmacol* 2006;372:354–61.
 34. Seely KA, Brents LK, Franks LN, et al. AM-251 and rimonabant act as direct antagonists at μ -opioid receptors: implications for opioid/cannabinoid interaction studies. *Neuropharmacology* 2012;63:905–15.
 35. Zádor F, Kocsis D, Borsodi A, Benyhe S. Micromolar concentrations of rimonabant directly inhibits delta opioid receptor specific ligand binding and agonist-induced G-protein activity. *Neurochem Int* 2014;67:14–22.
 36. Zádor F, Lénárt N, Csibrány B, et al. Low dosage of rimonabant leads to anxiolytic-like behavior via inhibiting expression levels and G-protein activity of kappa opioid receptors in a cannabinoid receptor independent manner. *Neuropharmacology* 2015;89:298–307.
 37. Zádor F, Otvös F, Benyhe S, et al. Inhibition of forebrain μ -opioid receptor signaling by low concentrations of rimonabant does not require cannabinoid receptors and directly involves μ -opioid receptors. *Neurochem Int* 2012;61:378–88.
 38. Deats SP, Adidharma W, Yan L. Hypothalamic dopaminergic neurons in an animal model of seasonal affective disorder. *Neurosci Lett* 2015;602:17–21.
 39. Nathan PJ, O'Neill BV, Napolitano A, Bullmore ET. Neuropsychiatric adverse effects of centrally acting antiobesity drugs. *CNS Neurosci Ther* 2011;17:490–505.
 40. Leibowitz SF. Hypothalamic paraventricular nucleus: interaction between alpha 2-noradrenergic system and circulating hormones and nutrients in relation to energy balance. *Neurosci Biobehav Rev* 1988;12:101–9.
 41. Gillard ER, Dang DQ, Stanley BG. Evidence that neuropeptide Y and dopamine in the perifornical hypothalamus interact antagonistically in the control of food intake. *Brain Res* 1993;628:128–36.
 42. Yang ZJ, Meguid MM. LHA dopaminergic activity in obese and lean Zucker rats. *NeuroReport* 1995;6:1191–4.
 43. Wellman PJ, Davies BT, Morien A, McMahon L. Modulation of feeding by hypothalamic paraventricular nucleus α 1- and α 2-adrenergic receptors. *Life Sci* 1993;53:669–79.
 44. Levin BE. Reduced paraventricular nucleus norepinephrine responsiveness in obesity-prone rats. *Am J Physiol* 1996;270:456–61.
 45. Brunetti L, Ferrante C, Orlando G, et al. Orexigenic effects of endomorphin-2 (EM-2) related to decreased CRH gene expression and increased dopamine and norepinephrine activity in the hypothalamus. *Peptides* 2013;48:83–8.
 46. Asakawa A, Inui A, Ueno N, et al. Endomorphin-1, an endogenous mu-opioid receptor-selective agonist,

- stimulates oxygen consumption in mice. *Horm Metab Res* 2000;32:51–2.
47. Cheng F, Li W, Zhou Y, et al. AdmetSAR: a comprehensive source and free tool for assessment of chemical ADMET properties. *J Chem Inf Model* 2012;52:3099–105.
48. Duchesnay D. Scikit-learn: machine learning in Python. *J Mach Learn Res* 2011;12:2825–30.
49. Riniker S, Landrum GA. Similarity maps – a visualization strategy for molecular fingerprints and machine-learning methods. *J Cheminform* 2013;5:43.